

Review

Chemical and Pharmacological Research of the Plants in Genus *Euphorbia*

Qing-Wen Shi, Xiao-Hui Su, and Hiromasa Kiyota

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Chemical and Pharmacological Research of the Plants in Genus *Euphorbia*

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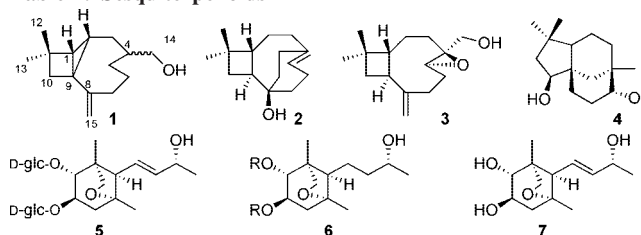
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Figure 1. Flower of *Euphorbia pekinensis* Rupr.

Table 1. Sesquiterpenoids



no.	name	plant	ref
1	euphinginol	<i>E. wangii</i>	10
2	cyclocaryophylla-4-en-8-ol	<i>E. wangii</i>	10
3	4β,5α-epoxy-4,5-dihydrocaryophyllen-14-ol	<i>E. wangii</i>	10
4	clovandiol	<i>E. wangii</i>	10
5	euphorbioside A	<i>E. resinifera</i>	11
6	euphorbioside B	<i>E. resinifera</i>	11
7	deglycosyl euphorbioside A	<i>E. resinifera</i>	11

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Xiao-Hui Su (b. 1983 in Qinhuangdao, Hebei Province). She got her Bachelor and Master degrees from School of Pharmaceutical Sciences, Hebei Medical University in 2005 and 2008. Her master thesis is studies on the antitumour components of *Euphorbia lathyris* seeds and flowers of *Inula japonica*.

1. Introduction

The genus *Euphorbia* is the largest in the spurge family, comprising more than 2000 species (Figure 1).¹ Some species of the genus *Euphorbia* have been used as medicinal plants for the treatment of skin diseases, gonorrhoea, migraine, and intestinal parasites and as wart cures.² The researched parts of the *Euphorbia* species include roots, seeds, latex, lactiferous tubes, stem wood, stem barks, leaves, and whole plants. Plants in the family Euphorbiaceae are well known for the chemical diversity of their isoprenoid constituents.³ Diterpenoids are the majority of the genus with many different core frameworks such as jatrophanes, lathyranes, tiglianens, ingenanes, myrsinols, etc. The triterpene alcohols found in the latex of *Euphorbia* species have been used as chemotaxonomic markers.^{4,5} In addition, sesquiterpenoids, phloracetophenones, cerebrosides, glycerols, flavonoids, and steroids were also obtained. The compounds isolated from genus *Euphorbia* and extracts perform many different activities,



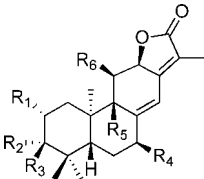
Hiromasa Kiyota (b. 1966 in Sendai, Japan), Associate Professor of Bioorganic Chemistry at Tohoku University, Japan. He received his B. S. degree (1989, soil science, Prof. Hidenori Wada) and M. S. degree (1991, organic chemistry, Prof. Kenji Mori and Associate Prof. Takeshi Kitahara) from the University of Tokyo. In 1991 he assumed the position of Assistant Professor at Prof. K. Mori's Laboratory, moved to Prof. Takayuki Oritani's Laboratory in 1994, and he was promoted to Associate Professor (Prof. Shigefumi Kuwahara) in 2002. In 1995 he received his PhD from the University of Tokyo (Prof. Kenji Mori and Prof. Takeshi Kitahara) about the synthesis of optically active insect pheromones. He joined the research group of Prof. Steven V. Ley at Cambridge University, U.K. as Visiting Academic (2001–2002). In 2003 he received The Japan Bioscience, Biotechnology and Agrochemistry Society Award for the Encouragement of Young Scientists. His research interests extend over a wide range of natural product chemistry, especially on the synthesis of biologically active compounds such as antibiotics, phytotoxins, plant hormones, insect pheromones, marine products, perfumery, etc.

including antiproliferation, modulability of multidrug resistance, cytotoxic activity, antimicrobial and antiinflammatory activity, etc. Biological activities including skin irritant, tumor promotion, and proinflammatory properties are attributed to the presence of specific classes of macro- and polycyclic diterpenes.^{6–9}

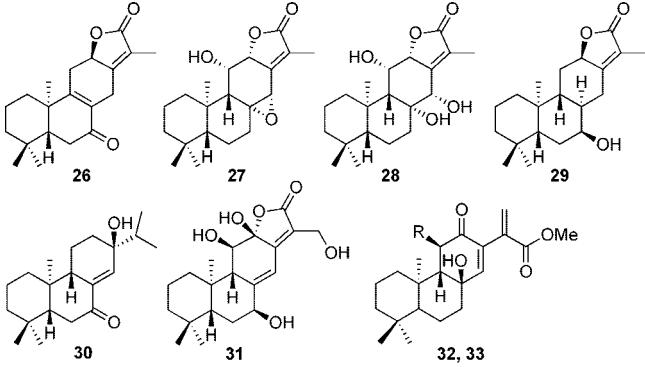
In this review article, we will summarize the phytochemical progress and list all of the compounds isolated from the

Table 2. *ent*-Abietanes-1

no.	name	R	plant	ref
8	17-acetoxyjolkinolide A	OAc	<i>E. fischeriana</i>	13
9	jolkinolide A	H	<i>E. fischeriana</i>	14
			<i>E. sessiliflora</i>	15
			<i>E. portulacoides</i>	16
10	17-hydroxyjolkinolide A	OH	<i>E. fischeriana</i>	13
			<i>E. acaulis</i>	17
			<i>E. caudicifolia</i>	18
			<i>E. portulacoides</i>	16
			<i>E. sessiliflora</i>	15
11	jolkinolide B	H	<i>E. fischeriana</i>	14
			<i>E. sessiliflora</i>	15
			<i>E. segueriana</i>	19
			<i>E. ebracteolata</i>	20
			<i>E. portulacoides</i>	16
12	17-hydroxyjolkinolide B	OH	<i>E. fischeriana</i>	14
13	17-acetoxyjolkinolide B	OAc	<i>E. fischeriana</i>	21
14	8 α ,14-dihydro-7-oxojolkinolide E	H	<i>E. characias</i>	22
15	8 α ,14-dihydro-7-oxohelioscopinolide A	OH	<i>E. semiperfoliata</i>	23
			<i>E. characias</i>	22

Table 3. *ent*-Abietanes-2


no.	name	R ₁	R ₂	R ₃	R ₄	R ₅	R ₆	plant	ref
16	7β-hydroxy- <i>ent</i> -abieta-8(14),13(15)-dien-12α,16-olide	H	H	H	OH	H	H	<i>E. seguieriana</i>	19
17	7β,9β-dihydroxy- <i>ent</i> -abieta-8(14),13(15)-dien-12α,16-olide	H	H	H	OH	OH	H	<i>E. seguieriana</i>	19
18	- <i>ent</i> -11β-hydroxyabieta-8(14),13(15)-dien-12β,16-olide	H	H	H	H	H	OH	<i>E. fischeriana</i>	13
19	jolkinolide E	H	H	H	H	H	H	<i>E. helioscopia</i>	25
20	helioscopinolide A	H	OH	H	H	H	H	<i>E. characias</i>	22
								<i>E. helioscopia</i>	25
								<i>E. pubescens</i>	26
21	helioscopinolide B	H	H	OH	H	H	H	<i>E. semiperfoliata</i>	23
								<i>E. characias</i>	22
								<i>E. helioscopia</i>	25
								<i>E. pubescens</i>	26
22	2α-hydroxyhelioscopinolide B	OH	OH	H	H	H	H	<i>E. helioscopia</i>	12
23	helioscopinolide C	=O	OH	H	H	H	H	<i>E. helioscopia</i>	25
24	helioscopinolide D	H	=O	=O	H	OH	H	<i>E. calyptrate</i>	27
25	helioscopinolide E	H	=O	=O	H	H	H	<i>E. calyptrate</i>	27

Table 4. *ent*-Abietanes-3


no.	name	R	plant	ref
26	7-oxo- <i>ent</i> -abieta-8,13(15)-dien-12α,16-olide	–	<i>E. seguieriana</i>	19
27	ebracteolatanolide A	–	<i>E. ebracteolata</i>	20
28	ebracteolatanolide B	–	<i>E. ebracteolata</i>	20
29	8α,14-dihydro-7β-hydroxyjolkinolide E	–	<i>E. terracina</i>	28
30	13β-hydroxy- <i>ent</i> -abiet-8(14)-en-7-one	–	<i>E. fischeriana</i>	21
31	languin B: 7β,11β,12β,17-tetrahydroxy- <i>ent</i> -abieta-8(14),13(15)-dien-16,12α-olide	–	<i>E. fischeriana</i>	13
32	methyl 8β-hydroxy-12-oxo- <i>ent</i> -abieta-13,15(17)-dien-16-oate	H	<i>E. portulacoides</i>	16
33	methyl 8β,11β-dihydroxy-12-oxo- <i>ent</i> -abieta-13,15(17)-dien-16-oate	OH	<i>E. portulacoides</i>	16

genus *Euphorbia* over the past few decades. Also included are the biological activities of compounds isolated in recent years and parts structure–activity relationships.

2. Chemical Constituents

2.1. Sesquiterpenoids (Table 1)

In 1997, Shi et al. reported the isolation of a new (**1**) and three known (**2–4**) sesquiterpenoids from *E. wangii*.¹⁰ This is the first investigation on sesquiterpenoids from the genus *Euphorbia*. In 2002, Fattorusso et al. isolated two novel bisnorsesquiterpene glycosides, euphorbiosides A (**5**) and B (**6**), as well as the aglycone of **5** (compound **7**) from more polar fractions of *E. resinifera*.¹¹ In 2006, Zhang et al. isolated two known nor-sesquiterpenoids, 4,5-dihy-

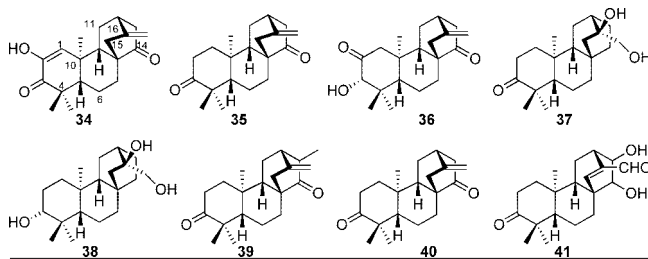
droblumenol A and aglycone of icaraside B2, from *E. helioscopia*.¹²

2.2. Higher Diterpenoids

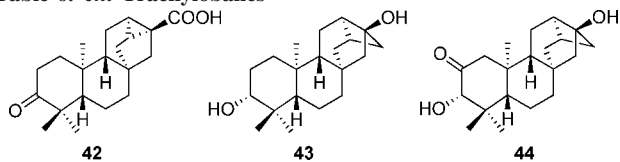
2.2.1. *ent*-Abietanes, *ent*-Atisanes, *ent*-Kauranes, *ent*-Isopimaranes, and *ent*-Pimaranes (Tables 1–9)

Polycyclic diterpenoids with a common 6/6/6-tricyclic ring are also major constituents of *Euphorbia*. Higher diterpenoids originated from geranylgeranyl diphosphate by “concertina-like” cyclization, *ent*-abietanes, *ent*-atisanes, *ent*-kauranes, *ent*-isopimaranes, and *ent*-pimaranes, are introduced as follows.

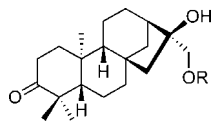
The *ent*-abietanes from this species usually contain fourth α,β-unsaturated γ-lactone ring. 2α-Hydroxyhelioscopinolide

Table 5. *ent*-Atisanes

no.	name	plant	ref
34	<i>ent</i> -2-hydroxyatisa-1,16(17)-diene-3,14-dione	<i>E. characias</i>	22
35	<i>ent</i> -atis-16(17)-ene-3,14-dione	<i>E. characias</i> <i>E. fidjiana</i>	22 29
36	<i>ent</i> -3 α -hydroxyatis-16(17)-ene-2,14-dione	<i>E. characias</i>	22
37	<i>ent</i> -16 α ,17-dihydroxyatisan-3-one	<i>E. quinquecostata</i> <i>E. fidjiana</i>	30 29
38	<i>ent</i> -atisane-3 β ,16 α ,17-triol	<i>E. quinquecostata</i> <i>E. fidjiana</i>	30 31
39	<i>ent</i> -(13 <i>R</i>)-hydroxyatis-16(17)-ene-3,4-dione	<i>E. fidjiana</i>	32
40	<i>ent</i> -atis-16(17)-ene-3,4-dione	<i>E. fidjiana</i>	32
41	<i>ent</i> -(13 <i>R</i> ,14 <i>R</i>)-dihydroxyatis-15-ene-15,17-dione	<i>E. fidjiana</i>	32

Table 6. *ent*-Trachylobanes

no.	name	plant	ref
42	<i>ent</i> -3-oxo-trachyloban-17-oic acid	<i>E. wallichii</i>	33
43	wallichanol A	<i>E. wallichii</i>	33
44	wallichanol B	<i>E. wallichii</i>	33

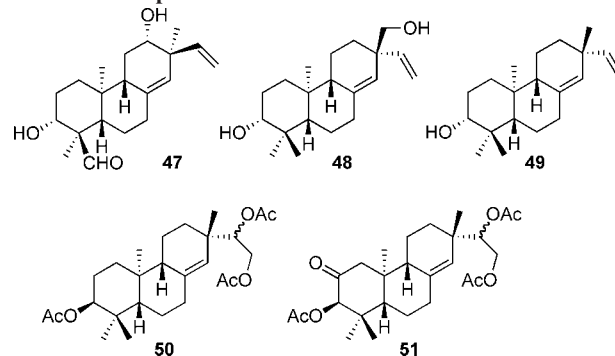
Table 7. *ent*-Kauranes

no.	name	R	plant	ref
45	16 β ,17-dihydroxy- <i>ent</i> -kauran-3-one	H	<i>E. characias</i>	22
			<i>E. portulacoides</i>	16
			<i>E. sieboldiana</i>	34
			<i>E. wallichii</i>	35
46	17-acetoxy-16 β -hydroxy- <i>ent</i> -kauran-3-one	Ac	<i>E. portulacoides</i>	16

B (22), which was reported as an intermediate in the course of structure elucidation of helioscopinolide C (23), was isolated from natural sources for the first time in 2006 by Zhang et al.¹² The occurrence of *ent*-atisanes in plants from the genus *Euphorbia* is very rare and, apart from *E. fidjiana*, has so far been documented only in the roots of the Indian species *E. acaulis* Roxb.³⁷

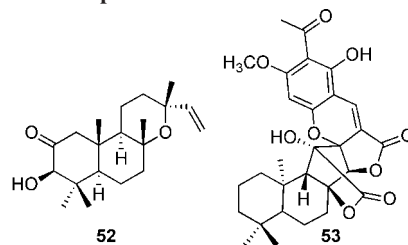
2.2.2. Other Diterpenoids

In 1997, a manoyloxide derivative, 3 β -hydroxy-2-oxomanoyloxide (52) was isolated from *E. segetalis* by Jakupovic et al.³⁸ Langduin C (53), a novel dimeric diterpenoid, was isolated from the roots of *E. fischeriana*.⁴⁰

Table 8. *ent*-Isopimaranes and *ent*-Pimaranes

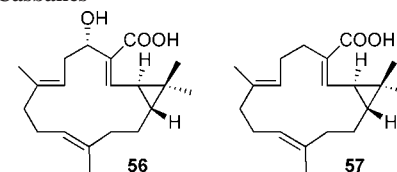
no.	name	plant	ref
47	3 α ,12 α -dihydroxy- <i>ent</i> -8(14),15-isopimaradien-18-al	<i>E. quinquecostata</i>	36
48	<i>ent</i> -pimara-8(14),15-diene-3 α ,17-diol	<i>E. fischeriana</i>	21
49	<i>ent</i> -pimara-8(14),15-dien-3 α -ol	<i>E. characias</i>	22
50	3 β ,15,16-triacetoxy- <i>ent</i> -pimar-8(14)-ene	<i>E. characias</i>	22
51	3 β ,15,16-triacetoxy- <i>ent</i> -pimar-8(14)-en-2-one	<i>E. characias</i>	22

Table 9. Other Diterpenoids



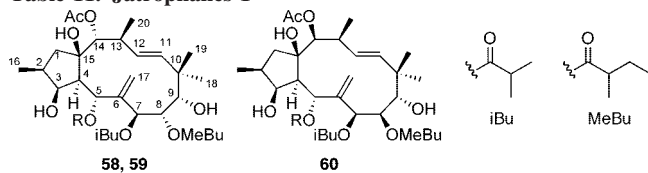
no.	name	plant	ref
52	3 β -hydroxy-2-oxomanoyloxide	<i>E. segetalis</i>	38
53	langduin D	<i>E. fischeriana</i>	39
54	langduin C	<i>E. fischeriana</i>	40
55	fischeria A	<i>E. fischeriana</i>	41

Table 10. Casbanes

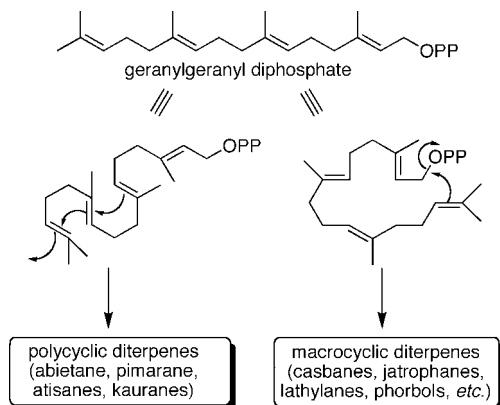
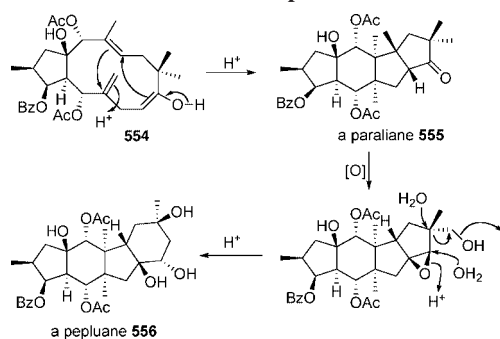


no.	name	plant	ref
56	yuexiandajisu A	<i>E. ebracteolata</i>	42
57	yuexiandajisu B	<i>E. ebracteolata</i>	42

penoid, was isolated from the roots of *E. fischeriana*.⁴⁰ In 1999, fischeria A (55), a novel norditerpene lactone from *E. fischeriana*, was isolated from the rhizomes of *E. fischeriana* Steud.⁴¹

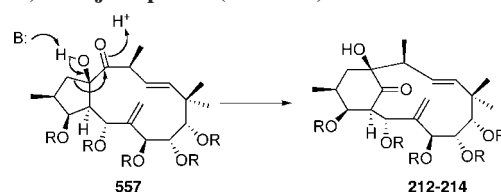
Table 11. Jatrophanes-1


no.	name	R	plant	ref
58	(2 <i>S</i> ,3 <i>S</i> ,4 <i>S</i> ,5 <i>S</i> ,7 <i>R</i> ,7 <i>S</i> ,8 <i>S</i> ,9 <i>S</i> ,13 <i>S</i> ,14 <i>R</i> ,15 <i>R</i>)-14-acetoxy-7-isobutyryloxy-5,8-bis(2-methylbutanoyloxy)jatropha-6(17),11 <i>E</i> -diene-3,9,15-triol	iBu	<i>E. terracina</i>	43
59	(2 <i>S</i> ,3 <i>S</i> ,4 <i>S</i> ,5 <i>S</i> ,7 <i>R</i> ,7 <i>S</i> ,8 <i>S</i> ,9 <i>S</i> ,13 <i>S</i> ,14 <i>R</i> ,15 <i>R</i>)-14-acetoxy-5,7-diisobutyryloxy-8-(2-methylbutanoyloxy)jatropha-6(17),11 <i>E</i> -diene-3,9,15-triol	MeBu	<i>E. terracina</i>	43
60	(2 <i>S</i> ,3 <i>S</i> ,4 <i>S</i> ,5 <i>S</i> ,7 <i>R</i> ,7 <i>S</i> ,8 <i>R</i> ,9 <i>S</i> ,13 <i>S</i> ,14 <i>S</i> ,15 <i>R</i>)-14-acetoxy-5,7-diisobutyryloxy-8-(2-methylbutanoyloxy)jatropha-6(17),11 <i>E</i> -diene-3,9,15-triol	—	<i>E. segetalis</i>	38

Scheme 1. Possible Biogenetic Route from Geranylgeranyl Diphosphate to Polycyclic and Macrocylic Diterpenoids**Scheme 2. Proposed Biogenesis of a Pepluane 556 Starting from a Suitable Substituted Jatropha 554**

2.3. Lower Diterpenoids

Considerable attention has been paid to the macrocyclic diterpenoids derived from cembrane cation because of their high chemical diversity and therapeutically relevant bioactivity. “Euphorbiaceae diterpenoids” include casbanes, jatrophanes, lathyrans, myrsinanes, tiglianes, ingenanes, segitanes, paralianes, pepluanes, and euphoractines as shown below.

Scheme 3. Pinacol-Type Rearrangement of a Jatropha to 1(15→14) Abeojatropha (212–214)

2.3.1. Casbanes (Table 10)

Two bicyclic diterpenoids with a casbane skeleton (**56** and **57**) were isolated from *E. ebracteolata*.⁴²

2.3.2. Jatrophanes (Tables 11–28)

Euphorbiaceae is a great rich source of jatropha and the related diterpenoids. Jatrophanes with various oxygenation stages and stereoisomers are listed in the tables. These compounds are usually substituted with various acyl groups, such as acetyl, propanoyl, butanoyl, isobutyryl, benzoyl, tigloyl, nicotinoyl, angeloyl, etc., and are sometimes called jatropha polyesters. Corea et al. carried out a molecular mechanic and dynamics calculation on amygdaloidins A–L (**120–131**).⁵⁶ The data obtained gave further information on the *endo*- and *exo*-type major conformations for the jatropha diterpenoids. On the other hand, the stereochemistry of hemiacetal type jatropha, esulatin C (**187**), at C-7, C-8, C-9, C-11, and C-13 could not be determined on the basis of NOESY correlations because of the high flexibility of this part of the molecule.⁵⁸ Compound **191** is the only jatropha with 12,15-epoxy ring.⁶⁸ 17-Bishomojatrophanes forming 6- or 8-membered lactone ring teracinolides (**192–210**) and salicinolide (**211**) were found.^{28,38,73–76} Salicifoline (**215**) is the first representative of a new type of tricyclic diterpenoids involving a novel 5/8/8 fused ring system, that is, 12,17-cyclojatropha.⁷⁶ Euphosalicin (**216**) also contains a new 9(10→18) abeojatropha skeleton.⁶³

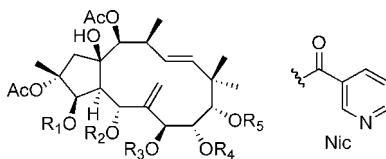
2.3.3. Lathyrans (Tables 29–33)

Lathyrans with a 5/11/3-membered ring are also very common in *Euphorbia* species. Several compounds are substituted with methoxy group (Tables 29 and 31). Phenylacetyl and methoxyphenylacetyl groups are rather diagnostic to lathyrans. Lathyranoic acid A (**260**) is the first seco-lathyrane diterpenoid in nature from *E. lathyris*.⁹¹ Lathyrone A (**261**) is a novel 1(15→14) abeolathyrane compound also isolated from *E. lathyris* (section 3).⁹¹

2.3.4. Myrsinanes, Cyclomyrsinanes, and Premyrcinanes (Tables 34–43)

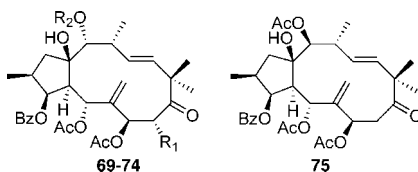
Myrsinanes and cyclomyrsinanes are derived from lathyrans via premyrsinanes (section 3). In addition to the normal myrsinanes (6,12-cyclojatrophanes) with a 5/7/5-ring carbon framework, compounds with a hemiacetal ring, a 13,17-epoxy ring, or a 10,13-epoxy ring are tabulated in Tables 34–39. The stereochemistry of all the frameworks and substituents are the same in myrsinanes. 9-Bishomomyrsinane **299** contains a δ -lactone ring.¹⁰⁷ The configuration of 12-position of eupobeotol (**306**) is reversed.¹¹¹ All the cyclomyrsinanes **300–305** have a 12,17-epoxy ring. Premyrsinanes **319–323** forms a rare acetyl hemiacetal moiety.

Table 12. Jatrophanes-2



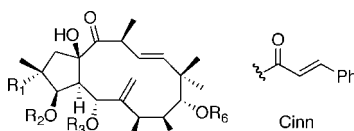
no.	name	R ₁	R ₂	R ₃	R ₄	R ₅	plant	ref
61	2,5,14-triacetoxy-3-benzoyloxy-7-isobutyryloxy-9-nicotinoyloxyjatropa-6(17),11 <i>E</i> -diene-8,15-diol	Bz	Ac	iBu	H	Nic	<i>E. peplus</i>	44
62	2,5,7,8,9,14-hexaacetoxy-3-benzoyloxyjatropa-6(17),11 <i>E</i> -dien-15-ol	Bz	Ac	Ac	Ac	Ac	<i>E. peplus</i>	44
63	2,5,9,14-tetraacetoxy-3-benzoyloxy-7-isobutyryloxyjatropa-6(17),11 <i>E</i> -diene-8,15-diol	Bz	Ac	iBu	H	Ac	<i>E. peplus</i>	44
64	2,5,7,14-tetraacetoxy-3-benzoyloxy-9-nicotinoyloxyjatropa-6(17),11 <i>E</i> -diene-8,15-diol	Bz	Ac	Ac	H	Nic	<i>E. peplus</i>	44
65	2,5,7,9,14-pentaacetoxy-3-benzoyloxyjatropa-6(17),11 <i>E</i> -diene-8,15-diol	Bz	Ac	Ac	H	Ac	<i>E. peplus</i>	44
66	pepluanin A	Bz	Ac	Ac	Ac	Nic	<i>E. peplus</i>	45
67	pepluanin B	Bz	Ac	MeBu	H	Nic	<i>E. peplus</i>	45
68	pepluanin C	Ac	iBu	Bz	Ac	Ac	<i>E. peplus</i>	45

Table 13. Jatrophanes-3

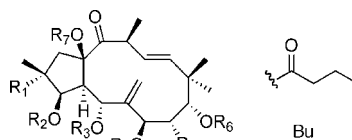


no.	name	R ₁	R ₂	plant	ref
69	(2 <i>S</i> *,3 <i>S</i> *,4 <i>R</i> *,5 <i>R</i> *,7 <i>S</i> *,8 <i>R</i> *,13 <i>R</i> *,14 <i>R</i> *,15 <i>R</i> *)-5,7,8,14-tetraacetoxy-3-benzoyloxy-15-hydroxyjatropa-6(17),11 <i>E</i> -dien-9-one	OAc	Ac	<i>E. semiperfoliata</i>	23
70	(2 <i>S</i> *,3 <i>S</i> *,4 <i>R</i> *,5 <i>R</i> *,7 <i>S</i> *,8 <i>R</i> *,13 <i>R</i> *,14 <i>R</i> *,15 <i>R</i> *)-5,7,14-triacetoxy-3-benzoyloxy-15-hydroxy-8-isobutyryloxyjatropa-6(17),11 <i>E</i> -dien-9-one	OiBu	Ac	<i>E. semiperfoliata</i>	23
71	(2 <i>S</i> *,3 <i>S</i> *,4 <i>R</i> *,5 <i>R</i> *,7 <i>S</i> *,8 <i>R</i> *,13 <i>R</i> *,14 <i>R</i> *,15 <i>R</i> *)-5,7-diacetoxy-3-benzoyloxy-8,14,15-trihydroxyjatropa-6(17),11 <i>E</i> -dien-9-one	OH	H	<i>E. semiperfoliata</i>	23
72	(2 <i>S</i> *,3 <i>S</i> *,4 <i>R</i> *,5 <i>R</i> *,7 <i>S</i> *,8 <i>R</i> *,13 <i>R</i> *,14 <i>R</i> *,15 <i>R</i> *)-5,7,8-triacetoxy-3-benzoyloxy-14,15-dihydroxyjatropa-6(17),11 <i>E</i> -dien-9-one	OAc	H	<i>E. semiperfoliata</i>	23
73	(2 <i>S</i> *,3 <i>S</i> *,4 <i>R</i> *,5 <i>R</i> *,7 <i>S</i> *,8 <i>R</i> *,13 <i>R</i> *,14 <i>R</i> *,15 <i>R</i> *)-5,7,14-triacetoxy-3-benzoyloxy-8,15-dihydroxyjatropa-6(17),11 <i>E</i> -dien-9-one	OH	Ac	<i>E. semiperfoliata</i>	23
74	(2 <i>S</i> *,3 <i>S</i> *,4 <i>R</i> *,5 <i>R</i> *,7 <i>R</i> *,13 <i>R</i> *,14 <i>R</i> *,15 <i>R</i> *)-5,7,14-triacetoxy-3-benzoyloxy-15-hydroxyjatropa-6(17),11 <i>E</i> -dien-9-one	H	Ac	<i>E. semiperfoliata</i>	23
75	(2 <i>S</i> *,3 <i>S</i> *,4 <i>R</i> *,5 <i>R</i> *,7 <i>R</i> *,13 <i>R</i> *,14 <i>S</i> *,15 <i>R</i> *)-5,7,14-triacetoxy-3-benzoyloxy-15-hydroxyjatropa-6(17),11 <i>E</i> -dien-9-one	—	—	<i>E. hyberna</i> subsp. <i>insularis</i> <i>E. semiperfoliata</i>	46 47

Table 14. Jatrophanes-4

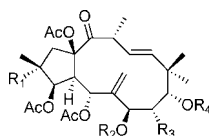


no.	name	R ₁	R ₂	R ₃	R ₄	R ₅	R ₆	plant	ref
76	(2 <i>S</i> *,3 <i>S</i> *,4 <i>R</i> *,5 <i>R</i> *,7 <i>S</i> *,8 <i>S</i> *,9 <i>S</i> *,13 <i>S</i> *,15 <i>R</i> *)-3,9,15-trihydroxy-5,7,8-triisobutyryloxyjatropa-6(17),11 <i>E</i> -dien-14-one	H	H	iBu	OiBu	OiBu	H	<i>E. segetalis</i>	38
77	(2 <i>S</i> *,3 <i>S</i> *,4 <i>R</i> *,5 <i>R</i> *,7 <i>S</i> *,8 <i>S</i> *,9 <i>S</i> *,13 <i>S</i> *,15 <i>R</i> *)-3,9,15-trihydroxy-7-isobutyryloxy-5,8-bis(2-methylbutanoyloxy)jatropa-6(17),11 <i>E</i> -dien-14-one	H	H	MeBu	OiBu	OMeBu	H	<i>E. segetalis</i>	38
78	(2 <i>S</i> *,3 <i>S</i> *,4 <i>R</i> *,5 <i>R</i> *,7 <i>S</i> *,8 <i>S</i> *,9 <i>S</i> *,13 <i>S</i> *,15 <i>R</i> *)-3,9,15-trihydroxy-5,7-diisobutyryloxy-8-(2-methylbutanoyloxy)jatropa-6(17),11 <i>E</i> -dien-14-one	H	H	iBu	OiBu	OMeBu	H	<i>E. segetalis</i>	38
79	1,5,8,9-tetraacetoxy-2-benzoyloxyacetoxo-7-isobutyryloxyjatropa-6(17),11 <i>E</i> -dien-14-one	OAc	BzOAc	Ac	OiBu	OAc	Ac	<i>E. segetalis</i>	38
80	5-acetoxy-3-benzoyloxy-9-cinnamoyloxy-15-hydroxyjatropa-6(17),11 <i>E</i> -dien-14-one	H	Bz	Ac	H	H	Cinn	<i>E. segetalis</i>	38
81	5-acetoxy-3,9-dicinnamoyloxy-15-hydroxyjatropa-6(17),11 <i>E</i> -dien-14-one	H	Cinn	Ac	H	H	Cinn	<i>E. segetalis</i>	38

Table 15. *Jatrphanes-5*

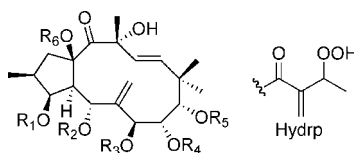
no.	name	R ₁	R ₂	R ₃	R ₄	R ₅	R ₆	R ₇	plant	ref
82	(2 <i>S</i> ,3 <i>S</i> ,4 <i>S</i> ,5 <i>R</i> ,7 <i>S</i> ,8 <i>S</i> ,9 <i>S</i> ,11 <i>E</i> ,13 <i>S</i> ,15 <i>R</i>)-5,7,8-tris(2-methylbutanoyloxy)-3,9,15-trihydroxyjatrpha-6(17),11-dien-14-one	H	H	MeBu	OMeBu	OMeBu	H	H	<i>E. terracina</i>	43
83	(2 <i>S</i> ,3 <i>S</i> ,4 <i>S</i> ,5 <i>R</i> ,7 <i>S</i> ,8 <i>S</i> ,9 <i>S</i> ,11 <i>E</i> ,13 <i>S</i> ,15 <i>R</i>)-3,9,15-trihydroxy-5,7,8-triisobutyryloxyjatrpha-6(17),11-dien-14-one	H	H	iBu	OiBu	OiBu	H	H	<i>E. terracina</i>	43
84	(2 <i>S</i> ,3 <i>S</i> ,4 <i>S</i> ,5 <i>R</i> ,7 <i>S</i> ,8 <i>S</i> ,9 <i>S</i> ,11 <i>E</i> ,13 <i>S</i> ,15 <i>R</i>)-3,9,15-trihydroxy-7-isobutyryloxy-5,8-bis(2-methylbutanoyloxy)jatrpha-6(17),11-dien-14-one	H	H	MeBu	OiBu	OMeBu	H	H	<i>E. terracina</i>	43
85	(2 <i>S</i> ,3 <i>S</i> ,4 <i>S</i> ,5 <i>R</i> ,7 <i>S</i> ,8 <i>S</i> ,9 <i>S</i> ,11 <i>E</i> ,13 <i>S</i> ,15 <i>R</i>)-3,9,15-trihydroxy-5,7-diisobutyryloxy-8-(2-methylbutanoyloxy)jatrpha-6(17),11-dien-14-one	H	H	iBu	OiBu	OMeBu	H	H	<i>E. terracina</i>	43
86	pepluanin D	H	Ac	Ac	Ac	H	Ac	Ac	<i>E. peplus</i>	45
87	pepluanin E	OAc	Bz	Ac	iBu	OH	Nic	H	<i>E. peplus</i>	45
88	2 α ,3 β ,5 α ,8 α ,9 α ,15 β -hexaacetoxy-7 β -benzoyloxyjatrpha-6(17),11 <i>E</i> -dien-14-one	OAc	Ac	Ac	Bz	OAc	Ac	Ac	<i>E. turczaninowii</i>	48
89	3 β ,5 α ,8 α ,9 α ,15 β -pentaacetoxy-7 β -benzoyloxyjatrpha-6(17),11 <i>E</i> -dien-14-one	H	Ac	Ac	Bz	OAc	Ac	Ac	<i>E. turczaninowii</i>	48
90	3 β ,5 α ,7 β ,8 α ,9 α ,15 β -hexaacetoxy-2 α -benzoyloxyjatrpha-6(17),11 <i>E</i> -dien-14-one	OBz	Ac	Ac	OAc	OAc	Ac	Ac	<i>E. turczaninowii</i>	48
91	(2 <i>S</i> ,3 <i>S</i> ,4 <i>S</i> ,5 <i>R</i> ,7 <i>S</i> ,8 <i>S</i> ,9 <i>S</i> ,13 <i>S</i> ,15 <i>R</i>)-3-acetoxy-5,9,15-trihydroxy-7-isobutyryloxy-8-(2-methylbutanoyloxy)-jatrpha-6(17),11 <i>E</i> -dien-14-one	H	Ac	H	iBu	OMeBu	H	H	<i>E. terracina</i>	43
92	(2 <i>S</i> *,3 <i>S</i> *,4 <i>R</i> *,5 <i>R</i> *,7 <i>S</i> *,8 <i>R</i> *,13 <i>S</i> *,15 <i>R</i> *)-5,7,8,9,15-pentaacetoxy-3 β -benzoyloxyjatrpha-6(17),11 <i>E</i> -dien-14-one	H	Bz	Ac	Ac	OAc	Ac	Ac	<i>E. mongolica</i>	49
93	(2 <i>R</i> ,3 <i>R</i> ,4 <i>R</i> ,5 <i>R</i> ,7 <i>S</i> ,8 <i>S</i> ,9 <i>S</i> ,11 <i>E</i> ,13 <i>S</i> ,15 <i>R</i>)-7,8,9-triacetoxy-3,15-dihydroxy-2,5-bis(2-methylbutanoyloxy)jatrpha-6(17),11-dien-14-one	OMeBu	H	MeBu	Ac	OAc	Ac	H	<i>E. obtusifolia</i>	50
94	(2 <i>R</i> ,3 <i>R</i> ,4 <i>R</i> ,5 <i>R</i> ,7 <i>S</i> ,8 <i>S</i> ,9 <i>S</i> ,11 <i>E</i> ,13 <i>S</i> ,15 <i>R</i>)-7,8,9-triacetoxy-3,7,8,9,15-pentahydroxy-2-isobutyryloxy-5-(2-methylbutanoyloxy)jatrpha-6(17),11-dien-14-one	OiBu	H	MeBu	Ac	OAc	Ac	H	<i>E. obtusifolia</i>	50
95	(2 <i>R</i> ,3 <i>R</i> ,4 <i>R</i> ,5 <i>R</i> ,7 <i>S</i> ,8 <i>S</i> ,9 <i>S</i> ,11 <i>E</i> ,13 <i>S</i> ,15 <i>R</i>)-7,8,9-triacetoxy-3,15-dihydroxy-2-nicotinoyloxy-5-(2-methylbutanoyloxy)jatrpha-6(17),11-dien-14-one	ONic	H	MeBu	Ac	OAc	Ac	H	<i>E. obtusifolia</i>	50
96	(2 <i>R</i> ,3 <i>R</i> ,4 <i>R</i> ,5 <i>R</i> ,7 <i>S</i> ,8 <i>S</i> ,9 <i>S</i> ,11 <i>E</i> ,13 <i>S</i> ,15 <i>R</i>)-8,9-diacetoxy-3,15-dihydroxy-7-isobutyryloxy-2,5-bis(2-methylbutanoyloxy)jatrpha-6(17),11-dien-14-one	OMeBu	H	MeBu	iBu	OAc	Ac	H	<i>E. obtusifolia</i>	50
97	(2 <i>R</i> ,3 <i>R</i> ,4 <i>R</i> ,5 <i>R</i> ,7 <i>S</i> ,8 <i>S</i> ,9 <i>S</i> ,11 <i>E</i> ,13 <i>S</i> ,15 <i>R</i>)-2,8,9-triacetoxy-3,15-dihydroxy-7-isobutyryloxy-5-(2-methylbutanoyloxy)jatrpha-6(17),11-dien-14-one	OAc	H	MeBu	iBu	OAc	Ac	H	<i>E. obtusifolia</i>	50
98	(2 <i>R</i> ,3 <i>R</i> ,4 <i>R</i> ,5 <i>R</i> ,7 <i>S</i> ,8 <i>S</i> ,9 <i>S</i> ,11 <i>E</i> ,13 <i>S</i> ,15 <i>R</i>)-7,9-diacetoxy-8-benzoyloxy-5,8,15-trihydroxy-2,3-bis(2-methylbutanoyloxy)jatrpha-6(17),11-dien-14-one	OMeBu	MeBu	H	Ac	OBz	Ac	H	<i>E. obtusifolia</i>	50
99	(2 <i>R</i> ,3 <i>R</i> ,4 <i>R</i> ,5 <i>R</i> ,7 <i>S</i> ,8 <i>S</i> ,9 <i>S</i> ,11 <i>E</i> ,13 <i>S</i> ,15 <i>R</i>)-8,9-diacetoxy-5,15-dihydroxy-7-isobutyryloxy-2,3-bis(2-methylbutanoyloxy)jatrpha-6(17),11-dien-14-one	OMeBu	MeBu	H	iBu	OAc	Ac	H	<i>E. obtusifolia</i>	50
100	(2 <i>R</i> ,3 <i>R</i> ,4 <i>R</i> ,5 <i>R</i> ,7 <i>S</i> ,8 <i>S</i> ,9 <i>S</i> ,11 <i>E</i> ,13 <i>S</i> ,15 <i>R</i>)-2,3,8,9-tetraacetoxy-15-hydroxy-5,7-diisobutyryloxyjatrpha-6(17),11-dien-14-one	OAc	Ac	iBu	iBu	OAc	Ac	H	<i>E. terracina</i>	43
101	(2 <i>R</i> ,3 <i>R</i> ,4 <i>R</i> ,5 <i>R</i> ,7 <i>S</i> ,8 <i>S</i> ,9 <i>S</i> ,11 <i>E</i> ,13 <i>S</i> ,15 <i>R</i>)-2,3,8,9-tetraacetoxy-5-benzoyloxy-15-hydroxy-7-isobutyryloxyjatrpha-6(17),11-dien-14-one	OAc	Ac	Bz	iBu	OAc	Ac	H	<i>E. terracina</i>	43
102	(2 <i>R</i> ,3 <i>R</i> ,4 <i>R</i> ,5 <i>R</i> ,7 <i>S</i> ,8 <i>S</i> ,9 <i>S</i> ,11 <i>E</i> ,13 <i>S</i> ,15 <i>R</i>)-2,5,8,9-tetraacetoxy-3-(benzoyloxyacetoxy)-15-hydroxy-7-isobutyryloxyjatrpha-6(17),11-dien-14-one	OAc	BzOAc	Ac	iBu	OAc	Ac	H	<i>E. terracina</i>	43
103	euphodendroidin A	OAc	H	iBu	Bz	OAc	Ac	H	<i>E. dendroides</i>	51
104	euphodendroidin B	OAc	H	MeBu	Bz	OAc	Ac	H	<i>E. dendroides</i>	51
105	euphodendroidin C	OAc	H	Nic	Bz	OAc	Ac	H	<i>E. dendroides</i>	51
106	euphodendroidin D	H	H	iBu	Bz	OAc	Ac	H	<i>E. dendroides</i>	51
107	euphodendroidin E	H	Ac	iBu	Bz	OAc	Ac	H	<i>E. dendroides</i>	51
108	euphodendroidin F	OH	Ac	iBu	Bz	OAc	Ac	H	<i>E. dendroides</i>	51
109	euphodendroidin G	OAc	Nic	Ac	iBu	OAc	Ac	H	<i>E. dendroides</i>	51
110	euphodendroidin H	H	Bz	Ac	iBu	OAc	Ac	H	<i>E. dendroides</i>	51
111	euphodendroidin I	ONic	Ac	iBu	Ac	OAc	Nic	Ac	<i>E. dendroides</i>	51
112	altotibetin A	H	Ac	Bz	Ac	OAc	Ac	Ac	<i>E. mongolica</i>	49
									<i>E. altotibetic</i>	52
113	altotibetin B	H	Ac	Bz	Bu	OAc	Ac	Ac	<i>E. altotibetic</i>	52
114	altotibetin C	OH	Ac	Bz	Ac	OAc	Ac	Ac	<i>E. altotibetic</i>	52
115	altotibetin D	OH	Ac	Bz	Bu	OAc	Ac	Ac	<i>E. altotibetic</i>	52

Table 16. Jatrophanes-6



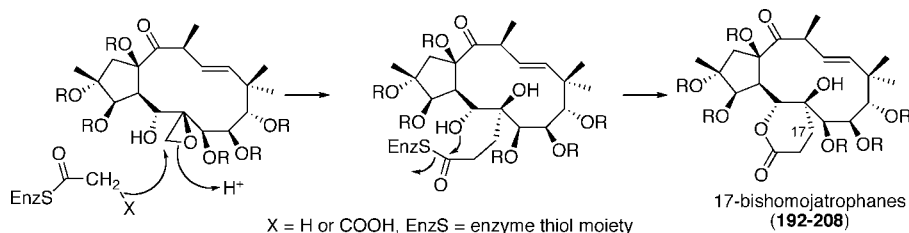
No.	name	R ₁	R ₂	R ₃	R ₄	plant	ref
116	esulatin D	OAc	Ac	H	Ac	<i>E. esula</i>	53
117	esulatin F	OAc	iBu	OAc	Ac	<i>E. esula</i>	54
118	(2 <i>R</i> *,3 <i>R</i> *,4 <i>S</i> *,5 <i>R</i> *,7 <i>R</i> *,9 <i>R</i> *,13 <i>R</i> *,15 <i>R</i> *)-2,3,5,7,15-pentaacetoxy-9-nicotinoyloxyljatropha-6(17),11 <i>E</i> -dien-14-one	OAc	Ac	H	Nic	<i>E. peplus</i>	44
119	3β,5α,7β,15β-tetraacetoxy-9-nicotinoyloxyljatropha-6(17),11 <i>E</i> -dien-14-one	H	Ac	H	Nic	<i>E. peplus</i>	55

Table 17. Jatrophanes-7

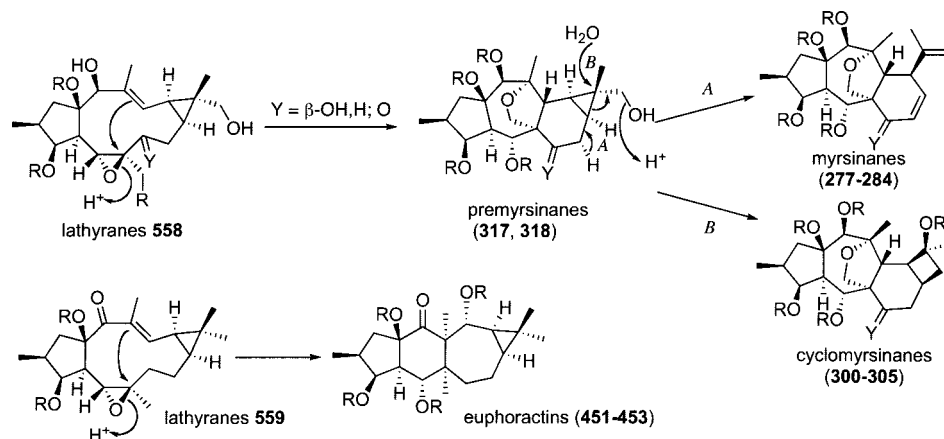


no.	name	R ₁	R ₂	R ₃	R ₄	R ₅	R ₆	plant	ref
120	amygdaloidins A	Ac	Ac	Ang	Ang	Nic	H	<i>E. amygdaloides</i>	56
121	amygdaloidins B	Ang	H	Ang	Ac	Nic	H	<i>E. amygdaloides</i>	56
122	amygdaloidins C	Hydrp	H	Ang	Ac	Nic	H	<i>E. amygdaloides</i>	56
123	amygdaloidins D	Ang	H	Ang	Ac	Ac	H	<i>E. amygdaloides</i>	56
124	amygdaloidins E	Ang	H	Hydrp	Ac	Ac	H	<i>E. amygdaloides</i>	56
125	amygdaloidins F	Ang	H	Hydrp	Ac	Ac	Ac	<i>E. amygdaloides</i>	56
126	amygdaloidins G	Ang	Ac	H	Hydrp	Ac	H	<i>E. amygdaloides</i>	56
127	amygdaloidins H	Hydrp	Ac	H	Ang	Ac	H	<i>E. amygdaloides</i>	56
128	amygdaloidins I	Ac	Hydrp	H	Ang	Ac	H	<i>E. amygdaloides</i>	56
129	amygdaloidins J	Hydrp	Ac	Ang	H	Ac	H	<i>E. amygdaloides</i>	56
130	amygdaloidins K	Ac	Hydrp	Ang	H	Ac	H	<i>E. amygdaloides</i>	56
131	amygdaloidins L	Ang	Ac	Hydrp	H	Ac	H	<i>E. amygdaloides</i>	56

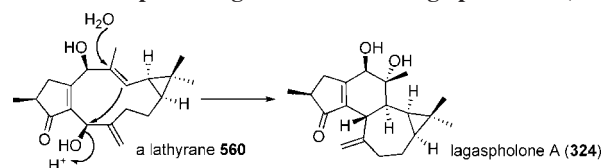
Scheme 4. Proposed Biogenesis of Lactones (192–208) by Incorporation of a C2 Unit (from Acetate or Malonate) into a Jatropane Precursor



Scheme 5. Proposed Biogenesis of Myrsinanes and Euphoractins from Epoxy Lathyrans

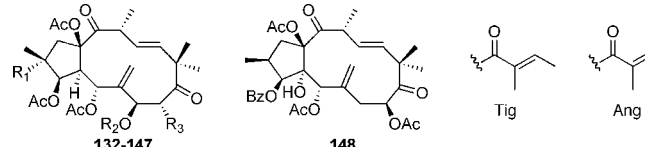


Scheme 6. Proposed Biogenetic Routes of Lagaspholone A (324)



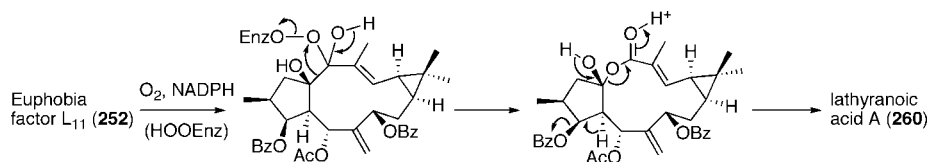
2.3.5. Jatropholanes (Table 44)

Lagaspholones A (324) and B (325) belong to jatropholanes are a new class of members with a 5,12-cyclojatropane skeleton.¹¹⁵

Table 18. *Jatrophanes-8*


no.	name	R ₁	R ₂	R ₃	plant	ref
132	(2 <i>R</i> *,3 <i>R</i> *,4 <i>S</i> *,5 <i>R</i> *,7 <i>S</i> *,8 <i>R</i> *,13 <i>R</i> *,15 <i>R</i> *)-2,3,5,7,15-pentaacetoxy-8-isobutyryloxyjatropha-6(17),11 <i>E</i> -diene-9,14-dione	OAc	Ac	OiBu	<i>E. semiperfoliata</i>	23
133	(2 <i>R</i> *,3 <i>R</i> *,4 <i>S</i> *,5 <i>R</i> *,7 <i>S</i> *,8 <i>R</i> *,13 <i>R</i> *,15 <i>R</i> *)-2,3,5,7,15-pentaacetoxy-8-tigloyloxyjatropha-6(17),11 <i>E</i> -diene-9,14-dione	OAc	Ac	OTig	<i>E. paralias</i> <i>E. semiperfoliata</i>	57 23
134	(2 <i>R</i> *,3 <i>R</i> *,4 <i>S</i> *,5 <i>R</i> *,7 <i>S</i> *,8 <i>R</i> *,13 <i>R</i> *,15 <i>R</i> *)-2,3,5,7,15-pentaacetoxy-8-benzoyloxyjatropha-6(17),11 <i>E</i> -diene-9,14-dione	OAc	Ac	OBz	<i>E. semiperfoliata</i>	23
135	(2 <i>R</i> *,3 <i>R</i> *,4 <i>S</i> *,5 <i>R</i> *,7 <i>S</i> *,8 <i>R</i> *,13 <i>R</i> *,15 <i>R</i> *)-3,5,7,8,15-pentaacetoxy-2-nicotinoyloxyjatropha-6(17),11 <i>E</i> -diene-9,14-dione	Nic	Ac	OAc	<i>E. semiperfoliata</i>	23
136	(2 <i>R</i> *,3 <i>R</i> *,4 <i>S</i> *,5 <i>R</i> *,7 <i>S</i> *,8 <i>R</i> *,13 <i>R</i> *,15 <i>R</i> *)-2,3,5,8,15-pentaacetoxy-7-benzoyloxyjatropha-6(17),11 <i>E</i> -diene-9,14-dione	OAc	Bz	OAc	<i>E. semiperfoliata</i>	23
137	(2 <i>R</i> *,3 <i>R</i> *,4 <i>S</i> *,5 <i>R</i> *,7 <i>S</i> *,8 <i>R</i> *,13 <i>R</i> *,15 <i>R</i> *)-2,3,5,8,15-pentaacetoxy-7-isobutyryloxyjatropha-6(17),11 <i>E</i> -diene-9,14-dione	OAc	iBu	OAc	<i>E. turczaninowii</i> <i>E. semiperfoliata</i>	58 23
138	(2 <i>R</i> *,3 <i>R</i> *,4 <i>S</i> *,5 <i>R</i> *,7 <i>S</i> *,8 <i>R</i> *,13 <i>R</i> *,15 <i>R</i> *)-3,5,7,8,15-pentaacetoxy-2-benzoyloxyjatropha-6(17),11 <i>E</i> -diene-9,14-dione	OBz	Ac	OAc	<i>E. semiperfoliata</i>	23
139	(2 <i>R</i> *,3 <i>R</i> *,4 <i>S</i> *,5 <i>R</i> *,7 <i>S</i> *,8 <i>R</i> *,13 <i>R</i> *,15 <i>R</i> *)-2,3,5,7,15-pentaacetoxy-8-hydroxyjatropha-6(17),11 <i>E</i> -diene-9,14-dione	OAc	Ac	OH	<i>E. semiperfoliata</i>	23
140	(2 <i>R</i> *,3 <i>R</i> *,4 <i>S</i> *,5 <i>R</i> *,7 <i>R</i> *,13 <i>R</i> *,15 <i>R</i> *)-2,3,5,15-tetraacetoxy-7-isobutyryloxyjatropha-6(17),11 <i>E</i> -diene-9,14-dione	OAc	iBu	H	<i>E. semiperfoliata</i>	23
141	esulatin B	OAc	Ac	H	<i>E. seguieriana</i> <i>E. semiperfoliata</i>	58 23
142	(2 <i>R</i> *,3 <i>R</i> *,4 <i>S</i> *,5 <i>R</i> *,7 <i>S</i> *,8 <i>R</i> *,13 <i>R</i> *,15 <i>R</i> *)-2,3,5,7,15-pentaacetoxy-8-isobutyryloxyjatropha-6(17),11 <i>E</i> -diene-9,14-dione	OAc	Ac	OiBu	<i>E. paralias</i>	59
143	(2 <i>R</i> *,3 <i>R</i> *,4 <i>S</i> *,5 <i>R</i> *,7 <i>S</i> *,8 <i>R</i> *,13 <i>R</i> *,15 <i>R</i> *)-2,3,5,7,15-pentaacetoxy-8-angeloyloxyjatropha-6(17),11 <i>E</i> -diene-9,14-dione	OAc	Ac	OAng	<i>E. paralias</i>	59
144	(2 <i>R</i> *,3 <i>R</i> *,4 <i>S</i> *,5 <i>R</i> *,7 <i>S</i> *,8 <i>R</i> *,13 <i>R</i> *,15 <i>R</i> *)-3,5,7,15-tetraacetoxy-8-isobutyryloxyjatropha-6(17),11 <i>E</i> -diene-9,14-dione	H	Ac	OiBu	<i>E. paralias</i>	59
145	(2 <i>R</i> *,3 <i>R</i> *,4 <i>S</i> *,5 <i>R</i> *,7 <i>S</i> *,8 <i>R</i> *,13 <i>R</i> *,15 <i>R</i> *)-2,3,5,7,15-pentaacetoxy-8-(2-methylbutanoyloxy)jatropha-6(17),11 <i>E</i> -diene-9,14-dione	OAc	Ac	OMeBu	<i>E. paralias</i>	59
146	(2 <i>R</i> *,3 <i>R</i> *,4 <i>S</i> *,5 <i>R</i> *,7 <i>S</i> *,8 <i>R</i> *,13 <i>R</i> *,15 <i>R</i> *)-2,3,5,7,15-pentaacetoxy-8-benzoyloxyjatropha-6(17),11 <i>E</i> -diene-9,14-dione	OAc	Ac	OBz	<i>E. paralias</i>	59
147	(2 <i>R</i> *,3 <i>R</i> *,4 <i>S</i> *,5 <i>R</i> *,7 <i>S</i> *,8 <i>R</i> *,13 <i>R</i> *,15 <i>R</i> *)-2,3,5,7,8,15-hexaacetoxyjatropha-6(17),11 <i>E</i> -diene-9,14-dione	OAc	Ac	OAc	<i>E. paralias</i>	59
148	pubescenol: 5 α ,8 β ,15 β -triacetoxy-3 β -benzoyloxy-4 α -hydroxy-13 β H-jatropha-6(17),11 <i>E</i> -diene-9,14-dione	—	—	—	<i>E. semiperfoliata</i> <i>E. pubescens</i>	23 26

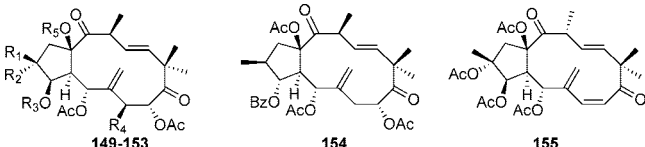
Scheme 7. Proposed Biogenesis of Lathyranic Acid A (260)

2.3.6. *Daphnanes* (Table 45)

Resiniferatoxin (326) and 327 forms intramolecular orthoester with phenylacetic acid.^{116,117,119} Langduin A (328),

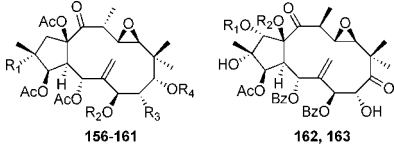
possessing an isopropyl group at C-14 and carbonyl group at C-13, is presumably derived from a tiglane by opening of the cyclopropane ring.⁸

Table 19. Jatrophanes-9



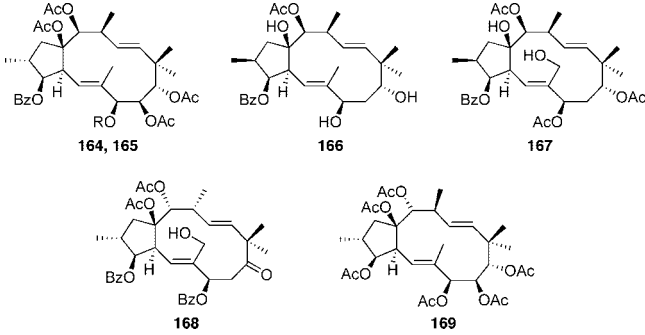
no.	name	R ₁	R ₂	R ₃	R ₄	R ₅	plant	ref
149	3β,5α,7β,8α,15β-pentaacetoxyljatropa-6(17),11E-diene-9,14-dione	Me	H	Ac	OAc	Ac	<i>E. turczaninowii</i>	48
150	(2 S,3 S,4 S,5 R,7 S,8 S,9 S,11 E,13 S,15 R)-3,5,8,15-tetraacetoxy-7-isobutyryloxyjatropa-6(17),11-diene-9,14-dione	Me	H	Ac	OiBu	Ac	<i>E. terracina</i>	43
151	(2 S*,3 S*,4 R*,5 R*,7 S*,8 R*,13 S*,15 R*)-5,7,8-triacetoxy-3-benzoyloxy-15-hydroxyjatropa-6(17),11 E-diene-9,14-dione	Me	H	Bz	OAc	H	<i>E. mongolica</i>	49
152	(2 S*,3 S*,4 R*,5 R*,8 R*,13 S*,15 R*)-5,8,15-triacetoxy-3-benzoyloxyjatropa-6(17),11 E-diene-9,14-dione	H	Me	Bz	H	Ac	<i>E. hyberna</i>	60
153	3β,5α,8α,15β-tetraacetoxyl-7β-benzoyloxyjatropa-6(17),11E-diene-9,14-dione	Me	H	Ac	OBz	Ac	<i>E. turczaninowii</i>	48
154	euphobubescenol	—	—	—	—	—	<i>E. pubescens</i>	61
155	esulatin E	—	—	—	—	—	<i>E. esula</i>	53

Table 20. Jatrophanes-10



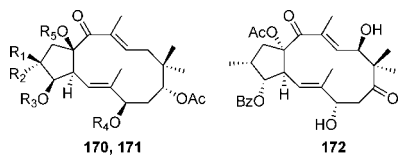
no.	name	R ₁	R ₂	R ₃	R ₄	plant	ref
156	kansuinin E	H	Ac	OBz	Nic	<i>E. kansui</i>	62
157	esulatin A	OAc	iBu	OAc	Ac	<i>E. esula</i> <i>E. salicifolia</i>	58 63
158	2α,3β,5α,9-tetraacetoxyl-11,12-epoxy-7β,8α-diisobutyryloxyjatroph-6(17)-en-14-one	OAc	iBu	OiBu	Ac	<i>E. salicifolia</i>	63
159	2α,3β,5α,6β,9α-pentaacetoxyl-11,12-epoxy-8α-isobutyryloxyjatroph-6(17)-en-14-one	OAc	Ac	OiBu	Ac	<i>E. salicifolia</i>	63
160	kansuinin F	H	Ac	OBz	Bz	<i>E. kansui</i>	64
161	kansuinin G	H	Ac	H	Nic	<i>E. kansui</i>	64
162	kansuinin C	Ac	H	—	—	<i>E. kansui</i>	65
163	kansuinin B	H	Ac	—	—	<i>E. kansui</i>	65

Table 21. Jatrophanes-11



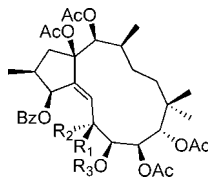
no.	name	R	plant	ref
164	(2 R*,3 S*,4 S*,7 S*,8 S*,9 S*,13 S*,14 S*,15 R*)-8,9,14,15-tetraacetoxy-3-benzoyloxyjatropa-5 E,11 E-dien-7-ol	H	<i>E. serrulata</i>	66
165	(2 R*,3 S*,4 S*,7 S*,8 S*,9 S*,13 S*,14 S*,15 R*)-7,8,9,14,15-pentaacetoxy-3-benzoyloxyjatropa-5 E,11 E-diene	Ac	<i>E. platyhyllus</i>	67
166	14β-acetoxy-3β-benzoyloxyjatropa-5E,11 E-diene-7β,9α,15β-triol	—	<i>E. helioscopia</i>	68
167	7β,9α,14β-triacetoxy-3β-benzoyloxyjatropa-5 E,11 E-diene-15β,17-diol	—	<i>E. helioscopia</i>	68
168	14α,15β-diacetoxy-3β,7β-dibenzoyloxy-17-hydroxy-(2βH,13βH)jatropa-5 E,11 E-dien-9-one	—	<i>E. helioscopia</i>	68
169	euphobubescene	—	<i>E. pubescens</i>	61

Table 22. Jatrophanes-12



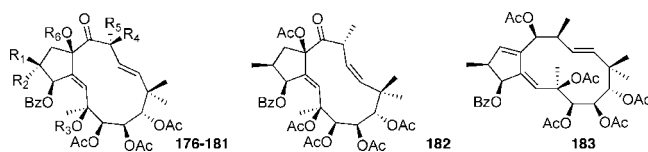
no.	name	R ₁	R ₂	R ₃	R ₄	R ₅	plant	ref
170	pubescens D	H	Me	Ac	Bz	H	<i>E. pubescens</i>	69
171	3β,9α,15β-triacetoxy-7β-butanoyloxyjatropha-5E,11 E-dien-14-one	Me	H	Ac	Bu	Ac	<i>E. pubescens</i>	70
172	euphoheliosnoid D	—	—	—	—	—	<i>E. helioscopia</i>	12

Table 23. Jatrophanes-13



no.	name	R ₁	R ₂	R ₃	plant	ref
173	(2 S,3 S,6 S,7 R,8 R,9 S,13 S,14 S,15 R)-7,8,9,14,15-pentaacetoxy-3-benzoyloxyjatropha-4 E,11 E-dien-6-ol	OH	Me	Ac	<i>E. serrulata</i>	66
174	(2 S*,3 S*,6 S*,7 R*,8 R*,9 S*,13 S*,14 S*,15 R*)-8,9,14,15-tetraacetoxy-3,7-dibenzoyloxyjatropha-4 E,11 E-dien-6-ol	OH	Me	Bz	<i>E. serrulata</i>	66
175	(2 S*,3* S, 6 R*,7 R*,8* R,9 S*,13 S*,14 S*,15 R*)-7,8,9,14,15-pentaacetoxy-3-benzoyloxyjatropha-4 E,11 E-dien-6-ol	Me	OH	Ac	<i>E. platyphyllos</i>	67

Table 24. Jatrophanes-14



no.	name	R ₁	R ₂	R ₃	R ₄	R ₅	R ₆	plant	ref
176	(2 R*,3 R*,6 S*,7 R*,8 R*,9 S*,13 S*,15 R*)-6,7,8,9,15-pentaacetoxy-3-benzoyloxy-2-hydroxyjatropha-4 E,11 E-dien-14-one	Me	OH	Ac	Me	H	Ac	<i>E. serrulata</i>	66
177	(2 R*,3 S*,6 S*,7 R*,8 R*,9 S*,13 S*,15 R*)-7,8,9,15-tetraacetoxy-3-benzoyloxy-6-hydroxyjatropha-4 E,11 E-dien-14-one	H	Me	H	Me	H	Ac	<i>E. serrulata</i> <i>E. platyphyllos</i>	66 67
178	(2 R*,3 S*,6 S*,7 R*,8 R*,9 S*,13 S*,15 R*)-6,7,8,9,15-pentaacetoxy-3-benzoyloxyjatropha-4 E,11 E-dien-14-one	H	Me	Ac	Me	H	Ac	<i>E. serrulata</i>	66
179	(2 R,3 R,6 R,7 R,8 R,9 S,13 S,15 R)-2,7,8,9-tetraacetoxy-3-benzoyloxy-6,15-dihydroxyjatropha-4 E,11 E-dien-14-one	Me	OAc	H	Me	H	H	<i>E. serrulata</i>	71
180	(2 R,3 R,6 S,7 R,8 R,9 S,13 S,15 R)-2,6,7,8,9,15-hexaacetoxy-3-benzoyloxyjatropha-4 E,11 E-dien-14-one	Me	OAc	Ac	Me	H	Ac	<i>E. serrulata</i>	71
181	serrulatin B	Me	H	Ac	H	Me	Ac	<i>E. serrulata</i>	72
182	(2 R,3 S,6 S,7 R,8 R,9 S,13 R,15 R)-6,7,8,9,15-pentaacetoxy-3-benzoyloxyjatropha-4 Z,11 E-dien-14-one	—	—	—	—	—	—	<i>E. serrulata</i>	73
183	(2 S*,3 S*,6 S*,7 R*,8 R*,9 S*,13 S*,14 S*)-6,7,8,9,14-pentaacetoxy-3-benzoyloxyjatropha-1(15),4 Z,11 E-triene	—	—	—	—	—	—	<i>E. serrulata</i>	73

2.3.7. Tiglianes (Tables 46–48)

12-*O*-Tetradecanoylphorbol 12-acetate (TPA) is a famous tumor promotor. A number of phorbol derivatives were isolated from *Euphorbia* species. Marco et al. presumed that **336** was not a natural product but rather an epimerization product of **331** during the isolation and chromatographic separation.⁵⁰ The C-6 of **358** and **359** is an aldehyde group.²¹

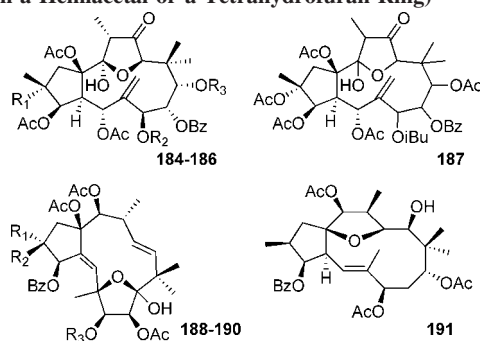
2.3.8. Ingenanes (Tables 49–52)

Ingenane diterpenoids have a very unique structural feature, that is, bicyclo[4.4.1]undecane core adopts a highly strained *inside–outside* skeleton. A large number of deriva-

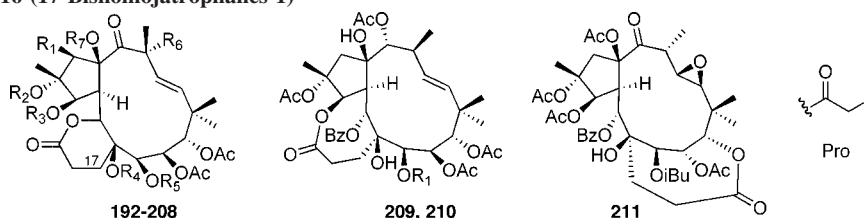
tives have been reported from this genus. Milliamine F isomer (**381**), isolated from *E. leuconeura*, has a unique [(benzamido)benzamido]benzoyl substituent.¹²⁸

2.3.9. Segetanes, Paralanes, Pepluanes, and Euphoractines (Tables 53–58)

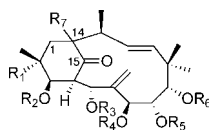
The skeleton of these 5/7/6/5-rings could be formed by transannular ring formation between 8,12- and 13,17-positions of the 12-membered ring of jatrophanes. Presegetanin (**436**) and **435** belong to a new class of carbon skeleton, presumably a biogenetic intermediate from jatrophanes to segetanes.^{38,138} Paralanes and pepluanes are

Table 25. Jatrophanes-15 (Jatrophanes with a Hemiacetal or a Tetrahydrofuran Ring)

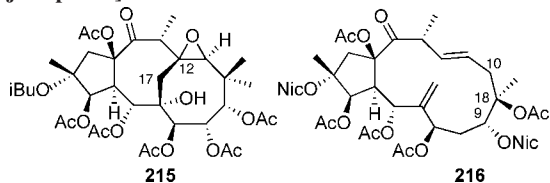
no.	name	R ₁	R ₂	R ₃	plant	ref
184	kansuinin A	H	Ac	Ac	<i>E. kansui</i>	65
185	kansuinin H	OH	Ac	Ac	<i>E. kansui</i>	64
186	kansuinin D	H	Ac	Nic	<i>E. kansui</i>	62
187	esulatin C	—	—	—	<i>E. esula</i>	58
188	serrulatin A: (2 <i>S</i> ,3 <i>S</i> ,6 <i>R</i> ,7 <i>S</i> ,8 <i>S</i> ,9 <i>S</i> ,13 <i>R</i> ,14 <i>S</i> ,15 <i>S</i>)-8,14,15-triacetoxy-3-benzoyloxy-6,9-epoxy-7-tigloyloxyjatropa-4 <i>Z</i> ,11 <i>E</i> -dien-9-ol	H	Me	Tig	<i>E. serrulata</i>	72
189	(2 <i>R</i> ,3 <i>S</i> ,6 <i>R</i> ,7 <i>S</i> ,8 <i>S</i> ,9 <i>S</i> ,13 <i>R</i> ,14 <i>S</i> ,15 <i>S</i>)-8,14,15-triacetoxy-3-benzoyloxy-6,9-epoxy-7-tigloyloxyjatropa-4 <i>E</i> ,11 <i>E</i> -dien-9-ol	Me	H	Tig	<i>E. serrulata</i>	71
190	(2 <i>R</i> ,3 <i>S</i> ,6 <i>S</i> ,7 <i>R</i> ,8 <i>S</i> ,9 <i>R</i> ,13 <i>R</i> ,14 <i>S</i> ,15 <i>R</i>)-8,14,15-triacetoxy-3,7-dibenzoyloxy-6,9-epoxyjatropa-4 <i>E</i> ,11 <i>E</i> -dien-9-ol	Me	H	Bz	<i>E. platyhyllus</i>	67
191	7β,9α,14β-triacetoxy-3β-benzoyloxy-12β,15β-epoxyjatroph-5 <i>E</i> -en-11β-ol	—	—	—	<i>E. helioscopia</i>	68

Table 26. Jatrophanes-16 (17-Bishomojatrophanes-1)

no.	name	R ₁	R ₂	R ₃	R ₄	R ₅	R ₆	R ₇	plant	ref
192	terracinolide A	H	Ac	Ac	Bz	iBu	H	Ac	<i>E. terracina</i>	73
									<i>E. segetalis</i>	38
193	terracinolide B	H	Ac	Ac	Ac	iBu	H	Ac	<i>E. terracina</i>	73
									<i>E. dendroides</i>	74
									<i>E. segetalis</i>	38
194	terracinolide C	H	Ac	H	Ac	iBu	H	Ac	<i>E. terracina</i>	75
									<i>E. dendroides</i>	74
									<i>E. segetalis</i>	38
195	terracinolide D	H	Ac	Ac	Bz	Ac	H	Ac	<i>E. terracina</i>	75
196	terracinolide E	H	Ac	Ac	Bz	Pr	H	Ac	<i>E. terracina</i>	75
									<i>E. segetalis</i>	38
197	terracinolide F	H	Ac	Ac	iBu	iBu	H	Ac	<i>E. terracina</i>	75
									<i>E. dendroides</i>	74
198	terracinolide G	H	Ac	Ac	Ac	iBu	H	H	<i>E. terracina</i>	75
199	terracinolide H	H	Ac	H	iBu	iBu	H	Ac	<i>E. segetalis</i>	38
200	terracinolide J	H	Ac	Ac	iBu	iBu	H	H	<i>E. dendroides</i>	74
201	terracinolide K	H	H	Ac	Ac	iBu	H	Ac	<i>E. dendroides</i>	74
202	terracinolide L	H	H	Ac	iBu	iBu	H	Ac	<i>E. dendroides</i>	74
203	13α-hydroxyterracinolide B	H	Ac	Ac	Ac	iBu	OH	Ac	<i>E. segetalis</i>	38
									<i>E. dendroides</i>	74
									<i>E. terracina</i>	28
204	13α-hydroxyterracinolide F	H	Ac	Ac	iBu	iBu	OH	Ac	<i>E. dendroides</i>	74
205	13α-hydroxyterracinolide G	H	Ac	Ac	Ac	iBu	OH	H	<i>E. terracina</i>	28
									<i>E. dendroides</i>	74
206	15- <i>O</i> -deacetyl-13α-hydroxyterracinolide A	H	Ac	Ac	Bz	iBu	OH	H	<i>E. terracina</i>	28
207	13α-hydroxyterracinolide I	OAc	Ac	Ac	Ac	iBu	OH	H	<i>E. segetalis</i>	38
									<i>E. terracina</i>	28
208	terracinolide I	OAc	Ac	Ac	Ac	iBu	H	H	<i>E. segetalis</i>	38
209	isoterracinolide A	iBu	—	—	—	—	—	—	<i>E. terracina</i>	28
210	isoterracinolide B	Pr	—	—	—	—	—	—	<i>E. terracina</i>	28
211	salicinolide	—	—	—	—	—	—	—	<i>E. salicifolia</i>	76

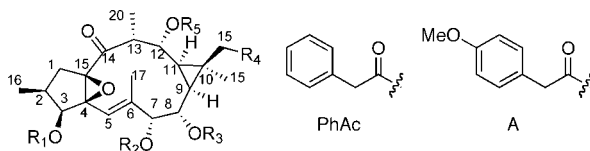
Table 27. Jatrophanes-17 [1(15→14) Abeojatrophanes]

no.	name	R ₁	R ₂	R ₃	R ₄	R ₅	R ₆	R ₇	plant	ref
212	(2 <i>S</i> ,3 <i>S</i> ,4 <i>R</i> ,5 <i>R</i> ,7 <i>S</i> ,8 <i>S</i> ,9 <i>S</i> ,11 <i>E</i> ,13 <i>S</i> ,14 <i>S</i>)-3,5,7,8,9,14-hexahydroxy-7-isobutyryloxy-5,8-bis(2-methylbutanoxy)-1(15→14)abeojatropha-6(17),11-dien-15-one	H	H	MeBu	iBu	MeBu	H	β-OH	<i>E. terracina</i>	43
213	abeodendroidin F	OH	Ac	iBu	Bz	Ac	Ac	β-OH	<i>E. dendroides</i>	74
214	<i>epi</i> -abeodendroidin F	OH	Ac	iBu	Bz	Ac	Ac	α-OH	<i>E. dendroides</i>	74

Table 28. Jatrophanes-18 [12,17-Cyclojatropane and 9(10→18) Abeojatropane]

no.	name	plant	ref
215	salicifoline	<i>E. salicifolia</i>	76
216	euphosalicin	<i>E. salicifolia</i>	63

derived from jatrophanes, and euphoractines are from lathyrans (section 3). Compound **445** possesses a rare aromatized D-ring.³⁸

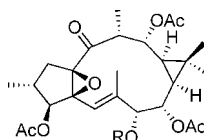
Table 29. Lathyrans-1

no.	name	R ₁	R ₂	R ₃	R ₄	R ₅	plant	ref
217	3β,12α-diacetoxy-19-hydroxy-7α,8α-ditigloyloxyngol	Ac	Tigl	Tigl	OH	Ac	<i>E. acurensis</i>	77
218	3β,12α,19-triacetoxy-7α-hydroxy-8α-ditigloyloxyngol	Ac	H	Tigl	OAc	Ac	<i>E. acurensis</i>	77
219	12α,19-diacetoxy-3β,7α-hydroxy-8α-ditigloyloxyngol	H	H	Tigl	OAc	Ac	<i>E. acurensis</i>	77
220	3β,8α,12α-triacetoxy-7α-isovaleryloxyngol	Ac	iVal	Ac	H	Ac	<i>E. acurensis</i>	77
221	3β,8α,12α-triacetoxy-7α-angeloyloxyngol	Ac	Ang	Ac	H	Ac	<i>E. acurensis</i>	77
222	3β,12α-diacetoxy-7α,8α-ditigloyloxyngol	Ac	Tigl	Tigl	H	Ac	<i>E. acurensis</i>	77
223	3β,8α,12α-triacetoxy-7α-tigloyloxyngol	Ac	Tigl	Ac	H	Ac	<i>E. acurensis</i>	77
224	3β,12α-diacetoxy-8α-methoxy-7α-tigloyloxyngol	Ac	Tigl	Me	H	Ac	<i>E. acurensis</i>	77
225	3β,12α-triacetoxy-7α-hydroxy-8α-tigloyloxyngol	Ac	H	Tigl	H	Ac	<i>E. acurensis</i>	77
							<i>E. lactea</i>	78
226	3β,7α,12α-triacetoxy-8α-isovaleryloxyngol	Ac	Ac	iVal	H	Ac	<i>E. tirucalli</i>	79
227	3β,7α,12α-triacetoxy-8α-benzoyloxyngol	Ac	Ac	Bz	H	Ac	<i>E. nivulia</i>	80
							<i>E. kamerunica</i>	81
							<i>E. hermentiana</i>	82
228	3β,12α-diacetoxy-7α-angeloyloxy-8α-methoxyngol	Ac	Ang	Me	H	Ac	<i>E. nivulia</i>	80
							<i>E. hermentiana</i>	81
229	12α-acetoxy-7α-angeloyloxy-3β-hydroxy-8α-methoxyngol	H	Ang	Me	H	Ac	<i>E. nivulia</i>	80
							<i>E. ingens</i>	83
230	3β,12α-diacetoxy-8α-benzoyloxy-7α-hydroxyngol	Ac	H	Bz	H	Ac	<i>E. nivulia</i>	80
231	3β,12α-diacetoxy-7α-benzoyloxy-8α-nicotinoyloxyngol	Ac	Bz	Nic	H	Ac	<i>E. nivulia</i>	80
232	3β-acetoxy-7α-angeloyloxy-12α-hydroxy-8α-methoxyngol	Ac	Ang	Me	H	H	<i>E. nivulia</i>	84
233	3β,12α-diacetoxy-8α-methoxy-7α-hydroxyngol	Ac	H	Me	H	Ac	<i>E. nivulia</i>	84
234	3β,12α-diacetoxy-7α-angeloyloxy-8α-hydroxyngol	Ac	Ang	H	H	Ac	<i>E. nivulia</i>	84
235	3β,12α,19-triacetoxy-8α-nicotinoyloxy-7α-phenylacetoxingol	Ac	PhAc	Nic	OAc	Ac	<i>E. poisonii</i>	85
236	3β,12α,19-triacetoxy-8α-hydroxy-7α-phenylacetoxingol	Ac	PhAc	H	OAc	Ac	<i>E. poisonii</i>	85
237	3β,19-diacetoxy-8α,12α-dihydroxy-7α-phenylacetoxingol	Ac	PhAc	H	OAc	H	<i>E. poisonii</i>	85
238	7α,8α,12α-triacetoxy-3β-phenylacetoxingol	PhAc	Ac	Ac	H	Ac	<i>E. officinarum</i>	86
239	7α,8α,12α-triacetoxy-3β-(<i>p</i> -methoxyphenyl)acetoxingol	A	Ac	Ac	H	Ac	<i>E. officinarum</i>	86
240	7α,12α-diacetoxy-8α-methoxy-3β-phenylacetoxingol	PhAc	Ac	Me	H	Ac	<i>E. officinarum</i>	86

2.4. Triterpenoids (Tables 59–65)

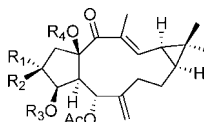
Tetracyclic, pentacyclic triterpenoids, some secotriterpenoids along with several other kinds triterpenoids are isolated from many plants of *Euphorbia* species. Among lanosterol type compounds, antiquol B (**460**) has a rare 19(10→9) abeoepuphane skeleton.¹⁴⁴ Several oleanes with shifted methyl and desmethyl groups (**477–482**) were isolated.^{20,26,146,150,153} 27-Nor-3β-hydroxycycloartan-25-one (**496**), (2*E*)-25,26,27-trinor-3β-hydroxycycloartan-22-en-24-al (**498**) and 25,26,27-trinor-3β-hydroxycycloartan-24-al (**499**) are rare *nor*-derivatives reported first in this species.¹³⁶ 24-Hydroperoxycycloartan-25-en-3β-ol (**500**) is the only C-24 hydroperoxy compound.¹³⁶ Peplusol (**508**) is an acyclic triterpene alcohol.¹⁴⁵ Anhydrobisfarnesol (**509**), an anhydro derivative of bisfarnesol, has been isolated from the latex of *E. lateriflora*. Its structure was confirmed by the synthesis using a photochemical isomerization.¹⁵⁸

Table 30. Lathyranes-2



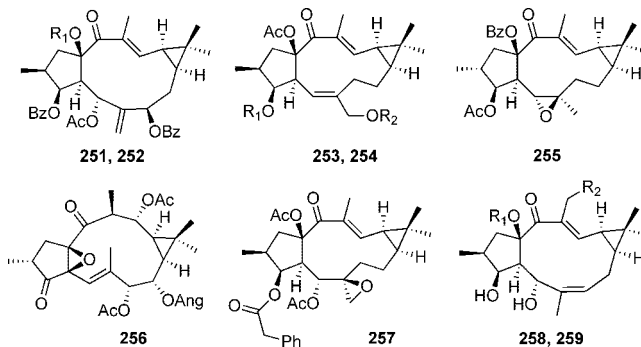
no.	name	R	plant	ref
241	3 β ,7 α ,8 α ,12 α -tetraacetoxy-2- <i>epi</i> -ingol	Ac	<i>E. portulacoides</i>	16
242	3 β ,8 α ,12 α -triacetoxy-7 α -isobutanoyloxy-2- <i>epi</i> -ingol	iBu	<i>E. portulacoides</i>	16
243	3 β ,8 α ,12 α -triacetoxy-7 α -methylbutanoyloxy-2- <i>epi</i> -ingol	MeBu	<i>E. portulacoides</i>	16
244	3 β ,8 α ,12 α -triacetoxy-7 α -benzoyloxy-2- <i>epi</i> -ingol	Bz	<i>E. portulacoides</i>	16

Table 31. Lathyranes-3.



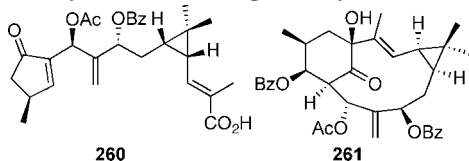
no.	name	R ₁	R ₂	R ₃	R ₄	plant	ref
245	(2 <i>R</i> *,3 <i>S</i> *,4 <i>R</i> *,5 <i>R</i> *,9 <i>S</i> *,11 <i>S</i> *,15 <i>R</i> *)-5 α ,15 β -diacetoxy-3 β -benzoyloxy-lathyrane-6(17),12 <i>E</i> -dien-14-one	Me	H	Bz	Ac	<i>E. hyberna</i>	60
246	(2 <i>R</i> *,3 <i>S</i> *,4 <i>R</i> *,5 <i>R</i> *,9 <i>S</i> *,11 <i>S</i> *,15 <i>R</i> *)-3,5,15-triacetoxy-lathyrane-6(17),12 <i>E</i> -dien-14-one	Me	H	Ac	Ac	<i>E. hyberna</i>	60
247	Euphorbia factor L ₃	H	Me	Bz	Ac	<i>E. villosa</i>	87
248	Euphorbia factor L ₈	H	Me	Nic	Ac	<i>E. lathyris</i>	88
249	(2 <i>S</i> *,3 <i>S</i> *,4 <i>R</i> *,5 <i>R</i> *,9 <i>S</i> *,11 <i>S</i> *,15 <i>R</i> *)-5-acetoxy-3-benzoyloxy-15-hydroxylathyrane-6(17),12 <i>E</i> -dien-14-one	H	Me	Bz	H	<i>E. lathyris</i>	89
250	(2 <i>S</i> *,3 <i>S</i> *,4 <i>R</i> *,5 <i>R</i> *,9 <i>S</i> *,11 <i>S</i> *,15 <i>R</i> *)-5,15-diacetoxy-3-phenylacetoxylathyrane-6(17),12 <i>E</i> -dien-14-one	H	Me	PhAc	Ac	<i>E. pithyusa</i> subsp. <i>cupanii</i>	90

Table 32. Lathyranes-4



no.	name	R ₁	R ₂	plant	ref
251	Euphorbia factor L ₂	Ac	—	<i>E. lathyris</i>	88
252	Euphorbia factor L ₁₁	H	—	<i>E. lathyris</i>	91
253	Euphorbia factor L ₁₀	C ₅ H ₁₁ CO	H	<i>E. lathyris</i>	92
254	15 β ,17-diacetoxy-3 β -benzoyloxyisolathyrone	Bz	Ac	<i>E. lathyris</i>	93
255	(2 <i>R</i> *,3 <i>S</i> *,4 <i>R</i> *,5 <i>R</i> *,6 <i>R</i> *,11 <i>S</i> *,15 <i>R</i> *)-3-acetoxy-15-benzoyloxy-5,6-epoxylathyrane-12 <i>E</i> -en-14-one	—	—	<i>E. villosa</i>	87
256	(2 <i>R</i> *,4 <i>R</i> *,7 <i>R</i> *,8 <i>S</i> *,9 <i>S</i> *,11 <i>R</i> *,12 <i>S</i> *,13 <i>S</i> *,15 <i>R</i> *)-7,12-diacetoxy-8-angeloyloxy-4,15-epoxylathyrane-5 <i>E</i> -ene-3,14-dione	—	—	<i>E. segetalis</i>	93
257	Euphorbia factors L ₁	—	—	<i>E. lathyris</i>	88
258	15 β -cinnamoyloxy-3 β ,5 α -dihydroxylathyrane-6Z,12 <i>E</i> -dien-14-one	Cinn	H	<i>E. kansuensis</i>	94
259	15 β -cinnamoyloxy-3 β ,5 α ,20-trihydroxylathyrane-6Z,12 <i>E</i> -dien-14-one	Cinn	OH	<i>E. kansuensis</i>	94

Table 33. Lathyranes-5 (Rearranged Lathyranes)

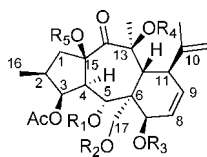


no.	name	plant	ref
260	lathyranoic acid A	<i>E. lathyris</i>	91
261	lathyrane A	<i>E. lathyris</i>	95

2.5. Steroids (Tables 66 and 67)

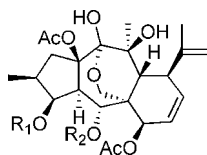
Tanaka et al. isolated several ergostane-type steroids (**510-515**) from *E. chamaesyce*.^{149,151} 5 α -Stigmastane-3 β ,6 α -diol (**517**) and 5 α -stigmastane-3 β ,5,6 β -triol (**519**) were found to be obtained in *E. boetica*.¹¹¹ In 2002, Rahman et al. reported the isolation of a new geniculatoside (**519**) from aerial parts of *E. geniculata* Linn.¹⁵⁹ In addition, β -sitosterol and daucosterol are present in many plants of this species, such

Table 34. Myrsinanes-1



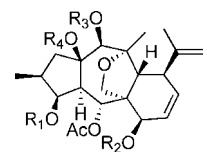
no.	name	R ₁	R ₂	R ₃	R ₄	R ₅	plant	ref
262	13-deacetylisodecypidone	Bu	H	Ac	H	Ac	<i>E. decipiens</i>	96
263	13-deacetylisodecypinone	Bz	H	Ac	H	Ac	<i>E. decipiens</i>	96
264	isodecypidone	Bu	H	Ac	Ac	Ac	<i>E. decipiens</i>	96
265	17-acetoxy-13-deacetyldecipinone	Bz	Ac	Ac	H	Ac	<i>E. decipiens</i>	97
266	13-deacetyldecipinone	Bz	Ac	Ac	H	H	<i>E. decipiens</i>	97
267	13-deacetyldecypidone	Bu	Ac	Ac	H	H	<i>E. decipiens</i>	97
268	decypinone	Bz	Ac	Ac	Ac	H	<i>E. decipiens</i>	98,99
269	decypidone	Bu	Ac	Ac	Ac	H	<i>E. decipiens</i>	98
270	isodecypinone	Bz	H	Ac	Ac	Ac	<i>E. decipiens</i>	98
271	17-acetoxy-13-deacetylisodecypidone	Bu	Ac	Ac	H	Ac	<i>E. decipiens</i>	100

Table 35. Myrsinanes-2



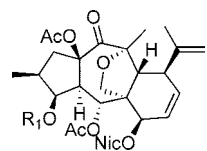
no.	name	R ₁	R ₂	plant	ref
272	3β,7β,15β-tri- <i>O</i> -acetyl-5α- <i>O</i> -nicotinoyl-13β,14β-dihydroxy-14-desoxo-14α,6α-epoxymethanomyrsinol	Ac	Nic	<i>E. decipiens</i>	101
273	decypinone ester A	Bu	Nic	<i>E. decipiens</i>	96
274	decypinone B	Ac	Bz	<i>E. decipiens</i>	102
275	decypinone C	Ac	Bu	<i>E. decipiens</i>	102
276	3β,7β,15β-tri- <i>O</i> -acetyl-5α- <i>O</i> -benzoyl-13β,14β-dihydroxy-14-desoxo-14α,6α-epoxymethanomyrsinol	Bu	Bz	<i>E. decipiens</i>	101

Table 36. Myrsinanes-3



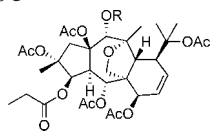
no.	name	R ₁	R ₂	R ₃	R ₄	plant	ref
277	euphorprolitherin C: 5α,15β-di- <i>O</i> -acetyl-7β,14β-di- <i>O</i> -benzoyl-14-desoxo-3β- <i>O</i> -propanoylmyrsinol	Pr	Bz	Bz	Ac	<i>E. prolifera</i>	103
278	5α,15β-di- <i>O</i> -acetyl-7β,14β-di- <i>O</i> -nicotinoyl-14-desoxo-3β- <i>O</i> -propanoylmyrsinol	Pr	Nic	Nic	Ac	<i>E. seguieriana</i>	19
279	3β,5α,15β-tri- <i>O</i> -acetyl-7β,14β-di- <i>O</i> -nicotinoyl-14-desoxomyrsinol	Ac	Nic	Nic	Ac	<i>E. seguieriana</i>	19
280	3β,5α,15β-tri- <i>O</i> -acetyl-7β- <i>O</i> -benzoyl-14β- <i>O</i> -nicotinoyl-14-desoxomyrsinol	Ac	Bz	Nic	Ac	<i>E. seguieriana</i>	19
281	5α,15β-di- <i>O</i> -acetyl-7β- <i>O</i> -benzoyl-14β- <i>O</i> -nicotinoyl-14-desoxo-3β- <i>O</i> -propanoylmyrsinol	Pr	Bz	Nic	Ac	<i>E. seguieriana</i>	19
282	5α,14β,15β-tri- <i>O</i> -acetyl-7β- <i>O</i> -benzoyl-14-desoxo-3β- <i>O</i> -propanoylmyrsinol	Pr	Bz	Ac	Ac	<i>E. seguieriana</i>	19
283	5α,14β,15β-tri- <i>O</i> -acetyl-7β- <i>O</i> -nicotinoyl-14-desoxo-3β- <i>O</i> -propanoylmyrsinol	Pr	Nic	Ac	Ac	<i>E. seguieriana</i>	19
284	5α,14β-di- <i>O</i> -acetyl-15β-hydroxy-7β- <i>O</i> -nicotinoyl-14-desoxo-3β- <i>O</i> -propanoylmyrsinol	Pr	Nic	Ac	H	<i>E. seguieriana</i>	19

Table 37. Myrsinanes-4



no.	name	R ₁	R ₂	plant	ref
285	15β- <i>O</i> -acetyl-3β- <i>O</i> -butanoyl-7β- <i>O</i> -nicotinoyl-5α- <i>O</i> -propanoylmyrsinol	Bu	Pr	<i>E. myrsinites</i>	104
286	15β- <i>O</i> -acetyl-3β,5α-di- <i>O</i> -butanoyl-7β- <i>O</i> -nicotinoylmyrsinol	Pr	Pr	<i>E. myrsinites</i>	104
287	15β- <i>O</i> -acetyl-7β- <i>O</i> -nicotinoyl-3β,5α-di- <i>O</i> -propanoylmyrsinol	Bu	Bu	<i>E. myrsinites</i>	104
288	15β- <i>O</i> -acetyl-5α- <i>O</i> -butanoyl-7β- <i>O</i> -nicotinoyl-3β- <i>O</i> -propanoylmyrsinol	Pr	Bu	<i>E. myrsinites</i>	104

Table 38. Myrsinanes-5



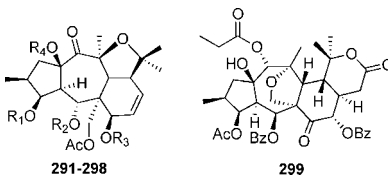
no.	name	R	plant	ref
289	euphorprolitherin A	Bz	<i>E. prolifera</i>	103
290	euphorprolitherin B	MeBu	<i>E. prolifera</i>	103

as *E. boetica*, *E. segetalis*, *E. altotibetic*, *E. aleppica*, *E. quinquecostata*, and *E. latifolia*.^{30,52,111,114,147,150}

2.6. Cerebrosides and Glycerols (Tables 68 and 69)

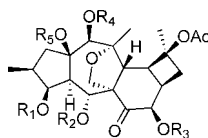
A complex mixture of four glucocerebrosides, **522** ($n = 21$), **523** ($n = 1, 3$), and **524**, was isolated from *E. peplis*.¹⁶³

Table 39. Myrsinanes-6



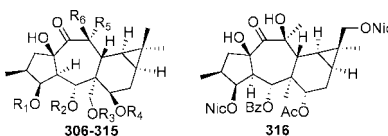
no.	name	R ₁	R ₂	R ₃	R ₄	plant	ref
291	3β-O-benzoyl-5α,7β,17-tri-O-acetyl-15β-hydroxycheiradone	Bz	Ac	Ac	H	<i>E. decipiens</i>	101
292	3β,5α,17-tri-O-acetyl-7β-O-benzoyl-15β-hydroxycheiradone	Ac	Ac	Bz	H	<i>E. decipiens</i>	105
293	3β,5α,15β,17-tetra-O-acetyl-7β-O-benzoylcheiradone	Ac	Ac	Bz	Ac	<i>E. decipiens</i>	105
294	3β,5α,15β,17-tetra-O-acetyl-7β-O-nicotinoylcheiradone	Ac	Ac	Nic	Ac	<i>E. decipiens</i>	105
295	15β-O-acetylcheiradone	Ac	Bz	Ac	Ac	<i>E. decipiens</i>	96
296	cheiradone	Ac	Bz	Ac	H	<i>E. cheiradenia</i>	106
297	cheiradone A	Ac	Bz	H	H	<i>E. cheiradenia</i>	106
298	cheiradone B	H	Bz	Ac	H	<i>E. cheiradenia</i>	106
299	myrsinol analog with C ₉ –C ₁₀ lactone ring	—	—	—	—	<i>E. prolifera</i>	107

Table 40. Cyclomyrsinanes

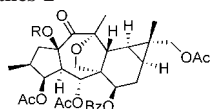


no.	name	R ₁	R ₂	R ₃	R ₄	R ₅	plant	ref
300	3β,5α,10β,14β,15β-penta-O-acetyl-8β-O-(2-methylbutanoyl)cyclomyrsinol	Ac	Ac	MeBuAc	Ac	Ac	<i>E. teheranica</i>	18
301	5α,10β,14β,15β-tetra-O-acetyl-8β-O-(2-methylbutanoyl)-3β-O-nicotinoylcyclomyrsinol	Nic	Ac	MeBuAc	Ac	Ac	<i>E. teheranica</i>	18
302	3β,10β,15β-tri-O-acetyl-8β-O-isobutyryl-5α,14β-O-dinicotinoylcyclomyrsinol	Ac	Nic	iBu	Nic	Ac	<i>E. seguieriana</i>	19
303	5α,10β,15β-tri-O-acetyl-8β,14β-O-dinicotinoyl-3β-O-propanoylcyclomyrsinol	Pr	Ac	Nic	Nic	Ac	<i>E. seguieriana</i>	19
304	euphorprolitherin D: 5α,10β,14β-tri-O-acetyl-8β-O-benzoyl-3β-O-propanoylcyclomyrsinol	Pr	Ac	Bz	Ac	H	<i>E. prolifera</i>	18
305	sPr5	Pr	Ac	Bz	Ac	Ac	<i>E. prolifera</i>	110

Table 41. Premyrsinanes-1



no.	name	R ₁	R ₂	R ₃	R ₄	R ₅	R ₆	plant	ref
306	eufoboetol	H	H	H	H	OH	Me	<i>E. boetica</i>	111
307	kandovanol ester A	Ac	Bz	Ac	Ac	Me	OH	<i>E. decipiens</i>	96
308	kandovanol ester B	Ac	Bu	Ac	Ac	Me	OH	<i>E. decipiens</i>	96
309	3β,7β,13β,17-tetraacetoxy-5α-isobutyryloxy-premyrsinol	Ac	iBu	Ac	Ac	Me	OAc	<i>E. pithyusa</i> subsp. <i>cupanii</i>	90
310	7β,13β,17-triacetoxy-5α-isobutyryloxy-3β-propanoyloxy-premyrsinol	Pr	iBu	Ac	Ac	Me	OAc	<i>E. pithyusa</i> subsp. <i>cupanii</i>	90
311	7β,13-diacetoxy-15α-isobutyryloxy-17-nicotinoyloxy-3β-propanoyloxy-premyrsinol	Pr	iBu	Nic	Ac	Me	OAc	<i>E. pithyusa</i> subsp. <i>cupanii</i>	90
312	7β,13β-diacetoxy-17-isobutyryloxy-5-(2-methylbutanoyloxy)-3β-propanoyloxy-premyrsinol	Pr	MeBu	iBu	Ac	Me	OAc	<i>E. pithyusa</i> subsp. <i>cupanii</i>	90
313	7β,13β-diacetoxy-5α,17-diisobutyryloxy-3β-propanoyloxy-premyrsinol	Pr	iBu	iBu	Ac	Me	OAc	<i>E. pithyusa</i> subsp. <i>cupanii</i>	90
314	7β,13β-diacetoxy-5α-isobutyryloxy-3β,17-dipropanoyloxy-premyrsinol	Pr	iBu	Pr	Ac	Me	OAc	<i>E. pithyusa</i> subsp. <i>cupanii</i>	90
315	7β,13β,17-triacetoxy-5α-benzoyloxy-3β-propanoyloxy-premyrsinol	Pr	Bz	Ac	Ac	Me	OAc	<i>E. pithyusa</i> subsp. <i>cupanii</i>	90
316	7α-O-acetyl-5β-O-benzoyl-13β-hydroxy-3β,18-dinicotinoyloxy-premyrsinol	—	—	—	—	—	—	<i>E. seguieriana</i>	109

Table 42. Premyrsinanes-2

no.	name	R	plant	ref
317	karajinone A	Ac	<i>E. decipiens</i>	112
318	karajinone B	H	<i>E. decipiens</i>	112

These were the first examples of glycosphingolipids from the *Euphorbia* genus. Other cerebrosides (522–526) and a sphingosine (527) have been isolated from several species.

Cateni et al. isolated three new glycolipids (528, 530, and 531) with anti-inflammatory activity from *E. cyparissias*.¹⁶⁷

2.7. Phenolics (Tables 70 and 71)

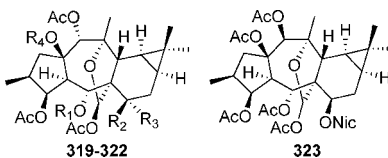
Several acetophenones and its glycoside were isolated from *E. portulacoides*, *E. quinquecostata*, *E. quinquecostata*, *E. fischeriana*, and *E. ebracteolata*.^{13,16,30,168} Unusual phenol dimers (541 and 542) were reported from *E. ebracteolata*.¹⁶⁸

2.8. Flavonoids (Table 72)

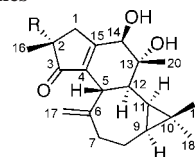
In 2004, a new flavonol glycoside, quercetin 3-*O*-6'-(3-hydroxyl-3-methylglutaryl)- β -D-glucopyranoside (543), and four known flavonoids, kaempferol 3-*O*-2''-galloyl- β -D-glucopyranoside, kaempferol 3-*O*-rutinoside, quercetin 3-*O*- β -D-glucopyranoside, and rutin, were isolated from the aerial parts of *E. ebracteolata*.¹⁶⁹ Nishimura et al. reported the isolation of one new flavonoid galactoside, quercetin 3-*O*-(2'',3''-digalloyl)- β -D-galactopyranoside (544), from *E. lunulata*, along with four known ones, quercetin 3-*O*-(2''-galloyl)- β -D-galactopyranoside (545), hyperin, and quercetin.¹⁷⁰ Recently, Zhang et al. reported six known flavonoids such as licochalcone A, 4,2',4'-trihydroxychalcone, echinatin, licochalcone B, glabrone, and 5,7,4'-trihydroxyflavanone from *E. helioscopia*.¹² These compounds were isolated from the species for the first time. In addition, the common flavonoids such as kaempferol, kaempferol 3-*O*-L-rhaside, kaempferol 3-*O*- β -D-glucopyranoside, quercetin, quercetin 3-*O*- β -D-glucopyranoside, and astragalins have been isolated from many plants of this species such as *E. latifolia*, *E. altotibetic*, and *E. aleppica*.^{52,113,114,147}

2.9. Miscellaneous Compounds (Table 73)

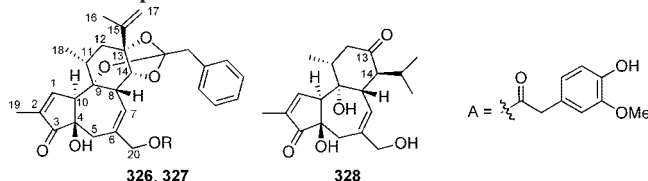
Lee et al. isolated a new ellagitannin, jolkinin (546), from the fresh whole plant of *E. jolkinii* in 2004.¹⁷¹ It has a unique hexacyclic structure attached to the 2,4-positions of 1-*O*-galloyl-3,6-(*R*)-hexahydroxydiphenyl- β -D-glucopyranose. 3,3',4'-

Table 43. Premyrsinanes-3

no.	name	R ₁	R ₂	R ₃	R ₄	plant	ref
319	aleppicatin A	Tigl	H	OAc	Ac	<i>E. aleppica</i>	113
320	aleppicatin B	Tigl	H	OAc	Tigl	<i>E. aleppica</i>	113
321	euphoreppine A	Ac	OTigl	H	Ac	<i>E. aleppica</i>	114
322	euphoreppine B	Ac	OTigl	H	Tigl	<i>E. aleppica</i>	114
323	3 β ,5 α ,14 β ,15 β ,17-penta- <i>O</i> -acetyl-7- <i>O</i> -nicotinoyl euphoppin	—	—	—	—	<i>E. decipiens</i>	96

Table 44. Jatropholanes

no.	name	R	plant	ref
324	lagaspholone A	H	<i>E. lagascae</i>	115
325	lagaspholone B	OH	<i>E. lagascae</i>	115

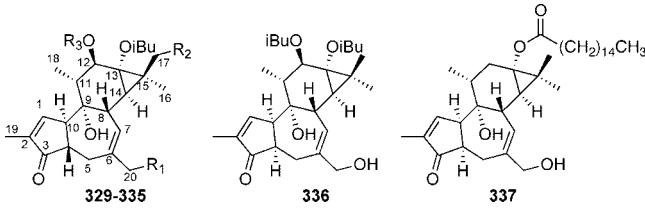
Table 45. Daphnanes

no.	name	R	plant	ref
326	20-(4-hydroxy-3-methoxyphenyl-acetoxy)-9,13,14-(orthophenyl-acetoxy)resiniferol (resiniferatoxin)	A	<i>E. poissonii</i>	116,117
			<i>E. resinifera</i>	118,119
327	20-hydroxy-9,13,14-(orthophenyl-acetoxy)resiniferol	H	<i>E. poissonii</i>	116
328	langduin A	—	<i>E. fischeriana</i>	8

Tri-*O*-methyl-4-*O*-[α -L-rhamnopyranosyl-(1'' \rightarrow 6'')- β -D-glucopyranosyl]ellagic acid has been isolated from *E. quinquecostata*.³⁰ In 2002, Su et al. isolated a new dihydrobenzo[*b*]furan neolignan, (–)-*trans*-9-acetyl-4,9'-di-*O*-methyl-3'-de-*O*-methyldehydrodiconiferyl alcohol (547), from the stem wood of *E. quinquecostata* along with 3,4-dimethoxycinnamaldehyde.³⁶

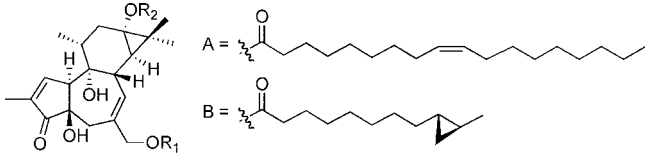
Coumarins in *Euphorbia* genus are not very rich. The main compounds are scopoletin (548),⁵² 7,7'-dihydroxy-6,8'-bicycoumarin (bicoumol, 549),³⁶ 6,7,8-trimethoxycoumarin (550),³⁰ and 6-hydroxy-7-methoxycoumarin (isoscopoletin, 551).³⁶ Two alkaloids, uracil and uridine, were isolated from *E. altotibetic* in 2003.⁵² In 1999, Che et al. reported the isolation of physcion from *E. fischeriana*.¹³ Gallic acid, 3,3'-di-*O*-methyl ellagic acid, and 3,4,3'-tri-*O*-methyl ellagic acid 4'-*O*- β -D-glucopyranoside were isolated from *E. fischeriana*, *E. sessiliflora*, and *E. lunulata*.^{13,15,75} Octacosyl ferulate (552 and 553),¹⁷² 1-glycerin hexadecanoate, 1-octacosanol, 9-*cis*-tricosene, 4-hydroxybenzoic acid, and its methyl ether were found to exist in *E. fischeriana*, *E. humifusa*, *E. latifolia*, and *E. aleppica*.^{84,113,145,147,148}

Table 46. Tiglides-1



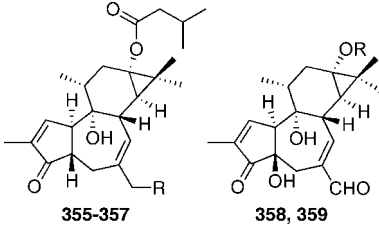
no.	name	R ₁	R ₂	R ₃	plant	ref
329	13 α -isobutyryloxy-4-deoxy-12 β -tigloyloxyphorbol	OH	H	Tigl	<i>E. semiperfoliata</i>	23
330	12 β ,13 α -diisobutyryloxy-4,20-dideoxyphorbol	H	H	iBu	<i>E. obtusifolia</i>	50
331	12 β ,13 α -diisobutyryloxy-4-deoxyphorbol	OH	H	iBu	<i>E. obtusifolia</i>	50
332	17-acetoxy-12,13 α -diisobutyryloxy-4-deoxyphorbol	OH	OAc	iBu	<i>E. obtusifolia</i>	50
333	17-acetoxy-12 β ,13 α -diisobutyryloxy-4,20-dideoxyphorbol	H	OAc	iBu	<i>E. obtusifolia</i>	50
334	20-acetoxy-12 β ,13 α -diisobutyryloxy-4-deoxyphorbol	OAc	H	iBu	<i>E. obtusifolia</i>	50
335	12 β -benzoyloxy-13 α -isobutyryloxy-4-deoxyphorbol	OH	H	Bz	<i>E. semiperfoliata</i>	23
336	12 β ,13 α -diisobutyryloxy-4-deoxy-4- <i>epi</i> -phorbol (4- <i>epi</i> -331)	—	—	—	<i>E. obtusifolia</i>	50
337	13 α -hexadecanoyl-4,12-dideoxy-4- <i>epi</i> -phorbol	—	—	—	<i>E. guyaniana</i>	120

Table 47. Tiglides-2



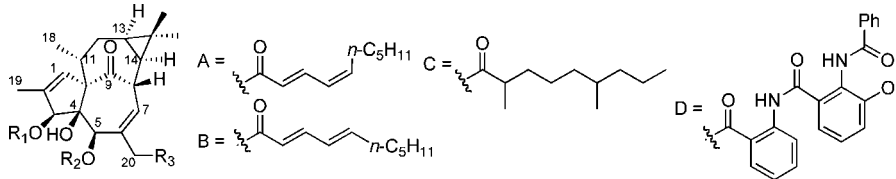
no.	name	R ₁	R ₂	plant	ref
338	20-acetoxy-13-isobutyryloxy-12-deoxyphorbol	Ac	iBu	<i>E. resinifera</i>	11
339	13-hexadecanoyloxy-12-deoxyphorbol	H	CO(CH ₂) ₁₄ CH ₃	<i>E. fischeriana</i>	8
340	prostratin: 13-acetoxy-12-deoxyphorbol	H	Ac	<i>E. fischeriana</i> <i>E. cornigera</i>	8 121
341	20-acetoxy-13-(9Z-octadecanoyloxy)-12-deoxyphorbol	Ac	A	<i>E. fischeriana</i>	21
342	20-acetoxy-13-angeloyloxy-12-deoxyphorbol	Ac	Ang	<i>E. poisonii</i> <i>E. resinifera</i>	122 11
343	20-acetoxy-12-deoxy-13-phenylacetoxyporbol	Ac	PhAc	<i>E. poisonii</i>	116
344	20-hydroxy-13-(<i>E</i> -9,10-methanoundecanoyloxy)-12-deoxyphorbol	H	B	<i>E. poisonii</i>	123
345	13-angeloyloxy-20-hydroxy-12-deoxyphorbol	H	Ang	<i>E. poisonii</i>	122
346	13-acetoxy-20-benzoyloxy-12-deoxyphorbol	Ac	Bz	<i>E. cornigera</i>	124
347	13-acetoxy-20- <i>p</i> -methoxybenzoyloxy-12-deoxyphorbol	Ac	<i>p</i> -MeOC ₆ H ₄ CO	<i>E. cornigera</i>	124
348	20-angeloyloxy-13-decanoyloxy-12-deoxyphorbol	CO(CH ₂) ₈ CH ₃	Ang	<i>E. cornigera</i>	124
349	13-decanoyloxy-20-tigloyloxy-12-deoxyphorbol	CO(CH ₂) ₈ CH ₃	Tigl	<i>E. cornigera</i>	124
350	13-acetoxy-20-decanoyloxy-12-deoxyphorbol	Ac	CO(CH ₂) ₈ CH ₃	<i>E. cornigera</i>	124
351	13-butanoyloxy-20-decanoyloxy-12-deoxyphorbol	Bu	CO(CH ₂) ₈ CH ₃	<i>E. cornigera</i>	124
352	20-decanoyloxy-13-hexanoyloxy-12-deoxyphorbol	CO(CH ₂) ₄ CH ₃	CO(CH ₂) ₈ CH ₃	<i>E. cornigera</i>	124
353	20-decanoyloxy-13-octanoyloxy-12-deoxyphorbol	CO(CH ₂) ₆ CH ₃	CO(CH ₂) ₈ CH ₃	<i>E. cornigera</i>	124
354	20-decanoyloxy-13-dodecanoyloxy-12-deoxyphorbol	CO(CH ₂) ₁₀ CH ₃	CO(CH ₂) ₈ CH ₃	<i>E. cornigera</i>	124

Table 48. Tiglides-3



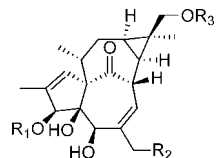
no.	name	R	plant	ref
355	13-(2,3-dimethylbutanoyloxy)-4,12,20-trideoxyphorbol	H	<i>E. pithyusa</i> subsp. <i>cupanii</i>	90
356	13-(2,3-dimethylbutanoyloxy)-4,12-dideoxyphorbol	OH	<i>E. pithyusa</i> subsp. <i>cupanii</i>	90
357	20-acetoxy-13-(2,3-dimethylbutanoyloxy)-4,12-dideoxyphorbol	OAc	<i>E. pithyusa</i> subsp. <i>cupanii</i>	90
358	13-acetoxy-12-deoxyphorbolaldehyde	Ac	<i>E. fischeriana</i>	21
359	13-hexadecanoyloxy-12-deoxyphorbolaldehyde	CO(CH ₂) ₁₄ CH ₃	<i>E. fischeriana</i>	21

Table 49. Ingenanes-1



no.	name	R ₁	R ₂	R ₃	plant	ref
360	20- <i>O</i> -(2' <i>E</i> ,4' <i>E</i> -decadienoyl)ingenol	H	H	OB	<i>E. kansui</i>	65
361	20- <i>O</i> -(2' <i>E</i> ,4' <i>Z</i> -decadienoyl)ingenol	H	H	OA	<i>E. kansui</i>	65
362	3β- <i>O</i> -(2' <i>E</i> ,4' <i>Z</i> -decadienoyl)ingenol	A	H	OH	<i>E. kansui</i>	65
363	3β- <i>O</i> -(2' <i>E</i> ,4' <i>E</i> -decadienoyl)ingenol	B	H	OH	<i>E. kansui</i>	65
364	5β- <i>O</i> -acetyl-3β- <i>O</i> -(2' <i>E</i> ,4' <i>Z</i> -decadienoyl)ingenol	A	Ac	OH	<i>E. kansui</i>	65
365	20- <i>O</i> -acetyl-3β- <i>O</i> -(2' <i>E</i> ,4' <i>Z</i> -decadienoyl)ingenol	A	H	OAc	<i>E. kansui</i>	65
366	20- <i>O</i> -acetyl-3β- <i>O</i> -(2' <i>E</i> ,4' <i>E</i> -decadienoyl)ingenol	B	H	OAc	<i>E. kansui</i>	65
367	20- <i>O</i> -decanoyl)ingenol	H	H	OCO(CH ₂) ₈ CH ₃	<i>E. kansui</i>	65
368	5β- <i>O</i> -(2' <i>E</i> ,4' <i>E</i> -decadienoyl)ingenol	H	B	OH	<i>E. kansui</i>	65
369	20- <i>O</i> -myristoyl)ingenol	H	H	OCO (CH ₂) ₁₂ CH ₃	<i>E. wallichii</i>	125
370	3β- <i>O</i> -myristoyl)ingenol	CO(CH ₂) ₁₂ CH ₃	H	OH	<i>E. wallichii</i>	125
371	3β- <i>O</i> -angeloyl)ingenol	Ang	H	OH	<i>E. peplus</i>	44
372	20- <i>O</i> -acetyl-3β- <i>O</i> -angeloyl)ingenol	Ang	H	OAc	<i>E. peplus</i>	44
373	5β,20-di- <i>O</i> -acetyl-3β- <i>O</i> -angeloyl)ingenol	Ang	Ac	OAc	<i>E. acurvensis</i>	77
					<i>E. canariensis</i>	7
374	3β- <i>O</i> -angeloyl-20-deoxyingenol	Ang	H	H	<i>E. paralias</i>	59
375	kansuiphorin C	Ac	Bz	H	<i>E. kansui</i>	126
376	Euphorbia factor L ₅ : 20- <i>O</i> -hexadecanoyl)ingenol	H	H	OCOC ₁₅ H ₃₁	<i>E. quinquecostata</i>	118
					<i>E. lathyris</i>	127
377	ingenol	H	H	OH	<i>E. lathyris</i>	88
378	Euphorbia factor L ₄	COC ₁₅ H ₃₁	H	OH	<i>E. lathyris</i>	88
379	Euphorbia factor L ₆	COC ₁₄ H ₁₇	H	OH	<i>E. lathyris</i>	88
380	ingenol 3β-(2,6-dimethylnonanoate)	C	H	OH	<i>E. resinifera</i>	11
381	milliamine F isomer	D	H	OPr	<i>E. leuconeura</i>	128
382	3β- <i>O</i> -(2' <i>E</i> ,4' <i>Z</i> -decadienoyl)-20-deoxyingenol	A	H	H	<i>E. kansui</i>	62
383	3β- <i>O</i> -(2' <i>E</i> ,4' <i>E</i> -decadienoyl)-20-deoxyingenol	B	H	H	<i>E. kansui</i>	62
384	20- <i>O</i> -acetyl)ingenol	H	H	OAc	<i>E. segetalis</i>	38
					<i>E. kansui</i>	129
385	20-deoxyingenol	H	H	H	<i>E. segetalis</i>	38
					<i>E. kansui</i>	129

Table 50. Ingenanes-2



no.	name	R ₁	R ₂	R ₃	plant	ref
386	20-acetoxy-3β- <i>O</i> -angeloyl-17-angeloyloxy)ingenol	Ang	OAc	Ang	<i>E. segetalis</i>	38
387	17-acetoxy-3β- <i>O</i> -angeloyl-20-deoxyingenol	Ang	H	Ac	<i>E. acurvensis</i>	77
388	3β- <i>O</i> -angeloyl-17-angeloyloxy-20-deoxyingenol	Ang	H	Ang	<i>E. paralias</i>	59
					<i>E. segetalis</i>	38
389	3β- <i>O</i> -benzoyl-17-benzoyloxy-20-deoxyingenol	Bz	H	Bz	<i>E. portulacoides</i>	16
390	20- <i>O</i> -hexadecanoyl-17-hydroxyingenol	H	OCO(CH ₂) ₁₄ CH ₃	H	<i>E. quinquecostata</i>	30
391	17-(2 <i>Z</i> ,4 <i>E</i> ,6 <i>Z</i> -2,4,6-tridecanoyloxy)ingenol	H	OH	2,4,6-decatrienoyl	<i>E. cauducifolia</i>	130
392	3β- <i>O</i> -angeloyl-17-(2 <i>Z</i> ,4 <i>E</i> ,6 <i>Z</i> -tridecanoyloxy)ingenol	Ang	OH	2,4,6-decatrienoyl	<i>E. cauducifolia</i>	130
393	3β- <i>O</i> -acetyl-20- <i>O</i> -angeloyl-17-hydroxyingenol	Ac	OAng	H	<i>E. cauducifolia</i>	130
394	17-acetoxy-3β- <i>O</i> -angeloyl)ingenol	Ang	OH	Ac	<i>E. cauducifolia</i>	130
395	20-acetoxy-3β- <i>O</i> -angeloyl-17-hydroxyingenol	Ang	OAc	H	<i>E. cauducifolia</i>	130
					<i>E. hermetiana</i>	131
396	3β- <i>O</i> -angeloyl-17-benzoyloxy)ingenol	Ang	OH	Bz	<i>E. cauducifolia</i>	130
					<i>E. canariensis</i>	132
397	20-acetoxy-3β- <i>O</i> -angeloyl-17-benzoyloxy)ingenol	Ang	OAc	Bz	<i>E. cauducifolia</i>	130
					<i>E. canariensis</i>	132
398	3β- <i>O</i> -angeloyl-17-benzoyloxy-20-deoxyingenol	Ang	H	Bz	<i>E. esula</i>	133

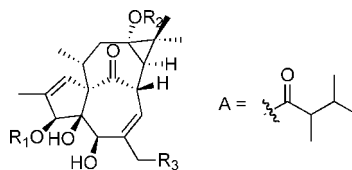
3. Hypothetical Biogenetic Route of the Rearranged Diterpenoids

Most polycyclic and macrocyclic diterpenoids are biosynthesized from geranylgeranyl diphosphate (Scheme 1).²²

3.1. Biosynthesis from Jatrophanes

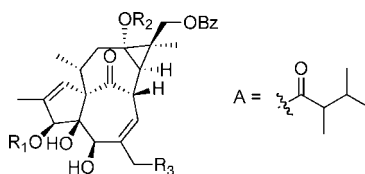
Bicyclic jatrophanes are converted to tetracyclic paralianes and pepluanes as shown in Scheme 2.^{44,139} Successive transannular reaction of a jatropane **554** would

Table 51. Ingenanes-3



no.	name	R ₁	R ₂	R ₃	plant	ref
399	20- <i>O</i> -acetyl-13β- <i>O</i> -(2,3-dimethylbutanoyl)-13α- <i>O</i> -dodecanoylingenol	A	CO(CH ₂) ₁₀ CH ₃	OAc	<i>E. kansui</i>	62
400	3β- <i>O</i> -(2,3-dimethylbutanoyl)-13α- <i>O</i> -dodecanoyl-20-deoxyingenol	A	CO(CH ₂) ₁₀ CH ₃	H	<i>E. kansui</i>	62
401	20- <i>O</i> -(2,3-dimethylbutanoyl)-13α- <i>O</i> -dodecanoylingenol	H	CO(CH ₂) ₁₀ CH ₃	OA	<i>E. kansui</i>	62
402	3β- <i>O</i> -(2,3-dimethylbutanoyl)-13α- <i>O</i> -dodecanoylingenol	A	CO(CH ₂) ₁₀ CH ₃	OH	<i>E. kansui</i> <i>E. cyparissias</i>	134 62 49
403	3β- <i>O</i> -benzoyl-13α- <i>O</i> -dodecanoylingenol	Bz	CO(CH ₂) ₁₀ CH ₃	OH	<i>E. kansui</i>	135
404	20- <i>O</i> -benzoyl-13α- <i>O</i> -dodecanoylingenol	H	CO(CH ₂) ₁₀ CH ₃	OBz	<i>E. kansui</i>	135
405	3β- <i>O</i> -(2,3-dimethylbutanoyl)-13α- <i>O</i> -octanoylingenol	A	CO(CH ₂) ₆ CH ₃	OH	<i>E. esula</i>	133
406	3β- <i>O</i> -benzoyl-13α- <i>O</i> -octanoylingenol	Bz	CO(CH ₂) ₆ CH ₃	OH	<i>E. esula</i>	133

Table 52. Ingenanes-4



no.	name	R ₁	R ₂	R ₃	plant	ref
407	13α-acetoxy-3β- <i>O</i> -benzoyl-17-benzoyloxyingenol	Bz	Ac	H	<i>E. segetalis</i>	38
408	13α-acetoxy-3β- <i>O</i> -angeloyl-17-benzoyloxyingenol	Ang	Ac	H	<i>E. segetalis</i>	38
409	17-benzoyloxy-3β,13α-bis- <i>O</i> -(2,3-dimethylbutanoyloxy)ingenol	A	A	H	<i>E. esula</i>	133
410	17-benzoyloxy-3β,13α-bis- <i>O</i> -(2,3-dimethylbutanoyloxy)-20-hydroxyingenol	A	A	OH	<i>E. esula</i>	133
411	17-benzoyloxy-13α,20-bis- <i>O</i> -(2,3-dimethylbutanoyloxy)-3β-hydroxyingenol	H	A	OA	<i>E. esula</i>	133
412	17-benzoyloxy-17,20-dihydroxy-13α-octanoyloxyingenol	H	CO(CH ₂) ₆ CH ₃	OH	<i>E. esula</i>	133
413	3β,17-dibenzoyloxy-20-hydroxy-13α-octanoyloxyingenol	Bz	CO(CH ₂) ₆ CH ₃	OH	<i>E. esula</i>	133
414	17,20-dibenzoyloxy-3β-hydroxy-13α-octanoyloxyingenol	H	CO(CH ₂) ₆ CH ₃	OBz	<i>E. esula</i>	133
415	3β- <i>O</i> -benzoyl-17-benzoyloxy-13α-octanoyloxyingenol	Bz	CO(CH ₂) ₆ CH ₃	H	<i>E. esula</i>	133
416	17-benzoyloxy-3β- <i>O</i> -(2,3-dimethylbutanoyloxy)-13α-octanoyloxy-20-hydroxyingenol	A	CO(CH ₂) ₆ CH ₃	OH	<i>E. esula</i>	133
417	17-benzoyloxy-20-(2,3-dimethylbutanoyloxy)-13α-octanoyloxy-3β-hydroxyingenol	H	CO(CH ₂) ₆ CH ₃	OA	<i>E. esula</i>	133
418	3β,13α,17-tri- <i>O</i> -benzoyloxy-20-hydroxyingenol	Bz	Bz	OH	<i>E. esula</i>	133
419	13α,17-dibenzoyloxy-3β- <i>O</i> -(2,3-dimethylbutanoyloxy)-20-hydroxyingenol	A	Bz	OH	<i>E. esula</i>	133
420	3β,17-dibenzoyloxy-3α- <i>O</i> -(2,3-dimethylbutanoyloxy)-20-hydroxyingenol	Bz	A	OH	<i>E. esula</i>	133
421	13α-acetoxy-17-benzoyloxy-3β- <i>O</i> -(2,3-dimethylbutanoyloxy)ingenol	A	Bz	H	<i>E. esula</i>	133

afford a 5/6/5/5-ring system of paraliene **555**. Oxidation followed by ring expansion and acetylation leads to a pepluane **556**. 1(15→14) *Abeojatropane* skeleton (**212**–**214**) could be derived from **557** by pinacol rearrangement (Scheme 3).⁴³ This mechanism can be applied to the biosynthesis of lathyranone A (**261**).⁹⁵ The C2-unit of bishomojatrophanes (**192**–**208**) is to be from acetyl or malonyl thiol ester moieties (Scheme 4).⁷⁵

3.2. Biosynthesis from Lathyranes

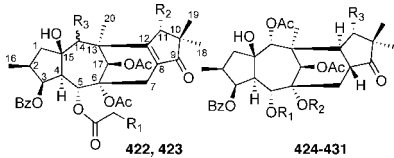
Three types of myrsinane diterpenoids and euphoractins would be formed from epoxy lathyranes (**558** or **559**) as shown in Scheme 5.¹⁹ The rare 5/6/7/3-ring system of lagaspholone A (**324**), a jatrophanolone-type skeleton, is from lathyranone **560** (Scheme 6).¹¹⁵ Lathyranic acid (**260**) would be an oxidative degradation product of Euphorbia factor L₁₁ (**252**) (Scheme 7).⁹¹

4. Biological Activity

4.1. Antiproliferative Activity

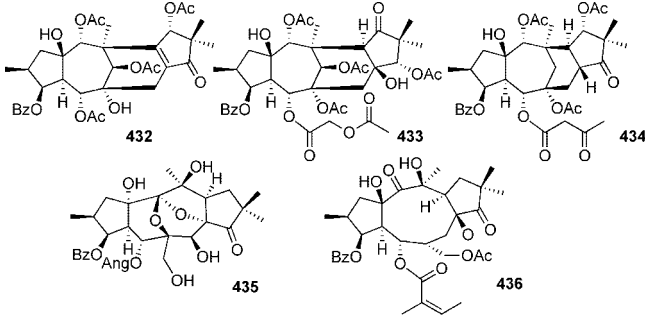
In 1997, Xu et al. reported that yuexiandajisu B (**57**) from *E. ebracteolata* inhibited proliferation of B lymphocytes *in vitro* preliminary bioassays.⁴² Valente et al. reported the *in vitro* effect of pubescenol (**147**), helioscopinolide A (**20**), helioscopinolide B (**21**), and pubescene D (**170**) on the human tumor cell lines MCF-7 (breast adenocarcinoma), NCI-H460 (nonsmall cell lung cancer), and SF-268 (CNS cancer). All four of the compounds were shown to be moderate inhibitors of the growth of these cell lines.²⁶ Putranjivain A, a tannin extracted from the whole plant of *E. jolkini* Bioss, was reported to inhibit the proliferation of MCF-7 by blocking cell cycle progression in the G₀/G₁ phase and inducing apoptosis.¹⁷³ In 2005, Nishimura et al. reported the proliferation activity of acetone extract of the whole plants of *E. lunulata* for insulin- and interleukin-10 (IL-10)-dependent cell lines.¹⁷⁰ Fractionation of the active extract led to the isolation of quercetin 3-*O*-(2'',3''-digalloyl)-

Table 53. Segetanes-1



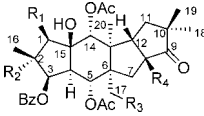
no.	name	R ₁	R ₂	R ₃	plant	ref
422	segetene B	OAc	H	α-OAc	<i>E. paralias</i>	57
423	segetene A	H	OAc	β-OAc	<i>E. paralias</i>	57
424	(2 S*,3 S*,4 R*,5 R*,6 R*,8 R*,12 S*,13 R*,14 R*,15 R*)-6,14,17-triacetoxy-5-(2-acetoxyacetoxy)-3-benzoyloxy-15-hydroxysegetan-9-one	COCH ₂ OAc	Ac	H	<i>E. segetalis</i>	38
425	(2 S*,3 S*,4 R*,5 R*,6 R*,8 R*,12 S*,13 R*,14 R*,15 R*)-6,11,14,17-tetraacetoxy-5-(2-acetoxyacetoxy)-3-benzoyloxy-15-hydroxysegetan-9-one	COCH ₂ OAc	Ac	OAc	<i>E. paralias</i> <i>E. segetalis</i>	59 38
426	(2 S*,3 S*,4 R*,5 R*,6 R*,8 R*,12 S*,13 R*,14 R*,15 R*)-6,14,17-triacetoxy-3-benzoyloxy-15-hydroxy-5-(2-hydroxyacetoxy)segetan-9-one	COCH ₂ OH	Ac	H	<i>E. paralias</i> <i>E. segetalis</i>	59 38
427	euphoportlandol B: 5α,11α,14α,17-tetraacetoxy-3β-benzoyloxy-6β,15β-dihydroxysegetan-9-one	Ac	H	OAc	<i>E. portlandica</i>	136
428	(2 S*,3 S*,4 R*,5 R*,6 R*,8 R*,11 S*,12 S*,13 R*,14 R*,15 R*)-5,6,11,14,17-pentaacetoxy-3-benzoyloxy-15-hydroxysegetan-9-one	Ac	Ac	OAc	<i>E. paralias</i>	59
429	paralinone B	COCH ₂ OAc	Ac	OAc	<i>E. paralias</i>	137
430	paralinone A	COCH ₂ OAc	Ac	H	<i>E. paralias</i>	137
431	segetanin A	Ac	Ac	OH	<i>E. paralias</i>	138

Table 54. Segetanes-2 and Presegetanes



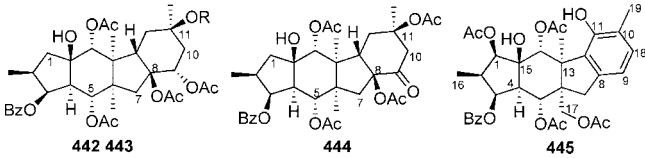
no.	name	plant	ref
432	euphoportlandol A: 5α,11α,14α,17-tetraacetoxy-3β-benzoyloxy-6β,15β-dihydroxyseget-8(12)-en-9-one	<i>E. portlandica</i>	136
433	segetanin B	<i>E. paralias</i>	138
434	(2 S*,3 S*,4 R*,5 R*,6 R*,8 R*,11 S*,12 S*,13 R*,14 R*,15 R*)-6,11,14-triacetoxy-3-benzoyloxy-15-hydroxy-5-(3-oxobutanoyl)-9-segetanone	<i>E. paralias</i>	59
435	(2 S*,3 S*,4 R*,5 R*,6 R*,7 S*,8 R*,12 R*,13 S*,14 R*,15 R*)-5-angeloyloxy-3-benzoyloxy-6,14:8,14-diepoxy-7,13,15,17-tetrahydroxy-15- <i>epi</i> -presegetan-9-one	<i>E. segetalis</i>	38
436	presegetanin	<i>E. paralias</i>	138

Table 55. Paralianes



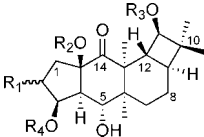
no.	name	R ₁	R ₂	R ₃	R ₄	plant	ref
437	(1 R*,2 R*,3 S*,4 R*,5 R*,6 R*,8 R*,12 S*,13 S*,14 R*,15 R*)-1,5,8,14-tetraacetoxy-3-benzoyloxy-15-hydroxyparalian-9-one	OAc	H	H	OAc	<i>E. segetalis</i>	38
438	(1 R*,2 R*,3 S*,4 R*,5 R*,6 R*,8 R*,12 R*,13 S*,14 R*,15 R*)-1,5,14-triacetoxy-3-benzoyloxy-15-hydroxyparalian-9-one	OAc	H	H	H	<i>E. paralias</i> <i>E. segetalis</i>	57 38
439	(2 R*,3 R*,4 S*,5 R*,6 R*,8 R*,12 R*,13 S*,14 R*,15 R*)-2,5,14-triacetoxy-3-benzoyloxy-15-hydroxyparalian-9-one	H	OAc	H	H	<i>E. segetalis</i>	38
440	(1 R*,2 R*,3 S*,4 R*,5 R*,6 R*,8 R*,12 R*,13 S*,14 R*,15 R*)-1,5,14,17-tetraacetoxy-3-benzoyloxy-15-hydroxyparalian-9-one	OAc	H	OAc	H	<i>E. segetalis</i>	38
441	(2 S*,3 S*,4 R*,5 R*,6 R*,8 S*,12 S*,13 S*,14 R*,15 R*)-5,8,14-triacetoxy-3-benzoyloxy-15-hydroxyparalian-9-one	H	H	H	OAc	<i>E. paralias</i>	59

Table 56. Pepluanes



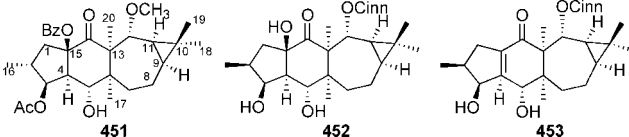
no.	name	R	plant	ref
442	(2 S*,3 S*,4 R*,5 R*,6 R*,8 R*,9 R*,10 R*,12 S*,13 S*,14 R*,15 R*)-5,8,9,11,14-pentaacetoxy-3-benzoyloxy-pepluan-15-ol	Ac	<i>E. peplus</i>	44
443	(2 S*,3 S*,4 R*,5 R*,6 R*,8 R*,9 R*,10 R*,12 S*,13 S*,14 R*,15 R*)-5,8,9,11,14-tetraacetoxy-3-benzoyloxy-pepluan-11,15 β -diol	H	<i>E. peplus</i>	19
444	pepluanone	—	<i>E. peplus</i>	40
445	(1 S*,2 R*,3 S*,4 R*,5 R*,6 R*,13 S*,14 R*,15 R*)-1,5,14,17-tetraacetoxy-3-benzoyloxy-8,10,(18)11-hexadehydropepluan-11,15-diol	—	<i>E. segetalis</i>	38

Table 57. Euphoractines-1



no.	name	R ₁	R ₂	R ₃	R ₄	plant	ref
446	3 β -acetoxy-11 β -methoxyeuphoractine D	α -Me	Bz	CH ₃	Ac	<i>E. villosa</i>	87
447	3 β -acetoxy euphoractine D	α -Me	Bz	H	Ac	<i>E. villosa</i>	87
448	euphoractine A	β -Me	Cinn	H	H	<i>E. micractina</i>	141
449	euphoractine C	β -Me	Bz	H	H	<i>E. micractina</i>	142
450	euphoractine D	α -Me	Bz	H	H	<i>E. micractina</i>	142

Table 58. Euphoractines-2



no.	name	plant	ref
451	no name	<i>E. villosa</i>	86
452	euphoractine B	<i>E. micractina</i>	141
453	euphoractine E	<i>E. micractina</i>	142

β -D-galactopyranoside (**544**), quercetin 3-O-(2''-galloyl)- β -D-galactopyranoside (**545**), quercetin, and gallic acid. Compounds **544** and **545** showed proliferation activity for BAF/InsR (insulin-dependent cell line), whereas quercetin and gallic acid showed stronger proliferation activity for BAF/IL10R than compounds **544** and **545**.¹⁷⁰

4.2. Cytotoxicity

Lathyrane diterpenoids **228**, **229**, and **234**, isolated from *E. nivulica*, showed significant cytotoxic activity against Colo 205, MT2, and CEM cell lines. The LD₅₀ values are almost the same in the three cell lines for the three compounds.⁸⁴ 17-Acetoxyjolkinoide B (**13**) and 13-hexadecanoyloxy-12-deoxyphorbol (**339**), obtained from the dried roots of *Euphorbia fischeriana*, exhibited potent cytotoxic activity to Ramos B cells with IC₅₀ values of 0.023 and 0.0051 μ g/mL, respectively.²¹ Daphnane diterpenoids **326**, **327**, and tiglane diterpenoids **342–345**, isolated from the bioactivity-guided fractionation of the latex of *E. poisonii*, showed strong cytotoxic selectivity for the human kidney carcinoma (A-498) cell line with potencies exceeding that of adriamycin by 10 000 times.¹²³ Compound **235**, also from *E. poisonii*,⁸⁵ modestly cytotoxic against A-549, MCF-7, HT, A-498, PC-3, and PACA-2, exhibited nonselective ED₅₀ values ranging from 2 to 4 μ g/mL, whereas **237** was weakly but selectively

active at ED₅₀ 15 μ g/mL against the prostate adenocarcinoma (PC-3).⁸⁵

4.3. Effects on the Cell Division

Ingenane-type diterpenoids **360–368**, **382**, **383**, **399–404**, and euphane triterpenoids **454–457**, all isolated from *E. kansui*,^{62,65,135,143} showed significant effects on the cell division of *Xenopus laevis* cells at the blastular stage. **360–368**, **382**, and **383** arrested cleavage significantly (0.5 μ g/mL of each compound resulted in >75% and 60% cleavage arrest, respectively). **402**, **403**, and **454–457** showed some activity (10 μ g/mL of each compound resulted in >60% and >50% cleavage arrest, respectively).

4.4. DNA-Damaging Activity

In a mutant yeast bioassay, compound **293**, isolated from *E. decipiens*, showed a positive response to DNA-damaging activity with IC₁₂ values of 750 μ g/mL against RS322Y (rad 52) and 1090 μ g/mL against wild-type LF 15 (Rad+).¹⁰⁵ Camptothecin was used as the standard drug, with IC₁₂ values of 12 μ g/mL for the mutant strain (rad 52) and 75 μ g/mL for the wild type.

4.5. Modularity of Multidrug Resistance

In a rhodamine 123 exclusion test using L5178 mouse lymphoma cells, euphosalicin (**216**) was found to be more active than verapamil in reversing multidrug resistance in mouse lymphoma cells.⁶³ The jatrophone diterpenoids **92**, **112**, and **151**, isolated from *E. mongolica*, also demonstrated a concentration-dependent effect in inhibiting the efflux pump activity of the tumor cells in the range 11.2–112 μ M.⁴⁹ Jatrophone diterpenoids **61**, **64**, and **119** (*E. peplus*), as well as two segetane diterpenoids, euphoportlandols B (**427**) and A (**432**) (*E. portlandica*), were found to be inhibitors of P-glycoprotein in the same test.^{66,136,115} The

Table 59. 6/6/6/5 Ring Triterpenoids

no.	name	R	plant	ref
454	kansenone	—	<i>E. kansui</i>	143
455	kansenonol	H ₂	<i>E. kansui</i>	143
456	11-oxo-kansenonol	O	<i>E. kansui</i>	143
457	kansenol		<i>E. kansui</i>	143
458	euphol		<i>E. antiquorum</i> <i>E. aleppica</i> <i>E. kansui</i>	144 114 143
459	antiquol C: eupa-7,9(11),24-trien-3β-ol		<i>E. antiquorum</i>	144
460	antiquol B: 19(10→9)abeo-8α(H),9β(H),10α(H)-eupa-5,24-dien-3β-ol		<i>E. antiquorum</i>	144
461	euphorbol: 24-methylenetirucall-8-en-3β-ol		<i>E. antiquorum</i>	144
462	<i>epi</i> -kansenone	O	<i>E. kansui</i>	143
463	lanosterol	H ₂	<i>E. peplus</i>	145
464	24-methylenelanosterol		<i>E. peplus</i>	145

Table 60. 6/6/6/6/5-Ring Triterpenoids-1

no.	name	R ₁	R ₂	plant	ref
465	lupeol acetate	Ac	Me	<i>E. altotibetic</i> <i>E. quinquecostata</i> <i>E. stygiana</i>	52 30 146
466	betulin	H	CH ₂ OH	<i>E. latifolia</i> <i>E. myrsinites</i>	147 104
467	betulinic acid	H	COOH	<i>E. latifolia</i>	147
468	lupeol	H	Me	<i>E. humifusa</i> <i>E. chamaesyce</i>	148 149

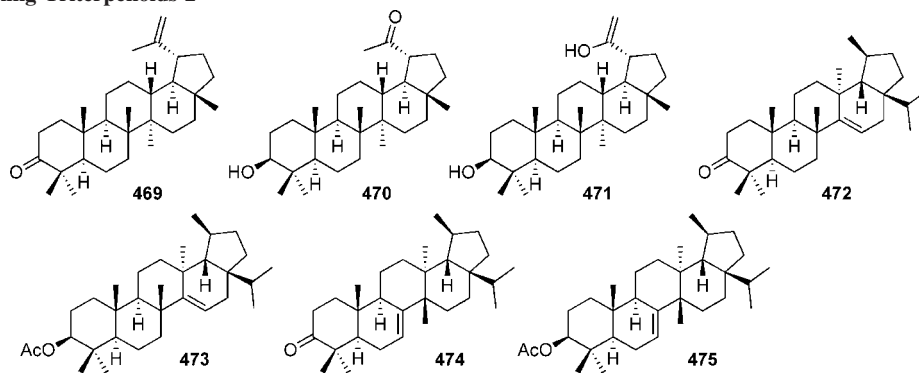
macrocyclic lathyrane polyester Euphorbia factor L₁₀ (**253**) showed powerful inhibition of the transport activity of P-glycoprotein in the experiment using one million K562/R7 human leukemic cells expressing high levels of P-glycoprotein.⁹²

Corea et al. studied on the structure–activity relationships of jatrophanes diterpenoids as modulators of multidrug resistance. In 2003, they isolated a series of 10 closely related jatrophanes **102** and euphodendroids A–I (**103–111**) from the Mediterranean spurge *E. dendroides* L., which served as a base for the establishment of structure–activity relationships within this class of P-glycoprotein inhibitors.⁵¹ The efficiency of euphodendroids to inhibit P-glycoprotein-

mediated daunomycin efflux was investigated by monitoring the intracellular accumulation of this drug. The results demonstrated the critical role of a free hydroxy at C-3, lacking of oxygenation at C-2 and the differential effect of substituent at C-5. Hydroxylation or acyloxylation at this C-2 was detrimental for the activity. The nicotinoyl derivative euphodendroidin C (**105**) was 2-fold less active compared to its 2-methylbutanoyl and isobutyryl analogs. The most powerful compound of the series, euphodendroidin D (**106**), outperformed cyclosporin by a factor of 2 to inhibit Pgp-mediated daunomycin transport. They also got 12 compounds, terracinolides B, C, H, J, K, L, 13α-OH terracinolides B, F, G, *abeo*dendroidin F, and *epi-abeo*dendroidin F (**193**, **194**, **199–205**, **213**, and **214**) from *E. dendroides* L.⁷⁴ All of the compounds were tested as inhibitors of the drug-efflux activity of P-glycoprotein from cancer cells. The main information was that the revertant activity of terracinolides and *abeo*jatrophanes was strongly affected by the presence of a free hydroxy group, with the following ranking of position: 3 < 15 < 13 < 2. In addition, substitution at position 6 affected the inhibitory ability in a way that dramatically depends on the location of the free hydroxy group. These observations suggest that jatrophanes and modified jatrophanes share a common gross pharmacophore, which is dramatically affected by changes of the oxygenation pattern.

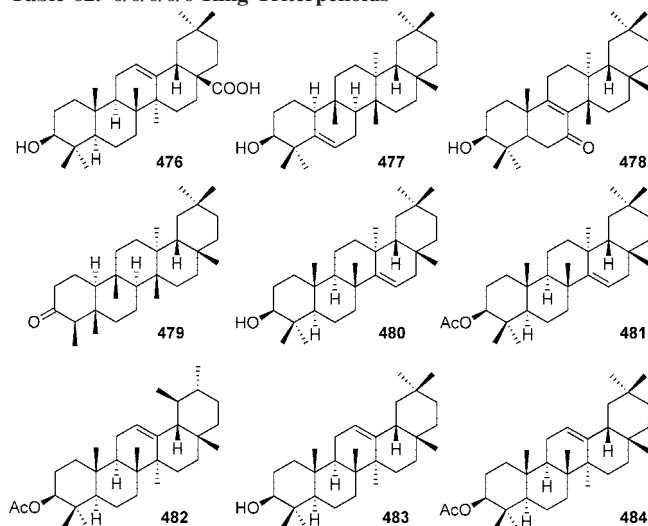
In 2004, Corea et al. went on researching the same structure–activity relationships.⁴⁵ The main object was to reveal the importance of substitutions on the medium-sized

Table 61. 6/6/6/5-Ring Triterpenoids-2



No.	name	plant	ref
469	lupenone	<i>E. segetalis</i>	150
		<i>E. stygiana</i>	146
470	3β-hydroxy-30-nor-lupan-20-one	<i>E. chamaesyce</i>	149
471	lup-20(30)-ene-3β,29-diol	<i>E. chamaesyce</i>	151
472	D-friedomadeir-14-en-3-one	<i>E. mellifera</i>	152
		<i>E. stygiana</i>	146
473	D-friedomadeir-14-en-3β-yl acetate	<i>E. stygiana</i>	146
		<i>E. mellifera</i>	152
474	D:C-friedomadeir-7-en-3-one	<i>E. stygiana</i>	146
		<i>E. stygiana</i>	146
475	D:C-friedomadeir-7-en-3β-yl acetate	<i>E. stygiana</i>	146

Table 62. 6/6/6/6-Ring Triterpenoids



no.	name	plant	ref
476	oleanolic acid	<i>E. latifolia</i>	147
477	glutinol	<i>E. chamaesyce</i>	153
		<i>E. segetalis</i>	150
		<i>E. chamaesyce</i>	153
478	3β-hydroxymultiflor-8-en-7-one	<i>E. chamaesyce</i>	153
479	friedeline	<i>E. segetalis</i>	130
480	taraxerone	<i>E. pubescens</i>	26
		<i>E. stygiana</i>	146
		<i>E. stygiana</i>	146
481	taraxeryl acetate	<i>E. stygiana</i>	146
482	α-amyirin acetate	<i>E. ebracteolata</i>	20
483	β-amyirin	<i>E. ebracteolata</i>	154
484	β-amyirin acetate	<i>E. ebracteolata</i>	154
		<i>E. fischeriana</i>	155

ring (carbons 8, 9, 14, and 15). Through the insight of seven jatrophone diterpenoids, pepluanins A–E (**66–68**, **86**, and **87**), **11**, and **65**, isolated from *E. peplus* L., they found that the activity was collapsed by the presence of C-8 hydroxy, whereas it increased with C-14 carbonyl, C-9 acetoxy, and C-15 hydroxy groups. Pepluanin A (**66**) showed a very high activity for a jatrophone diterpenoid, outperforming cyclosporin A by a factor of at least 2 in the inhibition of Pgp-mediated daunomycin transport.

4.6. Tumor Promoting Activity

Diterpene esters of the phorbol and ingenol types are known to be highly active tumor promoting agents that typically occur in members of the Euphorbiaceae.¹²⁸ Latex as well as total leaf extracts of *E. leuconera* exhibited Epstein–Barr-virus (EBV) inducing activity comparable to TPA (12-*O*-tetradecanoylphorbol acetate), a well-known tumor promoter. The activity of individual fractions correlated with their ingenol ester content.¹²⁸ Compounds **391–397**, obtained from the latex of *E. cauducifolia*, were evaluated for cocarcinogenic and tumor-promoting activity on the back skin of NMRI mice. After 24 weeks, an average tumor rate of 7% and an average tumor yield of 0.07 tumors/mouse were noticed. After 36 weeks, an average tumor rate of 36% was observed and the average tumor yield was 0.45 tumors/mouse.¹³⁰

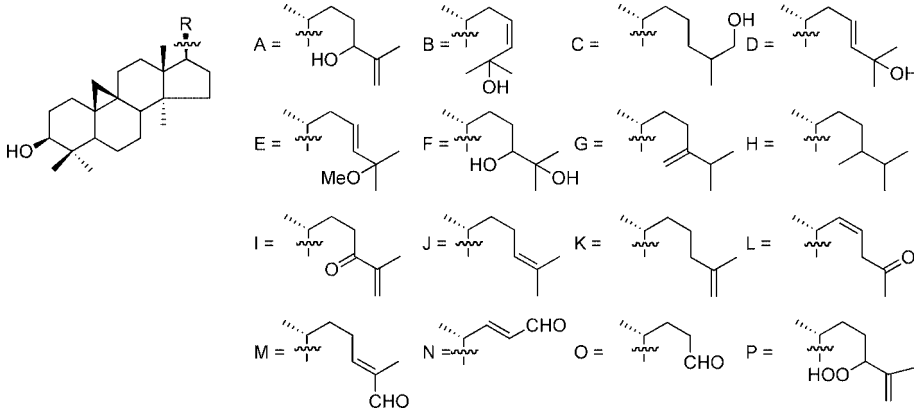
4.7. Pro-Inflammatory Activity

In 1999, Hohmann et al. investigated the irritant activities of some jatrophone diterpenes isolated from *E. peplus*. Only compound **77** was found to exert a weak pro-inflammatory activity on mouse ear of $ID_{50}^4 = ID_{50}^{24} = 29 \mu\text{g}/\text{ear}$ (the redness of the mouse ear was estimated 4 and 24 h after the application of solutions in Me_2CO). These data indicated that this type of diterpene does not play a significant role in the skin irritant activity of *Euphorbia* species.⁵⁵

4.8. Inhibition of Allergic Reactions

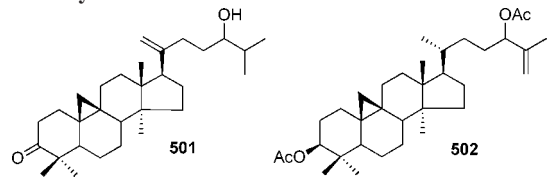
The water soluble fraction of *E. royleana* latex, showed dose-dependent anti-inflammatory and antiarthritic effects in different acute and chronic test models in rats and mice. It reduced the exudate volume and the migration of leukocytes and showed poor inhibitory effect on the granuloma formation induced by cotton pellets, while it had a low ulcerogenic score. The oral LD_{50} was more than 1500 mg/kg in both of rats and mice.¹⁷⁴ Oral administration of petroleum ether extract of the aerial parts of *E. splendens* caused significant inhibition of edema and produced inhibition of leukocyte migration and exudate volume in the affected tissues. The

Table 63. Cycloartanes-1



no.	name	R	plant	ref
485	cycloart-25-ene-3 β ,24-diol	A	<i>E. sessiliflora</i> <i>E. myrsinites</i> <i>E. portlandica</i> <i>E. aleppica</i> <i>E. altotibetic</i> <i>E. humifusa</i>	15 104 136 113 52 148
486	cycloart-23Z-ene-3 β ,25-diol	B	<i>E. sessiliflora</i> <i>E. portlandica</i> <i>E. humifusa</i> <i>E. chamaesyce</i>	15 136 148 153
487	cycloartane-3 β ,26-diol	C	<i>E. portlandica</i>	136
488	cycloart-23E-ene-3 β ,25-diol	D	<i>E. humifusa</i> <i>E. myrsinites</i> <i>E. altotibetic</i>	148 104 52
489	25-methoxycycloart-23E-en-3 β -ol	E	<i>E. sessiliflora</i>	15
490	cycloartane-3 β ,24,25-triol	F	<i>E. sessiliflora</i> <i>E. portlandica</i>	15 136
491	24-methylenecycloartan-3 β -ol	G	<i>E. myrsinites</i> <i>E. aleppica</i> <i>E. portlandica</i> <i>E. segetalis</i> <i>E. peplus</i> <i>E. pubescens</i> <i>E. ebracteolata</i>	104 113 136 150 148 26 154
492	cyclolaudanol	H	<i>E. myrsinites</i> <i>E. aleppica</i>	104 113
493	3 β -hydroxy-cycloart-25-en-24-one	I	<i>E. myrsinites</i> <i>E. portlandica</i> <i>E. aleppica</i>	104 136 113
494	cycloartenol	J	<i>E. peplus</i> <i>E. neriifolia</i> <i>E. segetalis</i>	145 156 150
495	cycloart-25-en-3 β -ol	K	<i>E. nivulia</i>	80
496	27-nor-3 β -hydroxycycloartan-25-one	L	<i>E. portlandica</i>	136
497	(24E)-3 β -hydroxycycloart-24-en-26-al	M	<i>E. portlandica</i>	136
498	(22E)-25,26,27-trinor-3 β -hydroxycycloart-22-en-24-al	N	<i>E. portlandica</i>	136
499	25,26,27-trinor-3 β -hydroxycycloartan-24-al	O	<i>E. portlandica</i>	136
500	24-hydroperoxycycloart-25-en-3 β -ol	P	<i>E. portlandica</i>	136

Table 64. Cycloartanes-2



no.	name	plant	ref
501	neriifolione	<i>E. neriifolia</i>	156
502	3 β ,24-diacetoxycycloart-25-ene	<i>E. portlandica</i>	136

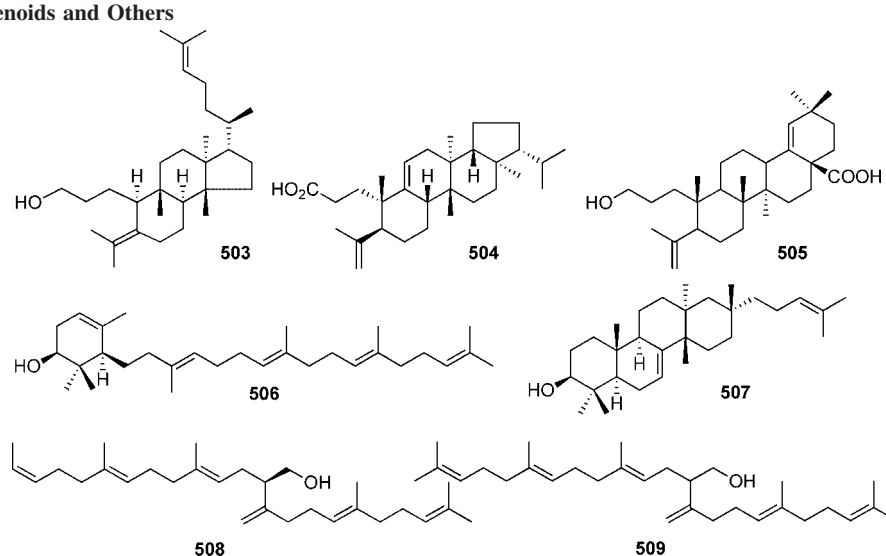
oral LD₅₀ in both rats and mice was approximately 1250 mg/kg.¹⁷⁵ A 95% ethanol extract from whole aerial parts of *E. hirta* (EH A001) showed antihistaminic, anti-inflammatory and immunosuppressive properties in various animal mod-

els.¹⁷⁶ *In vivo* tests, pepluanone (**444**), isolated from *E. peplus*, significantly reduced carrageenin-induced edema by 40% and 60%.¹⁴⁰ The lathyrane diterpenoid **229** showed significant PGE₂ inhibition using *in vitro* assay method employing Enzyme Immunoassay Kits. The IC₅₀ value for compound **229** was found to be 0.003 μ M compared to that of known PGE₂ inhibitor celecoxib (0.050 μ M).⁸⁰

4.9. Antimicrobial Activity

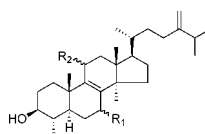
Ent-11 β -hydroxyabieta-8(14),13(15)-dien-16,12 α -olide (**16**) showed moderate to strong growth inhibition against *Bacillus cereus*, *B. subtilis*, *Micrococcus flavas*, *Moraxella catarrhalis*, *Neisseria sicca*, and *Candida albicans* CBS 5763 at 12.5 μ g/mL concentration. Jolkinolide A (**9**) also moderately inhibited

Table 65. Secotriterpenoids and Others



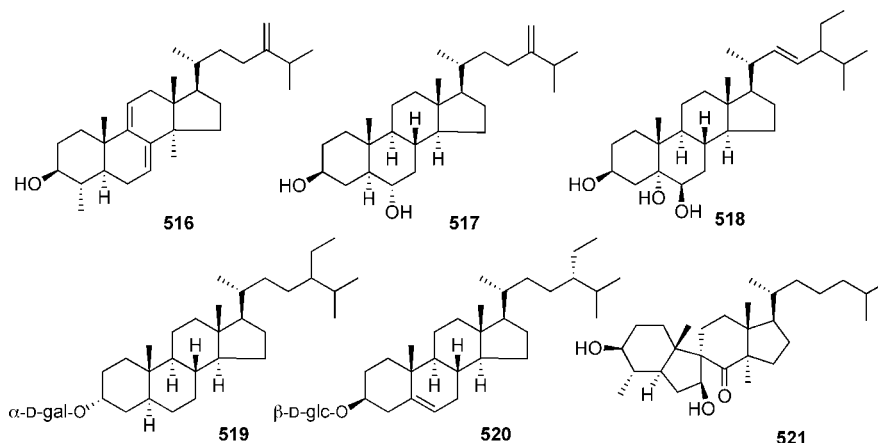
no.	name	plant	ref
503	isohelianol	<i>E. antiquorum</i>	144
504	3,4- <i>seco</i> -8β(H)-farna-4(23),9(11)-dien-3-oic acid	<i>E. chamaesyce</i>	153
505	3,4- <i>seco</i> -oleana-4(23),18-dien-3-oic acid	<i>E. chamaesyce</i>	157
506	camelliol C	<i>E. antiquorum</i>	144
507	lemmaphylla-7,21-dien-3β-ol	<i>E. antiquorum</i>	144
508	peplusol	<i>E. peplus</i>	145
509	anhydrobisfarnesol	<i>E. lateriflora</i>	158

Table 66. Steroids-1



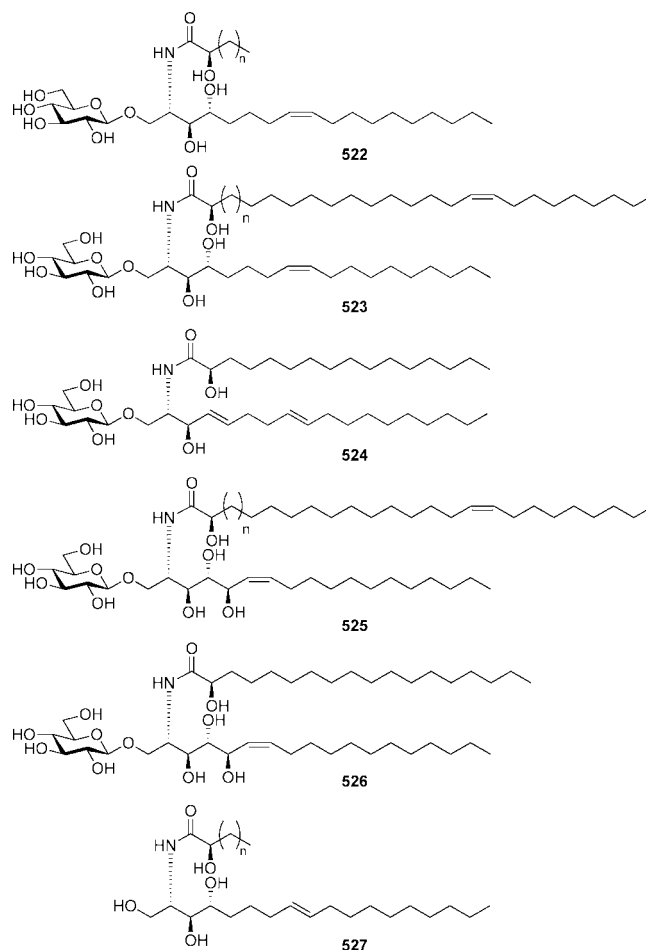
no.	name	R ₁	R ₂	plant	ref
510	3β-hydroxy-4α,14α-dimethyl-5α-ergosta-8,24(28)-dien-11-one	H ₂	=O	<i>E. chamaesyce</i>	151
511	3β,11α-dihydroxy-4α,14α-dimethyl-5α-ergosta-8,24(28)-dien-7-one	=O	α-OH	<i>E. chamaesyce</i>	151
512	3β,7α-dihydroxy-4α,14α-dimethyl-5α-ergosta-8,24(28)-dien-11-one	α-OH	=O	<i>E. chamaesyce</i>	151
513	3β-hydroxy-4α,14α-dimethyl-5α-ergosta-8,24(28)-dien-7-one	=O	H ₂	<i>E. chamaesyce</i>	149
514	3β-hydroxy-4α,14α-dimethyl-5α-ergosta-8,24(28)-dien-7,11-one	=O	=O	<i>E. chamaesyce</i>	149
515	obtusifoliol	H ₂	H ₂	<i>E. chamaesyce</i>	149

Table 67. Steroids-2



no.	name	plant	ref
516	4α,14α-dimethyl-5α-ergosta-7,9(11),24(28)-trien-3β-ol	<i>E. chamaesyce</i>	149
517	5α-stigmastane-3β,6α-diol	<i>E. boetica</i>	111
518	5α-stigmastane-3β,5,6β-triol	<i>E. boetica</i>	111
519	geniculatoside F	<i>E. geniculata</i>	159
520	3-(β-D-glucopyranosyloxy)stigmast-5-ene	<i>E. peplis</i>	160
521	(3 <i>S</i> ,4 <i>S</i> ,5 <i>S</i> ,7 <i>R</i> ,9 <i>R</i> ,14 <i>R</i>)-3,7-dihydroxy-4,14-dimethyl-7(8→9) <i>abeocholestan</i> -8-one	<i>E. officinarum</i>	86

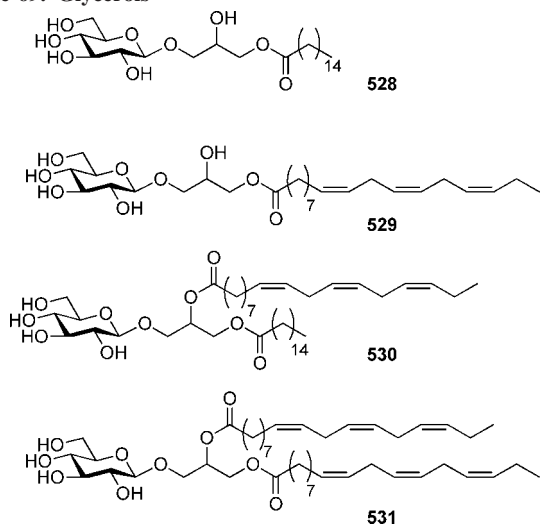
Table 68. Cerebrosides.



no.	name	<i>n</i>	plant	ref
522	(2 <i>S</i> ,3 <i>S</i> ,4 <i>R</i> ,8 <i>Z</i>)-1- <i>O</i> -(β- <i>D</i> -glucopyranosyl)-2-[(2' <i>R</i>)-2'-hydroxy(icosanoyl~octacosanoyl)amino]-8-octadecene-1,3,4-triol	17	<i>E. nicaeensis</i>	161
		19,20	<i>E. sororia</i>	162
		21	<i>E. peplis</i>	163
			<i>E. nicaeensis</i>	161
			<i>E. sororia</i>	162
523	(2 <i>S</i> ,3 <i>S</i> ,4 <i>R</i> ,8 <i>Z</i>)-1- <i>O</i> -(β- <i>D</i> -glucopyranosyl)-2-[(2' <i>R</i> ,15' <i>Z</i>)-2'-hydroxy(tetracos-15'-enoyl~octacos-19'-enoyl)amino]-8-octadecene-1,3,4-triol	22~25	<i>E. sororia</i>	162
		1	<i>E. peplis</i>	163
			<i>E. characias</i>	164
			<i>E. wulfenii</i>	165
		3	<i>E. peplis</i>	163
524	(2 <i>S</i> ,3 <i>S</i> ,4 <i>E</i> ,8 <i>E</i>)-1- <i>O</i> -(β- <i>D</i> -glucopyranosyl)-2-[(2' <i>R</i>)-2'-hydroxyhexadecanoylamino]-4,8-octadecadiene-1,3-diol	5	<i>E. wulfenii</i>	165
		—	<i>E. peplis</i>	163
		—	<i>E. characias</i>	166
525	(2 <i>S</i> ,3 <i>S</i> ,4 <i>R</i> ,5 <i>R</i> ,6 <i>Z</i>)-1- <i>O</i> -(β- <i>D</i> -glucopyranosyl)-2-[(2' <i>R</i> ,15' <i>Z</i>)-2'-hydroxy(tetracos-15'-enoyl~octacos-19'-enoyl)amino]-6-octadecene-1,3,4,5-tetraol	1	<i>E. characias</i>	166
		3	<i>E. characias</i>	166
		5	<i>E. characias</i>	166
526	(2 <i>S</i> ,3 <i>S</i> ,4 <i>R</i> ,5 <i>R</i> ,6 <i>Z</i>)-1- <i>O</i> -(β- <i>D</i> -glucopyranosyl)-2-[(2' <i>R</i>)-2'-hydroxyoctacosanoylamino]-6-octadecene-1,3,4,5-tetraol	—	<i>E. characias</i>	166
		—	<i>E. characias</i>	166
527	(2 <i>S</i> ,3 <i>S</i> ,4 <i>R</i> ,8 <i>E</i> ,2' <i>R</i>)-2-(2'-hydroxyeicosanoyl~hexacosanoylamino)-8-octadecene-1,3,4-triol	17~23	<i>E. sororia</i>	162

the growth of *M. catarrhalis* at 50 µg/mL concentration.¹⁵ Helioscopinolide A (**20**) and helioscopinolide B (**21**) showed significant activity against *Staphylococcus aureus* 6538P (2.5 µg/spot).²⁶ *In vitro* bioassays showed that yuexiandajisu A (**56**) exhibited antibacterial activity.⁴² A mixture of cerebrosides (**522**–**524**) from *E. peplis*¹⁶³ showed a synergistic antifungal activity against *Candida* spp. and *Cryptococcus neoformans* strain. Moreover, only a single compound **523**

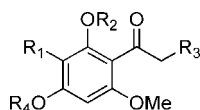
(*n* = 3) showed an interesting antitubercular activity with MIC of 40 µg/mL on reference strain and on two clinical isolates and a MIC of 80 µg/mL against clinical strain H242. Natarajan et al. researched the antibacterial activity of *E. fusiformis* against pathogenic strains of Gram positive (*Bacillus subtilis* and *S. aureus*) and Gram negative bacteria (*Escherichia coli*, *Klebsiella pneumoniae*, *Proteus vulgaris*, *Pseudomonas aeruginosa*, *Salmonella typhi* A, and *S. typhi*

Table 69. Glycerols

no.	name	plant	ref
528	1- <i>O</i> -(β -D-glucopyranosyl)-3-hexadecanoylglycerol	<i>E. cyparisias</i>	167
529	1- <i>O</i> -(β -D-glucopyranosyl)-3-(octadeca-9' Z,12' Z,15' Z-trienoyl)glycerol	<i>E. peplis</i>	168
530	1- <i>O</i> -(β -D-glucopyranosyl)-3-hexadecanoyl-2-(octadeca-9' Z,12' Z,15' Z-trienoyl)glycerol	<i>E. cyparisias</i>	167
531	1- <i>O</i> -(β -D-glucopyranosyl)-2,3-di(octadeca-9' Z,12' Z,15' Z-trienoyl)glycerol	<i>E. cyparisias</i>	167

B).¹⁷⁷ The different extracts differed significantly in their antibacterial properties with the methanolic extract being very effective followed by acetone and chloroform extracts. Aqueous and ethanolic extract showed the very least activity. Rootstock extracts had better antibacterial properties than leaf extracts. The results of this study supported the use of this plant in traditional medicine to treat fever, wound infections, and intestinal disorders.¹⁷⁷ The ethanolic extracts of aerial parts of *E. hirta* exhibited a broad spectrum of antimicrobial activity against *E. coli*, *P. vulgaris*, *P. aeruginosa*, and *S. aureus*.¹⁷⁸

Compound **438**, isolated from *E. paralias*, showed a moderate antiviral activity ($EC_{50} = 14$ mg/mL) against HIV-1 replication. The activity was based on the inhibition of virus-induced cytopathicity in MT-4 cells.⁵⁷ The 7 triterpenes (**458–461**, **503**, **506**, and **507**) isolated from *E. antiquorum* were examined for the inhibitory effects on Epstein–Barr virus early antigen (EBV-EA) activation induced by TPA.

Table 70. Phenolics-1

no.	name	R ₁	R ₂	R ₃	R ₄	plant	ref
532	2-hydroxy-4,6-dimethoxyacetophenone	H	H	H	Me	<i>E. portulacoides</i>	16
> 533	2,4,6-trimethoxyacetophenone	H	Me	H	Me	<i>E. portulacoides</i>	16
534	2-hydroxy-4,6-dimethoxy-3-methylacetophenone	Me	H	H	Me	<i>E. portulacoides</i>	16
535	2,4,6-trimethoxy-3-methylacetophenone	Me	Me	H	Me	<i>E. portulacoides</i>	16
536	2,2'-dihydroxy-4,6-dimethoxy-3-methylacetophenone	Me	H	OH	Me	<i>E. quinquecostata</i>	30
537	2,4-dihydroxy-6-methoxyacetophenone	H	H	H	H	<i>E. fischeriana</i>	13
538	2,4-dihydroxy-6-methoxy-3-methylacetophenone	Me	H	H	H	<i>E. ebracteolata</i>	168
539	2-hydroxy-6-methoxy-3-methylacetophenone 4- β -D-glucopyranoside	Me	H	H	glc	<i>E. ebracteolata</i>	168
540	ebractelatinoside C	Me	H	H	glc \rightarrow xyl(6 \rightarrow 1)	<i>E. ebracteolata</i>	168

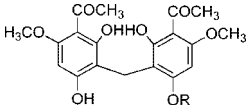
In this assay, compounds **458–461** and **503** showed 100% inhibition of activation at 1000 mol ratio/TPA.¹⁴⁴ The steroids **510–516**, isolated from *E. chamaesyce*, also exhibited potent inhibitory effects (100% inhibition of induction at 1000 mol ratio/TPA, about 80% inhibition at 500 mol ratio/TPA, and about 30% inhibition at 100 mol ratio/TPA) in the same assay. Compound **512** showed the most potent inhibitory effects in comparison to oleanolic acid on EBV-EA activation.¹⁵¹

4.10. Antidiarrheal Activity

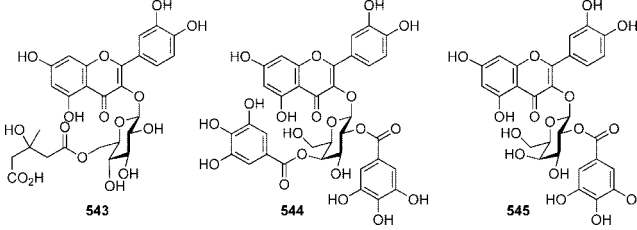
A significant antidiarrheal effect of the *E. paralias* extracts against castor oil-induced diarrhea in rats was achieved by 400 mg/kg.¹⁷⁹ It decreased the gastrointestinal movement as indicated by the significantly ($p < 0.01$) decreased distance traveled by the charcoal meal. The large dose of the extract was slightly more effective than the small one. The antidiarrheal effect was also evaluated on the motility of duodenum isolated from freshly slaughtered rabbits. The *E. paralias* methanol extract produced a transient stimulation followed by inhibition in doses of less than 0.05 mg/kg. Higher concentrations caused rapid muscle relaxation. Tannins, flavonoids, unsaturated sterols/triterpenoids, carbohydrates, lactones and proteins/amino acids were reported as major active constituents of the tested plants.¹⁷⁹ In 2006, Hore et al. reported the aqueous leaf extract of *E. hirta* decreased the gastrointestinal motility in normal rats and decreased the effect of castor oil-induced diarrhea in mice by 300 and 1000 mg/kg.¹⁸⁰

4.11. Antipyretic-Analgesic Activity

Myrsinane **266**, isolated from *E. decipens* whole plant chloroform extract, showed significant analgesic activity when administered to mice at dose of 5–20 mg/kg i.p. This activity is comparable to that of 100 mg/kg of aspirin or ibuprofen.¹⁸¹ Resiniferatoxin (**326**), an ultrapotent capsaicin analog present in the latex of *E. resinifera*, interacts at a specific membrane recognition site, expressed by primary sensory neurons mediating pain perception as well as neurogenic inflammation. Desensitization to **326** is a promising approach to mitigate neuropathic pain and other pathological conditions in which sensory neuropeptides released from capsaicin-sensitive neurons play a crucial role.¹⁸² Prostratin (**340**), obtained in *E. fischeriana*, showed significant analgesic and sedative activities. The 92% and 62% inhibitions were observed in sedative experiments with 20 mg/kg (p.o.) and 1 mg/kg (s.c.) in mice, respectively.⁸ The ethyl acetate

Table 71. Phenolics-2


no.	name	R	plant	ref
541	1,1-bis(2,6-dihydroxy-3-acetyl-4-methoxyphenyl)methane	H	<i>E. ebracteolata</i>	168
542	ebractelatinoside B	glc	<i>E. ebracteolata</i>	168

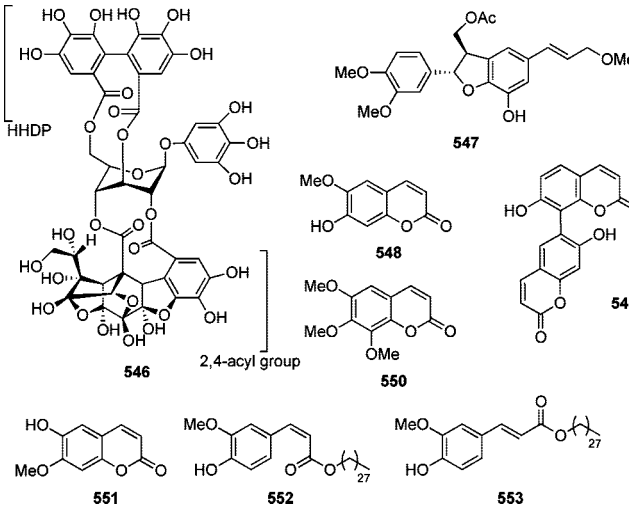
Table 72. Flavonoids


no.	name	plant	ref
543	quercetin 3-O-6'-(3-hydroxy-3-methylglutaryl)- β -D-glucopyranoside	<i>E. ebracteolata</i>	169
544	quercetin 3-O-(2'',3''-digalloyl)- β -D-galactopyranoside	<i>E. lunulata</i>	170
545	quercetin 3-O-(2''-galloyl)- β -D-galactopyranoside	<i>E. lunulata</i>	170

fraction from the residue of an 85% ethanol extract of the latex of *E. royleana* showed a dose related peripheral analgesic effect. The same fraction exhibited a significant antipyretic effect in hyperthermic rats and rabbits. The oral LD₅₀ was more than 2 g/kg in rats and mice.¹⁸³ Following an identified use of the plant as analgesic in traditional medicine, the hexane, chloroform and ethyl acetate extracts of *E. heterophylla* root were tested for antinociceptive activity in rats. All extracts showed significant effects at doses of 150/300 mg/kg.¹⁸⁴

4.12. Molluscicidal and Antifeedant Activities

In 2001 and 2002, Abdelgaleil et al. reported the molluscicidal and antifeedant activities of diterpenoids from *E. paralias*.^{57,185} Compounds **132**, **437**, and particularly **441** had high molluscicidal activities on *Biomphalaria alexandrina*. Antifeedant activity was tested by a conventional leaf disk method against third-instar larvae of *Spodoptera littoralis*.¹⁸⁵ Compounds **142** and **426** were good insect antifeedants with 66.8 and 45.8% antifeedant activity (1000 mg/mL), respectively. Kansanonol (**455**) and **143** were moderately active at 500 ppm, whereas compounds **132**, **144**, **424**, **428**, **437**, and **438** showed moderate activity at 500–1000 ppm.⁵⁷ In 2004, Singh et al. reported the aqueous and serially purified latex extracts of plants *E. pulcherima* and *E. hirta* had potent molluscicidal activity.¹⁸⁶ Sublethal doses (40 and 80% of LC₅₀) of aqueous and partially purified latex extracts of both the plants also significantly altered the levels of total protein, total free amino acid, nucleic acid (DNA and RNA), and the activity of enzyme protease and acid and alkaline phosphatase in various tissue of the snail *Lymnaea acuminata* in time and dose dependent manner.

Table 73. Miscellaneous Compounds


no.	name	plant	ref
546	jolkinin	<i>E. jolkinii</i>	171
547	(-)- <i>trans</i> -9-acetyl-4,9'-di-O-methyl-3'-de-O-methyldehydrodiconiferyl alcohol	<i>E. quinquecostata</i>	36
548	scopoletin	<i>E. lunulata</i>	52
549	7,7'-dihydroxy-6,8'-bicycoumarin (bicycoumarol)	<i>E. quinquecostata</i>	36
550	6,7,8-trimethoxycoumarin	<i>E. quinquecostata</i>	30
551	6-hydroxy-7-methoxycoumarin (isoscopoletin)	<i>E. quinquecostata</i>	36
552	octacosyl <i>cis</i> -ferulate	<i>E. hylonoma</i>	172
553	octacosyl <i>trans</i> -ferulate	<i>E. hylonoma</i>	172

4.13. Inhibitory Activity on the Mammalian Mitochondrial Respiratory Chain

Six diterpenoids (**330–334** and **336**) isolated from the latex of *E. obtusifolia* were evaluated for their inhibition of the NADH oxidase activity in submitochondrial particles from beef heart.¹⁸⁷ 12,13-Diisobutyryloxy-4,20-dideoxyphorbol (**330**) was the most potent inhibitor and showed an inhibitory concentration with IC₅₀ value of 2.06 ± 0.03 μ M. In the study, some structure–activity trends were suggested for the inhibitory activity of the mammalian mitochondrial respiratory chain of the 4-deoxyphorbol esters, in which two isobutyryloxy groups were located at C-12 and C-13. Compound **330**, the strongest inhibitor of the NADH oxidase activity, had an unfunctionalized *gem*-dimethylcyclopropane moiety and C-20 was not oxygenated. Less active compounds, such as **333**, **334**, and **335**, C-17 (OAc), and C-20 (OH), were oxygenated.

4.14. Antidipsogenic Activity

The effect of the methanol extract obtained from the leaves and stems of *E. hirta* on thirst was examined using Wistar rats.¹⁸⁸ Intraperitoneal administration of 10 mg/100 mg body wt of the extract significantly ($p < 0.05$) decreased the amount of water consumed by rats. This effect lasted for 2 h.

4.15. Survival Effect on Fibroblasts PGE₂ Inhibition Activity

Kansuinin E (**156**), isolated from the roots of *E. kansui*, exhibited a specific survival effect on fibroblasts that expressed TrkA with an ED₅₀ value of 0.23 μ g/mL when

compared with TrkB cells.⁶⁴ In contrast, kansuinins A (**184**), D (**186**), and F (**160**) enhanced the survival of both TrkA- and TrkB-expressing fibroblasts. TrkA is a high-affinity receptor for nerve growth factor. The survival of these cells was solely dependent on NGF treatment, and they normally died in the absence of NGF. The survival of TrkB-expressing cells was dependent on the presence of brain-derived neurotrophic factor (BDNF), a member of the neurotrophin family related to NGF.¹⁸⁹

4.16. PEP Inhibitory Activity

Prolyl endopeptidase (PEP) is the only serine protease that is known to cleave a peptide substrate in the C-terminal side of a proline residue and plays an important role in the metabolism of peptide hormones and neuropeptides and is recognized to be involved in learning and memory.¹⁹⁰ Low molecular weight inhibitors of PEP have been reported in the literature, but the majority of these were synthetic.¹⁹¹ Most of the natural PEP inhibitors isolated have been of microbial origin whereas PEP inhibitors from plants have rarely been investigated.¹⁹² The myrsinol-type diterpenoids **276**, **291**, and **292**, isolated from *E. decipiens*, were active against PEP.^{101,105} Compounds **276** and **291** showed IC₅₀ of 3.2 and 10.5 μM , respectively, with the positive control of Bacitracin. Compound **292** exhibited an IC₅₀ of $16.9 \pm 1.3 \mu\text{M}$, which was compared with the positive control of PEP (Z-Pro-prolinal, IC₅₀ of $1.27 \pm 0.01 \mu\text{M}$).

4.17. Urease Inhibitory Activity

Studies on the enzyme inhibition have led to the discoveries of drugs. Urease inhibitors have recently attracted much attention as potential new antiulcer drugs. Unfortunately, only a few natural products with this activity have been discovered. In 2003, Ahmad et al. reported the isolation of decipinol ester A (**273**) from *E. decipiens*.¹⁰¹ It was the first naturally occurring urease inhibitor.

4.18. Angiotensin Converting Enzyme Inhibiting Activity

The methanol extract obtained from the leaves and stems of *E. hirta* inhibited the activity of angiotensin converting enzyme (ACE) by 90 and 50% at 500 and 160 μg , respectively, using enzyme linked immunosorbent assay (ELISA).¹⁸⁸

4.19. Other Activities

3,4-Dimethoxycinnamaldehyde, isolated from *E. quinquecostata*, was significantly active in the induction of quinone reductase (QR) in Hepa1c1c7 hepatoma cells and in the inhibition of the transformation of murine epidermal JB6 cells with a CD (concentration to double induction) value of 9.5 $\mu\text{g}/\text{mL}$ (52.8 μM) (QR assay) and an IC₅₀ value of 2.3 $\mu\text{g}/\text{mL}$ (12.8 μM) (JB6 assay), respectively.³⁶ The water and ethanol extracts (50 and 100 mg/kg) of *E. hirta* produced time-dependent increase in urine output. Electrolyte excretion was also significantly affected by the plant extracts.¹⁹³ The water extract increased the urine excretion of Na^+ , K^+ , and HCO_3^- . In contrast, the ethanol extract increased the excretion of HCO_3^- , decreased the loss of K^+ , and had little effect on renal removal of Na^+ .

5. Conclusion

The genus *Euphorbia* is widespread all over the world. The diterpenoids with jatrophone, lathyrane, tigliane, ingenane, *ent*-abietane, and myrsinol skeletons are among the most studied diterpenoids isolated from *Euphorbia* plants. Other types of diterpenoids, such as segetane, paraliene, pepluane, euphoractine, *ent*-atisnae, *ent*-kaurane, and casbane, along with triterpenes, sesquiterpenoids, steroids, and flavonoids are also important components. The rare compounds are cerebrosides, ellagitannin, neolignan, and manoyloxide, which isolated from *E. characias*,¹⁶³ *E. jolkinii*,¹⁷¹ *E. quinquecostata*,³⁶ and *E. segetalis*,³⁸ respectively. Also, several enzymes have been isolated from the Euphorbiaceae family. The present review shows the majority of the compounds isolated from *Euphorbia*. But the structures of very common compounds such as organic acids and aliphatic hydrocarbons are not given.

The biological research on *Euphorbia* species has supported the use of some plants in traditional medicines or revealed the new activities on modern pharmacological levels. The biological activity includes antiproliferative activity, modulability of multidrug resistance, cytotoxic activity, DNA-damaging activity, antiviral activity, PEP inhibitory activity, antidiarrheal activity, molluscicidal and antifeedant activities, antimicrobial activity, antiinflammatory activity, antipyretic-analgesic activity, inhibitory activity on the mammalian mitochondrial respiratory chain, PGE₂ inhibition activity, angiotensin converting enzyme inhibiting activity, antidipsogenic activity, survival effect on fibroblasts, inhibitory activity on urease, effects on the cell division, tumor promoting activity, etc.

The insight of structure–activity relationships study on jatrophone diterpenoids as modulators of multidrug resistance has given us more detailed information about the active core framework and substituents.^{45,51,74} It could help fellow researchers to find more active P-glycoprotein inhibitors.

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7. References

- (1) Jassbi, A. R. *Phytochemistry* **2006**, *67*, 1977.
- (2) Singla, A. K.; Pathak, K. *Fitoterapia* **1990**, *61*, 483.
- (3) Jury, S. L.; Reynolds, T.; Cutler, D. F.; Evans, J. F., Eds. *The Euphorbiales: Chemistry, Taxonomy, and Economic Botany*; Academic Press: London, 1987.
- (4) Ponsinet, G.; Ourisson, G. *Phytochemistry* **1968**, *7*, 89.
- (5) Nielsen, P. E.; Nishimura, H.; Liang, Y.; Calvin, M. *Phytochemistry* **1979**, *18*, 103.
- (6) Evans, J. F.; Taylor, S. E. *Prog. Chem. Org. Nat. Prod.* **1983**, *44*, 1.
- (7) Marco, J. A.; Sanz-Cervera, J. F.; Yuste, A. *Phytochemistry* **1997**, *45*, 563.
- (8) Ma, Q. G.; Liu, W. Z.; Wu, X. Y.; Zhou, T. X.; Qin, G. W. *Phytochemistry* **1997**, *44*, 663.
- (9) Cateni, F.; Falsone, G.; Zilic, J. *Mini-Rev. Med. Chem.* **2003**, *3*, 425.
- (10) Shi, J. G.; Shi, Y. P.; Ji, Z. J. *Phytochemistry* **1997**, *45*, 343.
- (11) Fattorusso, E.; Lanzotti, V.; Tagliatalata-Scafati, O.; Tron, G. C.; Appendino, G. *Eur. J. Org. Chem.* **2002**, *71*.
- (12) Zhang, W.; Guo, Y. W. *Chem. Pharm. Bull.* **2006**, *54*, 1037.
- (13) Che, C. T.; Zhou, T. X.; Ma, Q. G.; Qin, G. W.; Williams, I. D.; Wu, H. M.; Shi, Z. S. *Phytochemistry* **1999**, *52*, 117.

- (14) Liu, G. F.; Fu, Y. Q.; Yang, Z. Q.; Zhao, H. Q.; Fan, X. M. *Zhongyao Tongbao* **1988**, *5*, 291.
- (15) Suththivaiyakit, S.; Thapsut, M.; Prachayasittikul, V. *Phytochemistry* **2000**, *53*, 947.
- (16) Morgenstern, T.; Bittner, M.; Silva, M.; Aqueveque, P.; Jakupovic, J. *Phytochemistry* **1996**, *41*, 1149.
- (17) Satti, N. K.; Suri, O. P.; Dhar, K. L.; Atal, C. K.; Kawasaki, T.; Miyahara, K.; Kawano, S. *Phytochemistry* **1986**, *25*, 1411.
- (18) Ahmad, S.; Seligmann, O.; Wagner, H.; Hussain, G. *Phytochemistry* **1977**, *16*, 1844.
- (19) Jeske, F.; Jakupovic, J.; Berendsohn, W. *Phytochemistry* **1995**, *40*, 1743.
- (20) Wang, W. X.; Ding, X. B. *Acta Pharm. Sin.* **1998**, *33*, 128.
- (21) Wang, Y. B.; Huang, R.; Wang, H. B.; Jin, H. Z.; Lou, L. G.; Qin, G. W. *J. Nat. Prod.* **2006**, *69*, 967.
- (22) Appendino, G.; Belloro, E.; Tron, G. C.; Jakupovic, J.; Ballero, M. *Fitoterapia* **2000**, *71*, 134.
- (23) Appendino, G.; Jakupovic, J.; Tron, G. C.; Jakupovic, J.; Milon, V.; Ballero, M. *J. Nat. Prod.* **1998**, *61*, 749.
- (24) Lal, A. R.; Cambie, R. C.; Rutledge, P. S.; Woodgate, P. D. *Phytochemistry* **1990**, *29*, 2239.
- (25) Shizuri, Y.; Kosemura, S.; Yamamura, S.; Ohba, S.; Ito, M.; Saito, Y. *Chem. Lett.* **1983**, 65.
- (26) Valente, C.; Pedro, M.; Duarte, A.; Nascimento, M. S. J.; Abreu, P. M.; Ferreira, M. J. U. *J. Nat. Prod.* **2004**, *67*, 902.
- (27) Borghi, D.; Ballabio, M.; Arlandini, E.; Crespi Perellino, N.; Minghetti, A.; Francesco Vincieri, F. *J. Nat. Prod.* **1991**, *54*, 1503.
- (28) Marco, J. A.; Sanz-Cervera, J. F.; Yuste, A.; Jakupovic, J. *J. Nat. Prod.* **1999**, *62*, 110.
- (29) Lal, A. R.; Cambie, R. C.; Rutledge, P. S.; Woodgate, P. D. *Phytochemistry* **1990**, *29*, 1925.
- (30) Mbwambo, Z. H.; Lee, S. K.; Mshiu, E. N.; Pezzuto, J. M.; Kinghorn, A. D. *J. Nat. Prod.* **1996**, *59*, 1051.
- (31) Satti, N. K.; Suri, O. P.; Thaper, R. K.; Kachroo, P. L. *Phytochemistry* **1988**, *27*, 1530.
- (32) Lal, A. R.; Cambie, R. C.; Rutledge, P. S.; Woodgate, P. D.; Rickard, C. E. F.; Clark, G. R. *Tetrahedron Lett.* **1989**, *30*, 3205.
- (33) Pan, L.; Zhou, P.; Zhang, X. F.; Peng, S.; Ding, L. S.; Qiu, S. X. *Org. Lett.* **2006**, *8*, 2775.
- (34) Ding, Y. L.; Jia, Z. J. *Phytochemistry* **1991**, *30*, 2412.
- (35) Wang, H.; Zhang, X. F.; Luo, X. D. *Nat. Prod. Res. Dev.* **2006**, *18*, 53.
- (36) Su, B. N.; Park, E. J.; Mbwambo, Z. H.; Santarsiero, B. D.; Mesecar, A. D.; Fong, H. H. S.; Pezzuto, J. M.; Kinghorn, A. D. *J. Nat. Prod.* **2002**, *65*, 1278.
- (37) Satti, N. K.; Suri, O. P.; Dhar, K. L.; Kawasaki, T.; Miyahara, K.; Noda, N. *J. Nat. Prod.* **1987**, *50*, 790.
- (38) Jakupovic, J.; Jeskt, F.; Morgenstern, T.; Tschritzis, F.; Marco, J. A.; Berendsohn, W. *Phytochemistry* **1998**, *47*, 1583.
- (39) Wang, Y. B.; Yao, G. M.; Wang, H. B.; Qin, G. W. *Chem. Lett.* **2005**, *34*, 1160.
- (40) Zhou, T. X.; Bao, G. H.; Ma, Q. G.; Qin, G. W.; Che, C. T.; Lv, Y.; Wang, C.; Zheng, Q. T. *Tetrahedron Lett.* **2003**, *44*, 135.
- (41) Pei, Y. H.; Koike, K.; Han, B.; Jia, Z. H.; Nikaïdo, T. *Tetrahedron Lett.* **1999**, *40*, 951.
- (42) Xu, Z. H.; Sun, J.; Xu, R. S.; Qin, G. W. *Phytochemistry* **1998**, *49*, 149.
- (43) Marco, J. A.; Sanz-Cervera, J. F.; Yuste, A.; Jakupovic, J.; Jeske, F. *Phytochemistry* **1998**, *47*, 1621.
- (44) Jakupovic, J.; Morgenstern, T.; Bittner, M.; Silva, M. *Phytochemistry* **1998**, *47*, 1601.
- (45) Corea, G.; Fattorusso, E.; Lanzotti, V.; Motti, R.; Simon, P. N.; Dumontet, C.; Di Pietro, A. *J. Med. Chem.* **2004**, *47*, 988.
- (46) Appendino, G.; Spagliardi, P.; Ballero, M.; Seu, G. *Fitoterapia* **2002**, *73*, 576.
- (47) Miglietta, A.; Gabriel, L.; Appendino, G.; Bocca, C. *Cancer Chemother. Pharmacol.* **2003**, *51*, 67.
- (48) Liu, L. G.; Tan, R. X. *J. Nat. Prod.* **2001**, *64*, 1064.
- (49) Hohmann, J.; Rédei, D.; Forgo, P.; Molnár, J.; Dombi, G.; Zorig, T. *J. Nat. Prod.* **2003**, *66*, 976.
- (50) Marco, J. A.; Sanz-Cervera, J. F.; Checa, J.; Palomares, E.; Fraga, B. M. *Phytochemistry* **1999**, *52*, 479.
- (51) Corea, G.; Fattorusso, E.; Lanzotti, V.; Tagliatalata-Scafati, O.; Appendino, G.; Ballero, M.; Simon, P. N.; Dumontet, C.; Di Pietro, A. *J. Med. Chem.* **2003**, *46*, 3395.
- (52) Pan, L.; Zhang, X. F.; Deng, Y.; Wang, H.; Wu, D. G.; Luo, X. D. *Helv. Chim. Acta* **2003**, *86*, 2525.
- (53) Günther, G.; Hohmann, J.; Vasas, A.; Máthé, I.; Dombi, Gy.; Jerkovich, Gy. *Phytochemistry* **1998**, *47*, 1309.
- (54) Günther, G.; Martinek, T.; Dombi, G.; Hohmann, J.; Vasas, A. *Magn. Reson. Chem.* **1999**, *37*, 365.
- (55) Hohmann, J.; Vasas, A.; Günther, G.; Dombi, Gy.; Blazsó, G.; Falkay, Gy.; Máthé, I.; Jerkovich, Gy. *Phytochemistry* **1999**, *51*, 673.
- (56) Corea, G.; Fattorusso, C.; Fattorusso, E.; Lanzotti, V. *Tetrahedron* **2005**, *61*, 4485.
- (57) Abdelgaleil, S. A. M.; Kassem, S. M. I.; Doe, M.; Baba, M.; Nakatani, M. *Phytochemistry* **2001**, *58*, 1135.
- (58) Hohmann, J.; Vasas, A.; Günther, G.; Máthé, I.; Evanics, F.; Dombi, G.; Jerkovich, Gy. *J. Nat. Prod.* **1997**, *60*, 331.
- (59) Jakupovic, J.; Morgenstern, T.; Marco, J. A.; Berendsohn, W. *Phytochemistry* **1998**, *47*, 1611.
- (60) Ferreira, A. M. V. D.; Carvalho, L. H. M.; Carvalho, M. J. M.; Sequeira, M. M.; Silva, A. M. S. *Phytochemistry* **2002**, *61*, 373.
- (61) Valente, C.; Pedro, M.; Ascenso, J. R.; Abreu, P. M.; Nascimento, M. S. J.; Ferreira, M. J. U. *Planta Med.* **2004**, *70*, 244.
- (62) Wang, L. Y.; Wang, L. N.; Yao, X. S.; Miyata, S.; Kitanaka, S. *Chem. Pharm. Bull.* **2003**, *51*, 935.
- (63) Hohmann, J.; Evanics, F.; Dombi, G.; Molnár, J.; Szabó, P. *Tetrahedron* **2001**, *57*, 211.
- (64) Pan, Q.; Ip, F. C. F.; Ip, N. Y.; Zhu, H. X.; Min, Z. D. *J. Nat. Prod.* **2004**, *67*, 1548.
- (65) Wang, L. Y.; Wang, N. L.; Yao, X. S.; Miyata, S.; Kitanaka, S. *J. Nat. Prod.* **2002**, *65*, 1246.
- (66) Hohmann, J.; Molnár, J.; Rédei, D.; Evanics, F.; Forgo, P.; Kálmán, A.; Argay, G.; Szabó, P. *J. Med. Chem.* **2002**, *45*, 2425.
- (67) Hohmann, J.; Forgo, P.; Csupor, D.; Schlosser, G. *Helv. Chim. Acta* **2003**, *86*, 3386.
- (68) Lu, Z. Q.; Guan, S. H.; Li, X. N.; Chen, G. T.; Zhang, J. Q.; Huang, H. L.; Liu, X.; Guo, D. A. *J. Nat. Prod.* **2008**, *71*, 873.
- (69) Valente, C.; Ferreira, M. J. U.; Abreu, P. M.; Gyémánt, N.; Ugoicsai, K.; Hohmann, J.; Molnár, J. *Planta Med.* **2004**, *70*, 81.
- (70) Valente, C.; Ferreira, M. J. U.; Abreu, P. M.; Pedro, M.; Cerqueira, F.; Nascimento, M. S. J. *Planta Med.* **2003**, *69*, 361.
- (71) Rédei, D.; Hohmann, J.; Evanics, F.; Forgo, P.; Szabó, P.; Máthé, I. *Helv. Chim. Acta* **2003**, *86*, 280.
- (72) Hohmann, J.; Rédei, D.; Evanics, F.; Kálmán, A.; Argay, G.; Bartók, T. *Tetrahedron* **2000**, *56*, 3619.
- (73) Marco, J. A.; Sanz-Cervera, J. F.; Yuste, A.; Jakupovic, J.; Lex, J. *J. Org. Chem.* **1996**, *61*, 1707.
- (74) Corea, G.; Fattorusso, E.; Lanzotti, V.; Tagliatalata-Scafati, O.; Appendino, G.; Ballero, M.; Simon, P. N.; Dumontet, C.; Di Pietro, A. *Bioorg. Med. Chem.* **2003**, *11*, 5221.
- (75) Marco, J. A.; Sanz-Cervera, J. F.; Yuste, A.; Jakupovic, J. *Phytochemistry* **1997**, *45*, 137.
- (76) Hohmann, J.; Evanics, F.; Dombi, G.; Szabó, P. *Tetrahedron Lett.* **2001**, *42*, 6581.
- (77) Marco, J. A.; Sanz-Cervera, J. F.; Roperio, F. J.; Checa, J.; Fraga, B. M. *Phytochemistry* **1998**, *49*, 1095.
- (78) Ahmed, A. A.; Couladis, M.; Mahmoud, A. A.; Adams, A.; Mabry, T. J. *Fitoterapia* **1999**, *70*, 140.
- (79) Khan, A. Q.; Malik, A. *J. Nat. Prod.* **1990**, *53*, 728.
- (80) Ravikanth, V.; Reddy, V. L. N.; Rao, T. P.; Diwan, P. V.; Ramakrishna, S.; Venkateswarlu, Y. *Phytochemistry* **2002**, *59*, 331.
- (81) Connolly, D. J.; Fakunle, O. C.; Rycroft, S. D. *Tetrahedron Lett.* **1984**, *25*, 3773.
- (82) Lin, L. J.; Kinghorn, D. *Phytochemistry* **1983**, *22*, 2795.
- (83) Opferkuch, H. J.; Hecker, E. *Tetrahedron Lett.* **1973**, 3611.
- (84) Ravikanth, V.; Reddy, V. L. N.; Reddy, A. V.; Ravinder, K.; Rao, T. P.; Ram, T. S.; Kumar, K. A.; Vamanrao, D. P.; Venkateswarlu, Y. *Chem. Pharm. Bull.* **2003**, *51*, 431.
- (85) Fatope, M. O.; Zeng, L.; Ohayagha, J. E.; McLaughlin, J. L. *Bioorg. Med. Chem. Lett.* **1996**, *4*, 1679.
- (86) Daoubi, M.; Marquez, N.; Mazoir, N.; Benharref, A.; Hernández-Galán, R.; Muñoz, E.; Collado, I. G. *Bioorg. Med. Chem.* **2007**, *15*, 4577.
- (87) Vasas, A.; Hohmann, J.; Forgo, P.; Szabó, P. *Tetrahedron* **2004**, *60*, 5025.
- (88) Appendino, G.; Tron, G. C.; Cravotto, G.; Palmisano, G.; Jakupovic, J. *J. Nat. Prod.* **1999**, *62*, 76.
- (89) Hohmann, J.; Evanics, F.; Vasas, A.; Dombi, G.; Jerkovich, G.; Máthé, I. *J. Nat. Prod.* **1999**, *62*, 176.
- (90) Appendino, G.; Belloro, E.; Tron, G. C.; Jakupovic, J.; Ballero, M. *J. Nat. Prod.* **1999**, *62*, 1399.
- (91) Liao, S. G.; Zhan, Z. J.; Yang, S. P.; Yue, J. M. *Org. Lett.* **2005**, *7*, 1379.
- (92) Appendino, G.; Porta, C. D.; Conseil, G.; Sterner, O.; Mercalli, E.; Dumontet, C.; Pietro, A. D. *J. Nat. Prod.* **2003**, *66*, 140.
- (93) Adolf, W.; Köhler, I.; Hecker, E. *Phytochemistry* **1984**, *23*, 1461.
- (94) Wang, H.; Zhang, X.; Zhou, Y.; Ascenso, J. R.; Abreu, P. M.; Nascimento, M. S. J.; Ferreira, M. J. U. *Planta Med.* **2004**, *70*, 244.
- (95) Gao, S.; Liu, H. Y.; Wang, Y. H.; He, H. P.; Wang, J. S.; Di, Y. T.; Li, C. S.; Fang, X.; Hao, X. J. *Org. Lett.* **2007**, *9*, 3453.
- (96) Zahid, M.; Husani, S. R.; Abbas, M.; Pan, Y.; Jassbi, A. R.; Asim, M.; Parvez, M.; Voelter, W.; Ahmad, V. U. *Helv. Chim. Acta* **2001**, *84*, 1980.

- (97) Ahmad, V. U.; Hussain, H.; Jassbi, A. R.; Zahid, M.; Hussain, J.; Bukhari, I. A.; Yasin, A.; Choudhary, M. I. *Pol. J. Chem.* **2002**, *76*, 1699.
- (98) Ahmad, V. U.; Jassbi, A. R.; Parvez, M. *Tetrahedron* **1998**, *54*, 1573.
- (99) Jassbi, A. R.; Fukushima, Y.; Tahara, S. *Helv. Chim. Acta* **2002**, *85*, 1706.
- (100) Ahmad, V. U.; Hussain, H.; Hussain, J.; Bukhari, I. A.; Yasin, A.; Choudhary, M. I.; Dar, A. Z. *Naturforsch.* **2002**, *57C*, 1066.
- (101) Ahmad, V. U.; Hussain, J.; Hussain, H.; Jassbi, A. R.; Ullah, F.; Lodhi, M. A.; Yasin, A.; Choudhary, M. I. *Chem. Pharm. Bull.* **2003**, *51*, 719.
- (102) Ahmad, V. U.; Jassbi, A. R. *Planta Med.* **1998**, *64*, 732.
- (103) Zhang, W. J.; Chen, D. F.; Hou, A. J. *Chin. J. Chem.* **2004**, *22*, 103.
- (104) Öksüz, S.; Gürek, F.; Gil, R. R.; Pengsuparp, T.; Pezzuto, J. M.; Cordell, G. A. *Phytochemistry* **1995**, *38*, 1457.
- (105) Ahmad, V. U.; Hussain, H.; Jassbi, A. R.; Hussain, J.; Bukhari, I. A.; Yasin, A.; Aziz, N.; Choudhary, M. I. *J. Nat. Prod.* **2003**, *66*, 1221.
- (106) Abbas, M.; Jassbi, A. R.; Zahid, M.; Ali, Z.; Alam, N.; Akhtar, F.; Choudhary, M. I.; Ahamad, V. U. *Helv. Chim. Acta* **2000**, *83*, 2751.
- (107) Zhang, J.; Yang, C. J.; Wu, D. G. *Chin. Tradit. Herb. Drugs* **1998**, *29*, 73.
- (108) Ahmad, V. U.; Jassbi, A. R. *J. Nat. Prod.* **1999**, *62*, 1016.
- (109) Öksüz, S.; Gürek, F.; Qiu, S. X.; Cordell, G. A. *J. Nat. Prod.* **1998**, *61*, 1198.
- (110) Wu, D. G.; Sorg, B.; Hecher, E. *J. Nat. Prod.* **1995**, *58*, 408.
- (111) Ferreira, M. J. U.; Ascenso, J. R. *Phytochemistry* **1999**, *51*, 439.
- (112) Ahmad, V. U.; Jassbi, A. R. *Phytochemistry* **1998**, *48*, 1217.
- (113) Öksüz, S.; Gürek, F.; Lin, L. Z.; Gil, R. R.; Pezzuto, J. M.; Cordell, G. A. *Phytochemistry* **1996**, *42*, 473.
- (114) Shi, Y. P.; Jia, Z. J.; Jamil, L.; Sadiq, S. *Phytochemistry* **1995**, *40*, 1219.
- (115) Duarte, N.; Ferreira, M. J. U. *Org. Lett.* **2007**, *9*, 489.
- (116) Schmidt, R. J.; Evans, J. F. *Phytochemistry* **1976**, *15*, 333.
- (117) Schmidt, R. J.; Evans, J. F. *Phytochemistry* **1976**, *15*, 1778.
- (118) Hergenbahn, M.; Adolf, W.; Hecker, E. *Tetrahedron Lett.* **1975**, *1595*.
- (119) Adolf, W.; Sorg, B.; Hergenbahn, M.; Hecker, E. *J. Nat. Prod.* **1982**, *45*, 347.
- (120) Haba, H.; Lavaud, C.; Harkat, H.; Alabdul Magid, A.; Marcourt, L.; Benkhaled, M. *Phytochemistry* **2007**, *68*, 1255.
- (121) Miana, G. A.; Bashir, M.; Evans, F. J. *Planta Med.* **1985**, *51*, 353.
- (122) Schmidt, R. J.; Evans, J. F. *Lloydia* **1977**, *40*, 225.
- (123) Fatope, M. O.; Zeng, L.; Ohayaga, J. E.; Shi, G.; McLaughlin, J. L. *J. Med. Chem.* **1996**, *39*, 1005.
- (124) Baloch, I. B.; Baloch, M. K.; Saqib, Q. N. *Eur. J. Med. Chem.* **2008**, *43*, 274.
- (125) Li, Y. L.; Suo, Y. R. *Zhongcaoyao* **2005**, *36*, 1763.
- (126) Pan, D. J.; Hu, C. Q.; Chang, J. J. *Phytochemistry* **1991**, *30*, 1018.
- (127) Itokawa, H.; Ichihara, Y.; Watanabe, K.; Takeya, K. *Planta Med.* **1989**, *55*, 271.
- (128) Vogg, G.; Mattes, E.; Rothenburger, J.; Hertkorn, N.; Achatz, S.; Sandermann, H. *Phytochemistry* **1999**, *51*, 289.
- (129) Uemura, D.; Ohwaki, H.; Hirata, Y.; Chen, Y. P.; Hsu, H. Y. *Tetrahedron Lett.* **1974**, *29*, 2527.
- (130) Baloch, I. B.; Baloch, M. K.; Saqib, Q. N. *Helv. Chim. Acta* **2005**, *88*, 3145.
- (131) Lin, L. J.; Marshall, G. T.; Kinghorn, A. D. *J. Nat. Prod.* **1983**, *46*, 723.
- (132) Lin, L. J.; Kinghorn, A. D. *J. Agric. Food Chem.* **1983**, *31*, 396.
- (133) Lu, Z. Q.; Yang, M.; Zhang, J. Q.; Chen, G. T.; Huang, H. L.; Guan, S. H.; Ma, C.; Liu, X.; Guo, D. A. *Phytochemistry* **2008**, *69*, 812.
- (134) Ott, H. H.; Hecker, E. *Experientia* **1981**, *37*, 88.
- (135) Kamimura, D.; Ijichi, K.; Shigetani, S.; Baba, M.; Fujiwara, M.; Yokota, T.; Tokuhisa, K.; Watanabe, H. *Chem. Abstr.* **1996**, *126*, 65375; JP Patent 08245379, 1996.
- (136) Madureira, A. M.; Gyémánt, N.; Ascenso, J. R.; Abreu, P. M.; Molnár, J.; Ferreira, M. J. U. *J. Nat. Prod.* **2006**, *69*, 950.
- (137) Öksüz, S.; Gürek, F.; Yang, S.; Lin, L.; Cordell, G. A.; Pezzuto, J. M.; Wagner, H.; Lotter, H. *Tetrahedron* **1997**, *53*, 3215.
- (138) Barile, E.; Lanzotti, V. *Org. Lett.* **2007**, *9*, 3603.
- (139) Hohmann, J.; Günther, G.; Vasas, A.; Kálmán, A.; Argay, G. *J. Nat. Prod.* **1999**, *62*, 107.
- (140) Corea, G.; Fattorusso, E.; Lanzotti, V.; Meglío, P.; Maffia, P.; Grassia, G.; Ialenti, A.; Ianaro, A. *J. Med. Chem.* **2005**, *48*, 7055.
- (141) Shi, J. G.; Jia, Z. J.; Yang, L. *Phytochemistry* **1993**, *32*, 208.
- (142) Shi, J. G.; Jia, Z. J. *Phytochemistry* **1995**, *38*, 1445.
- (143) Wang, L. Y.; Wang, N. L.; Yao, X. S.; Miyata, S.; Kitanaka, S. *J. Nat. Prod.* **2003**, *66*, 630.
- (144) Akihisa, T.; Wijeratne, E. M. K.; Tokuda, H.; Enjo, F.; Toriumi, M.; Kimura, Y.; Koike, K.; Nikaido, T.; Tezuka, Y.; Nishino, H. *J. Nat. Prod.* **2002**, *65*, 158.
- (145) Berkowitz, J. D.; Giner, J.-L.; Andersson, T. *J. Nat. Prod.* **2000**, *63*, 267.
- (146) Lima, E. M. C.; Medeiros, J. M. R.; Davin, L. B. *Phytochemistry* **2003**, *63*, 421.
- (147) Zhang, L. H.; Zhang, G. T.; Ye, W. C.; Yin, Z. Q.; Zheng, D.; Zhao, S. X. *Nat. Prod. Res. Dev.* **2006**, *18*, 58.
- (148) Liu, R. H.; Kong, L. Y. *Nat. Prod. Res. Dev.* **2005**, *17*, 437.
- (149) Tanaka, R.; Kasubuchi, K.; Kita, S.; Matsunaga, S. *Phytochemistry* **1999**, *41*, 457.
- (150) Ferreira, M. J. U.; Margarida, A.; Ascenso, J. R. *Phytochemistry* **1998**, *49*, 179.
- (151) Tanaka, R.; Kasubuchi, K.; Kita, S.; Tokuda, H.; Nishino, H.; Matsunaga, S. *J. Nat. Prod.* **2000**, *63*, 99.
- (152) Ferreira, M. J. U. Doctoral Thesis, University of Lisbon, Lisbon, 1990.
- (153) Tanaka, R.; Ida, T.; Kita, S.; Kamisako, W.; Matsunaga, S. *Phytochemistry* **1996**, *41*, 1163.
- (154) Fu, G. M.; Yu, B. Y.; Zhu, D. N. *J. Chin. Pharm. Uni.* **2003**, *34*, 377.
- (155) Liu, W. C.; He, Z. Y.; Gu, X. F.; Wu, X. Y.; Qin, G. W. *J. Chin. Mater. Med.* **2001**, *26*, 180.
- (156) Ilyas, M.; Parveen, M.; Amin, K. M. Y. *Phytochemistry* **1998**, *48*, 561.
- (157) Tanaka, R.; Ida, T.; Takaoka, Y.; Kita, S.; Kamisako, W.; Matsunaga, S. *Phytochemistry* **1994**, *36*, 129.
- (158) Faure, S.; Connolly, J. D.; Fakunle, C. O.; Piva, O. *Tetrahedron* **2000**, *56*, 9647.
- (159) Rahman, A.; Ali, M.; Khan, N. Z. *Pharmazie* **2002**, *57*, 643.
- (160) Cateni, F.; Zilic, J.; Falsone, G.; Kralj, B.; Tubaro, A.; Sosa, S. *Pharm. Pharmacol. Lett.* **2001**, *11*, 49.
- (161) Cateni, F.; Zilic, J.; Falsone, G.; Hollan, F.; Frausin, F.; Scarcia, V. *Farmaco* **2003**, *58*, 809.
- (162) Zhang, W. K.; Xu, J. K.; Zhang, X. Q.; Yao, X. S.; Ye, W. C. *Chem. Phys. Lipids* **2007**, *148*, 77.
- (163) Cateni, F.; Zilic, J.; Falsone, G.; Scialino, G.; Banfi, E. *Bioorg. Med. Chem. Lett.* **2003**, *13*, 4345.
- (164) Falsone, G.; Cateni, F.; Baumgartner, M.; Lucchini, V.; Wagner, H.; Seligmann, O. Z. *Naturforsch.* **1994**, *49C*, 135.
- (165) Falsone, G.; Cateni, F.; Vrevez, I.; Wagner, H.; Haslinger, E.; Presser, A. *Pharm. Pharmacol. Lett.* **1998**, *8*, 16.
- (166) Falsone, G.; Cateni, F.; Katusian, F.; Wagner, H.; Seligmann, O.; Pellizer, G.; Asaro, F. Z. *Naturforsch.* **1993**, *48C*, 1121.
- (167) Cateni, F.; Falsone, G.; Vitrotti, E.; Zilic, J.; Birkofer, L.; Della Loggia, R.; Sosa, S. *Pharm. Pharmacol. Lett.* **2000**, *10*, 65.
- (168) Wang, W. X.; Ding, X. B. *Acta Pharm. Sin.* **1999**, *34*, 514.
- (169) Liu, X.; Ye, W. C.; Yu, B.; Zhao, S. X.; Wu, H. M.; Che, C. T. *Carbohydr. Res.* **2004**, *339*, 891.
- (170) Nishimura, H.; Wang, L. Y.; Kusano, K.; Kitanaka, S. *Chem. Pharm. Bull.* **2005**, *53*, 305.
- (171) Lee, S. H.; Tanaka, T.; Nonaka, G. I.; Nishioka, I. *J. Nat. Prod.* **2004**, *67*, 1018.
- (172) Ruan, H. L.; Zhou, X. F.; Zhang, Y. H.; Pi, H. F.; Wu, J. Z.; Sun, H. D. *Fitoterapia* **2007**, *78*, 72.
- (173) Kuo, P. L.; Cho, C. Y.; Hsu, Y. L.; Lin, T. C.; Lin, C. C. *Toxicol. Appl. Pharmacol.* **2006**, *213*, 37.
- (174) Bani, S.; Kaul, A.; Jaggi, B. S.; Suri, K. A.; Suri, O. P.; Sharma, O. P. *Fitoterapia* **2000**, *71*, 655.
- (175) Bani, S.; Singh, S.; Singh, G. B.; Banerjee, S. K. *Phytother. Res.* **1997**, *11*, 76.
- (176) Singh, G. D.; Kaiser, P.; Youssouf, M. S.; Singh, S.; Khajuria, A.; Koul, A.; Bani, S.; Kapahi, B. K.; Satti, N. K.; Suri, K. A.; Johri, R. K. *Phytother. Res.* **2006**, *20*, 316.
- (177) Natarajan, D.; Britto, S. J.; Srinivasan, K.; Nagamurugan, N.; Mohanasundari, C.; Perumal, G. *J. Ethnopharmacol.* **2005**, *102*, 123.
- (178) Sudhakar, M.; Rao Ch, V.; Rao, P. M.; Raju, D. B.; Venkateswarlu, Y. *Fitoterapia* **2006**, *77*, 378.
- (179) Atta, A. H.; Mouneir, S. M. *Phytother. Res.* **2005**, *19*, 481.
- (180) Hore, S. K.; Ahuja, V.; Mehta, G.; Kumar, P.; Pandey, S. K.; Ahmad, A. H. *Fitoterapia* **2006**, *77*, 35.
- (181) Ahmad, V. U.; Hussain, H.; Bukhari, I. A.; Hussain, J.; Jassbi, A. R.; Dar, A. *Fitoterapia* **2005**, *76*, 230.
- (182) Appendino, G.; Szallasi, A. *Life Sci.* **1997**, *60*, 681.
- (183) Bani, S.; Suri, K. A.; Suri, O. P.; Sharma, O. P. *Phytother. Res.* **1997**, *11*, 597.
- (184) Vamsidhar, I.; Habeeb Mohammed, A.; Nataraj, B.; Madhusudana Rao, C.; Ramesh, M. *Fitoterapia* **2000**, *71*, 562.
- (185) Abdelgaleil, S. A. M.; El-Aswad, A. F.; Nakatani, M. *Pest. Manag. Sci.* **2002**, *58*, 479.
- (186) Singh, S. K.; Yadav, R. P.; Singh, D.; Singh, A. *Enviro. Toxicol. Pharm.* **2004**, *15*, 87.

- (187) Betancur-Galvis, L.; Palomares, E.; Marco, J. A.; Estornell, E. *J. Ethnopharmacol.* **2003**, *85*, 279.
- (188) Williams, L. A. D.; Gossell-Williams, M.; Sajabi, A.; Barton, E. N.; Fleischhacker, R. *Phytother. Res.* **1997**, *11*, 401.
- (189) Ip, N. Y.; Yancopoulos, G. D. *Annu. Rev. Neurosci.* **1996**, *19*, 491.
- (190) Anis, I.; Ahmed, S.; Malik, A.; Yasin, A.; Choudhary, M. I. *Chem. Pharm. Bull.* **2002**, *50*, 515.
- (191) Wallen, E. A. A.; Christiaans, J. A. M.; Forsberg, M. F.; Venäläinen, P. T. M.; Gynther, J. *J. Med. Chem.* **2002**, *45*, 4581.
- (192) Lee, K. H.; Kwak, J. H.; Lee, K. B.; Song, K. S. *Arch. Pharm. Res.* **1998**, *21*, 207.
- (193) Johnson, P. B.; Abdurahman, E. M.; Tiam, E. A.; Abdu-Aguye, I.; Hussaini, I. M. *J. Ethnopharmacol.* **1999**, *65*, 63.

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